

Original research

Whole-genome sequencing of monozygotic twins discordant for schizophrenia indicates multiple genetic risk factors for schizophrenia



Jinsong Tang ^{a,1}, Yu Fan ^{b,1}, Hong Li ^{a,j,1}, Qun Xiang ^{b,c}, Deng-Feng Zhang ^b,
Zongchang Li ^a, Ying He ^a, Yanhui Liao ^a, Ya Wang ^d, Fan He ^e, Fengyu Zhang ^a,
Yin Yao Shugart ^f, Chunyu Liu ^g, Yanqing Tang ^{h,*}, Raymond C.K. Chan ^{d,*},
Chuan-Yue Wang ^{e,*}, Yong-Gang Yao ^{b,c,i,*}, Xiaogang Chen ^{a,*}

^a Institute of Mental Health, National Clinical Research Center for Mental Health Disorders and National Technology Institute of Psychiatry, and Key Laboratory of Psychiatry and Mental Health of Hunan Province, The Second Xiangya Hospital, Central South University, Changsha 410011, China

^b Key Laboratory of Animal Models and Human Disease Mechanisms of the Chinese Academy of Sciences & Yunnan Province, Kunming Institute of Zoology, Kunming 650223, China

^c Kunming College of Life Science, University of Chinese Academy of Sciences, Kunming 650204, China

^d Neuropsychology and Applied Cognitive Neuroscience Laboratory, and CAS Key Laboratory of Mental Health, Institute of Psychology, Chinese Academy of Sciences, Beijing 100101, China

^e Beijing Key Laboratory of Mental Disorders, Department of Psychiatry, Beijing Anding Hospital, and Center of Schizophrenia, Beijing Institute for Brain Disorders and Laboratory of Brain Disorders of the Ministry of Science and Technology, Capital Medical University, Beijing 100088, China

^f Unit on Statistical Genomics, Intramural Research Programs, National Institute of Mental Health, NIH, Bethesda 20892, USA

^g Institute of Human Genetics, University of Illinois at Chicago, Chicago, IL 60607, USA

^h Department of Psychiatry, The First Affiliated Hospital of China Medical University, Shenyang 110122, China

ⁱ CAS Center for Excellence in Brain Science and Intelligence Technology, Chinese Academy of Sciences, Shanghai 200031, China

^j Department of Psychiatry, The First Affiliated Hospital of Zhengzhou University, Zhengzhou 450052, China

ARTICLE INFO

Article history:

Received 1 March 2017

Received in revised form

29 April 2017

Accepted 9 May 2017

Available online 8 June 2017

Keywords:

Whole-genome sequencing

Schizophrenia

Monozygotic twin

De novo mutation

Combined effect

Susceptibility

ABSTRACT

Schizophrenia is a common disorder with a high heritability, but its genetic architecture is still elusive. We implemented whole-genome sequencing (WGS) analysis of 8 families with monozygotic (MZ) twin pairs discordant for schizophrenia to assess potential association of *de novo* mutations (DNMs) or inherited variants with susceptibility to schizophrenia. Eight non-synonymous DNMs (including one splicing site) were identified and shared by twins, which were either located in previously reported schizophrenia risk genes (p.V24689I mutation in TTN, p.S2506T mutation in GCN1L1, IVS3+1G > T in DOCK1) or had a benign to damaging effect according to *in silico* prediction analysis. By searching the inherited rare damaging or loss-of-function (LOF) variants and common susceptible alleles from three classes of schizophrenia candidate genes, we were able to distill genetic alterations in several schizophrenia risk genes, including GAD1, PLXNA2, RELN and FEZ1. Four inherited copy number variations (CNVs; including a large deletion at 16p13.11) implicated for schizophrenia were identified in four families, respectively. Most of families carried both missense DNMs and inherited risk variants, which might suggest that DNMs, inherited rare damaging variants and common risk alleles together conferred to schizophrenia susceptibility. Our results support that schizophrenia is caused by a combination of multiple genetic factors, with each DNM/variant showing a relatively small effect size.

Copyright © 2017, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, and Genetics Society of China. Published by Elsevier Limited and Science Press. All rights reserved.

* Corresponding authors.

E-mail addresses: yanqingtang@163.com (Y. Tang), rckchan@psych.ac.cn (R.C.K. Chan), wang.cy@163.net (C.-Y. Wang), yaoyg@mail.kiz.ac.cn (Y.-G. Yao), chenxghn@163.com (X. Chen).

¹ These authors contributed equally to this work.

1. Introduction

Schizophrenia is a severe chronic mental disorder affecting about 0.5% of the world populations (Saha et al., 2005). While it is believed that the etiology of schizophrenia is multifactorial, genetic epidemiological studies have indicated that schizophrenia is highly heritable (Sullivan et al., 2003). Genome-wide association studies (GWAS) have identified a number of common single nucleotide polymorphisms (SNPs) and copy number variations (CNVs) associated with schizophrenia (Schizophrenia Psychiatric Genome-Wide Association Study Consortium, 2011; Shi et al., 2011; Yue et al., 2011; Sullivan et al., 2012; Ma et al., 2013; Ripke et al., 2013; Wu et al., 2017). Although some CNVs were replicated and have not been found in mentally healthy cohort, they are not unique to schizophrenia (Stefansson et al., 2014). A recent large GWAS of schizophrenia reported 108 independent genetic variants that met the level of genome-wide significance (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014). However, these common variants all together only explain a small portion of genetic risk. The “missing heritability” is still a preeminent case in schizophrenia (McClellan and King, 2010; Lee et al., 2012; Zuk et al., 2014).

Several studies had analyzed rare genetic variants or *de novo* mutations (DNMs) in schizophrenia through using either whole-genome sequencing (WGS) or whole-exome sequencing (WES) technology (Need et al., 2012; Fromer et al., 2014; Wang et al., 2015). While no specific genes associated with schizophrenia were identified, these studies demonstrated that patients with schizophrenia tend to have an enrichment of rare risk variants or DNMs in genes involved in synaptic function and in those encoding the activity-regulated cytoskeleton (ARC)-associated proteins and N-methyl-D-aspartate receptor (NMDAR) complexes, as well as fragile X mental retardation protein (FMRP) (Fromer et al., 2014). Mutations in the nongenic (Bae et al., 2014), noncoding RNA (Kwon et al., 2013), and large CNV regions (Liu et al., 2002; Xu et al., 2008; CNV and Schizophrenia Working Groups of the Psychiatric Genomics Consortium, 2017) may play important roles in the development of neuropsychiatric disorders such as schizophrenia. WGS is powerful for systematically evaluating various genetic risk factors such as common and rare variants, DNMs, and CNVs that may contribute to neuropsychiatric disorders (Jiang et al., 2013; Yuen et al., 2015). In this study, we performed WGS of 8 monozygotic (MZ) twin pairs discordant for schizophrenia and their parents to assess the potential role of DNMs or inherited genetic variants in liability of schizophrenia.

2. Results

2.1. Alignment and coverage statistics

WGS was performed at the BGI using the Illumina HiSeq platform with a depth of 30× (Table S1). Sequence calling was made for each individual using the Genome Analysis Toolkit (GATK) (McKenna et al., 2010). We detected an average of 3,141,201 single nucleotide variants (SNVs) (min = 3,096,496; max = 3,182,102; SD = 21,670) and 701,678 indels (min = 678,058; max = 720,000; SD = 11,501) per individual genome. Greater than 95% of these variations were SNPs in dbSNP142 (Bhagwat, 2010). Each sequenced individual had an average of 18,006 SNVs and 335 indels that were located in the coding region (Fig. S1A). Members of the 8 families (32 individuals) shared 735,818 SNVs and 169,308 indels, with no apparent family-specific pattern of SNVs and indels (Table S2; Fig. S1B and C). According to the minor allele frequency (MAF) in the 1000 Genomes Project (Sudmant et al., 2015), we detected around 6 million common SNVs (MAF > 5%), 94,344 low

frequency SNVs (MAF = 0.5%–5%), and 73,453 rare SNVs (MAF < 0.5%) on average in each of the 32 individuals (Fig. S1D).

2.2. Detection and validation of DNMs

We applied forestDNM (Michaelson et al., 2012) to detect DNMs in trios (mother, father, and one offspring), combined the results of GATK (McKenna et al., 2010) and pindel (Ye et al., 2009) to find *de novo* indels, and employed four algorithms (Breakdancer (Chen et al., 2009), CNVnator (Abzyzov et al., 2011), Delly (Rausch et al., 2012) and Lumpy (Layer et al., 2014)) to detect the *de novo* CNVs. Mutations shared by the MZ twins but not in their parents are counted as germline mutations. Mutations that were not shared by MZ twins are referred to as somatic mutations (Koren et al., 2012). While no *de novo* CNVs were detected, a total of 502 putative *de novo* SNVs (60.2 per individual) and 12 putative *de novo* indels (1.5 per individual) were detected in 8 MZ twin pairs (Fig. 1; Tables S3 and S4). There might be plausible mutation clusters across the genome, although our observation was based on a limited number of individuals (Fig. 1). We randomly selected 37 DNMs for validation on mother, father, and offspring using Sanger sequencing, of which 3 failed to be amplified, 20.6% (7/34) were false-positive variant calls, and 8.8% (3/34) were true-positive-inherited SNPs falsely called as negative in one parent (Table S5). The overall validation rate was comparable or higher than those of previous studies (Michaelson et al., 2012; Jiang et al., 2013; Genome of the Netherlands Consortium, 2014). We did not validate the 12 putative *de novo* indels, simply because they are all located in non-coding region, and the overall frequency of *de novo* indels in our sample was consistent with the previous study (Jiang et al., 2013).

2.3. DNMs and father's age at conception of child

We estimated the genome-wide mutation rate based on the germline DNMs per offspring (37–81 DNMs/offspring) as $1.18\text{--}2.58 \times 10^{-8}$ per generation, which is lower than the expected rate (Haldane, 2004). To gain more statistical power, we combined our data with others reported previously (Michaelson et al., 2012) to examine the association of the number of DNMs with parents' age at conception of child. We performed the correlation analyses of the number of DNMs in the offspring with maternal and paternal age at conception, respectively. Significant associations were observed with both maternal ($p\text{-value} = 2.60 \times 10^{-3}$) and paternal age ($p\text{-value} = 5.08 \times 10^{-5}$). Because the parent ages were highly correlated ($R^2 = 0.83$), we carried out a conditional analysis of multiple regression models to separate the age effect of the father and/or mother. We observed a significant positive correlation ($p\text{-value} = 0.009$) between father's age at conception of child and the number of DNMs in the offspring, whereas mother's age at conception ($p\text{-value} = 0.976$) was not significantly correlated with the number of DNMs (Fig. 2).

2.4. DNMs and potential pathogenicity

Nearly a half of DNMs were located in the intergenic region, only 8 DNMs (7 missense and 1 synonymous DNMs) shared between twins were located in the exon (Tables S4 and S6). In 8 probands, 4 (50%) were found to carry at least one *de novo* missense mutation. The *de novo* indels were located in the non-coding region. *In silico* functional analysis using both PolyPhen-2 (Adzhubei et al., 2010, 2013) and SIFT (Ng, 2003; Kumar et al., 2009) predicted that 4 of the 7 missense mutations would be deleterious (Table 1).

Approximately 63.3% (314/504 = 61.5%; excluding 10 unvalidated DNMs out of 37 randomly selected DNMs for validation) DNMs had been annotated with known and predicted regulatory

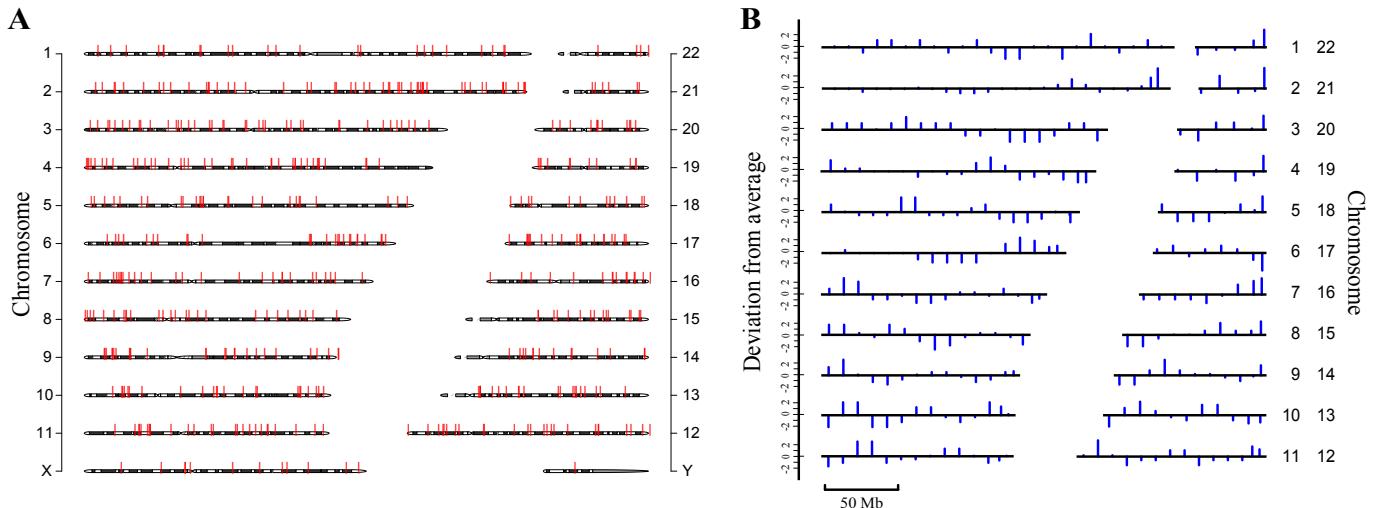


Fig. 1. The distribution of DNMs in the genomes of the 8 schizophrenia families. **A:** The location of DNMs in chromosomes 1–22, X and Y. The DNMs are marked with red “|”. **B:** The distribution difference of DNMs located on different regions. The distribution difference is represented by the deviation from the average which was calculated by subtracting the average number of DNMs in 40 Mb genomic region from the average number of DNMs in whole chromosome, with a sliding window of 20 Mb genomic region for counting.

elements by RegulomeDB (Boyle et al., 2012), including DNAase hypersensitivity sites, binding sites of transcription factors, and promoter regions (Table S7). Approximately two-thirds of these DNMs (334/504 = 66.3%) are transitions. DNMs are located in CpG sites, including 4 in the CpG islands. Because all these DNMs are rare mutations, we could not retrieve the related information, such as expression quantitative trait loci (eQTL) (Yang et al., 2010). We arbitrarily classified these non-coding DNMs into different categories according to the location of each mutation in predicted regulatory elements of known schizophrenia susceptibility genes (3 classes of gene sets (see Methods and materials)), genes that were not reported to be associated with schizophrenia, and the intergenic regions. There was no apparent enriched pattern for more non-coding DNMs in regulatory elements in known schizophrenia genes (p -value > 0.01 for all three comparisons, fisher's exact test).

Most of the identified DNMs (92.1%; 464/504; excluding 10 unvalidated DNMs out of 37 randomly selected DNMs for validation) were shared by the paired twins, whereas only 40 DNMs were detected in one sibling of the twin, including 20 DNMs in the probands and 20 in the non-affected siblings. Among the 20 DNMs

in the probands, 13 DNMs were located in the intron region, 1 DNM was located in the 5'-upstream region, and the remaining 6 DNMs were located in the intergenic region (Table 2). It is noteworthy that the majority of DNMs in the probands seemed to be enriched in the probands of 3 families (B84, 7 DNMs; B113, 5 DNMs; B124, 3 DNMs) (Table 2). Although these DNMs were not located in the coding region, we could not exclude the possibility that they had roles in gene regulation, as accumulating evidence showing a regulatory effect of mutations/variants in the non-coding region (Ward and Kellis, 2012; Roussos et al., 2014; Xiao et al., 2017). One intronic DNM in proband of B8 family was located in the intron of *NXPH1* (neurexophilin 1) – a gene located in a region with an autism risk CNV (Gai et al., 2012). The *NXPH1* is a ligand for the *NRXN1* protein, which has been reported to be involved in schizophrenia (Missler et al., 1998; Gauthier et al., 2011). The *NRXN1* gene had a significantly differential expression in schizophrenia patients compared with controls (Chen et al., 2013; Lanz et al., 2015) (Table 2).

DNMs shared by twins might also affect schizophrenia in the analyzed families. We focused on the 8 DNMs that were shared by the twins and would lead to residue changes or intron retention.

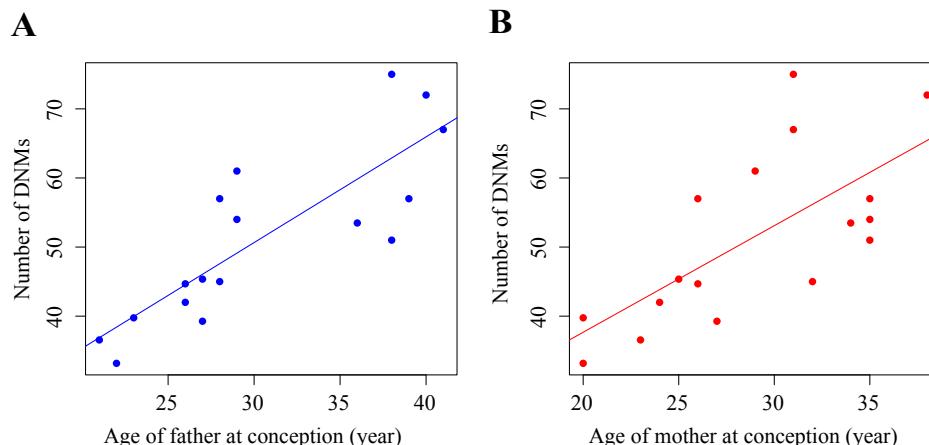


Fig. 2. Correlation between the number of DNMs and parents' age at conception. The number of DNMs in the offspring is plotted against the age of the father (A) and the mother (B). The number of mutations increases with the age of father at conception (p -value = 0.009). The data reported by Michaelson et al. (2012) were (re-)analyzed together with our data.

Table 1

The *de novo* missense and splice-site mutations in 8 pairs of MZ twins.

Sample	Location	Genotype	Gene	Transcript ID	Mutant	SIFT prediction	PolyPhen-2 prediction
B53 B54	chr12:120569035	C/G	GCN1L1	NM_006836	p.S2506T	Tolerated	Benign
B53 B54	chr17:39595049	G/T	KRT38	NM_006771	p.P265H	Damaging	Damaging
B53 B54	chr2:179436794	C/T	TTN	NM_001256850	p.V24689I	NA	NA
B62 B63	chr2:242283282	A/G	SEPT2	NM_001008491	p.N271S	Tolerated	Benign
B93 B94	chr19:57037004	C/G	ZNF471	NM_020813	p.A523G	Damaging	Damaging
B93 B94	chr16:2147410	G/A	PKD1	NM_000296	p.R3438W	Damaging	Damaging
B103 B104	chr10:128776253	G/T	DOCK1	NM_001290223	IVS3+1G > T	NA	NA
B123 B124	chr19:35449194	T/C	ZNF792	NM_175872	p.N522S	Tolerated	Damaging

The potential deleterious effect of each variant was predicted by using SIFT (Ng, 2003; Kumar et al., 2009) and PolyPhen-2 (Adzhubei et al., 2010, 2013). NA, not available. Tolerated, benign and damaging were defined according to SIFT scores (>0.05, tolerated; < 0.05, damaging) and PolyPhen-2 HDIV scores (>0.453, damaging; ≤ 0.452, benign).

Table 2

The *de novo* mutations in the probands and differential expression of the hit genes.

Sample	Location	Genotype	Gene and function	Differential expression (p-value)					RegulomeDB score ^c	
				GSE53987 ^a			GSE35978 ^b			
				Hippocampus	Prefrontal cortex	Striatum	Cerebellum	Prefrontal cortex		
B104	chr2:153664053	G/A	Intergenic	NA	NA	NA	NA	NA	7	
B113	chr4:122576011	T/C	Intergenic	NA	NA	NA	NA	NA	4	
B113	chr20:52177664	A/G	RP4_724E16.2 intron	NA	NA	NA	NA	NA	6	
B113	chr15:96155017	C/A	Intergenic	NA	NA	NA	NA	NA	6	
B113	chr9:127918364	G/A	PPP6C intron	6.2E-06	0.057	0.007	0.75	0.007	6	
B113	chr12:26742696	A/G	ITPR2 intron	0.498	0.002	0.9	0.035	0.961	7	
B124	chr1:230530315	T/C	PGBD5 intron	0.465	0.231	0.003	0.647	7.7E-05	2b	
B124	chr17:12638660	T/C	MYOCD intron	0.888	0.958	0.969	0.6826817	0.127	5	
B124	chr17:12638661	A/G	MYOCD intron	0.888	0.958	0.969	0.6826817	0.127	5	
B54	chr12:76731688	G/T	Intergenic	NA	NA	NA	NA	NA	4	
B63	chr4:1331141	T/C	MAEA intron	2.3E-05	0.446	0.069	0.383	0.001	4	
B84	chr17:41382052	T/C	LINCO0854 5'-upstream	NA	NA	NA	NA	NA	2b	
B84	chr13:45830708	C/T	GTF2F2 intron	0.002	0.698	0.273	0.672	0.159	6	
B84	chr9:94497890	T/C	ROR2 intron	0.012	0.617	0.843	0.9733785	0.00468	6	
B84	chr7:8681356	T/C	NXPH1 intron	2.2E-06	0.865	0.007	0.086	0.00034	7	
B84	chr4:36920104	A/C	Intergenic	NA	NA	NA	NA	NA	7	
B84	chr6:149959476	T/A	KATNA1 intron	0.73	0.766	0.995	0.4299414	0.358	7	
B84	chr2:167878685	A/G	XIRP2 intron	0.036	0.854	0.298	0.6894866	0.643	7	
LJ	chr6:12749681	C/G	PHACTR1 intron	0.008	0.939	0.043	0.4530216	0.0263	3a	
LJ	chr19:3354318	T/C	Intergenic	NA	NA	NA	NA	NA	7	

^a Data from Lanz et al. (2015).

^b Data from Chen et al. (2013).

^c The RegulomeDB (Boyle et al., 2012) score refers to the following available datatypes for a single coordinate. 2b, transcription factor (TF) binding + any motif + DNase Footprint + DNase peak; 3a , TF binding + any motif + DNase peak; 4, TF binding + DNase peak; 5, TF binding or DNase peak; 6, other; 7, no data. NA, not available.

Among the 7 shared non-synonymous mutations (Table 1), three (p.P265H in KRT38; p.A523G in ZNF471 and p.R3438W in PKD1) were predicted to be damaging according to the overlapping results of multiple *in silico* program affiliated predictions. We searched the 7 genes harboring the above non-synonymous mutations (Table 1) in the NPdenovo database (Li et al., 2016) to detect whether these genes had DNM in schizophrenia or other neurodevelopmental disorders. We only found that the TTN gene contained one DNM according to database search, but the DNM is different from the one observed in this study. We observed a similar pattern of family-specific enrichment of shared missense DNM as described above for DNM in the probands, particularly for families B5 and B9 (Table 1). It is worth noting that the TTN (titin; p.V24689I) and GCN1L1 (GCN1 eIF2 alpha kinase activator homolog; p.S2506T) genes identified in family B5 also exhibited *de novo* non-synonymous mutations in schizophrenia patients (Fromer et al., 2014; Wang et al., 2015) and autism patients (Iossifov et al., 2012; Fromer et al., 2014), but the mutations are different from this study. Using the DAPPLE web server (Rossin et al., 2011), we explored protein-protein interactions (PPIs) among 23 genes hit by DNM in the coding regions, UTR regions, 3'-downstream regions or 5'-upstream regions, and the schizophrenia top genes (identified

by convergent functional genomics (CFG) score) of the SZDB data set (Wu et al., 2017) (Fig. S2). We found that TTN and GCN1L1 proteins interacted with a variety of schizophrenia-related proteins, such as DISC1 (disrupted in schizophrenia 1) (Hodgkinson et al., 2004; Thomson et al., 2014; Huang, 2015) and AKT1 (encoding serine-threonine protein kinase) (Emamian et al., 2004; Schwab et al., 2005). However, we need to state that the exact role of these two genes in neurodevelopment of schizophrenia remains to be clarified.

The splice site mutation (IVS3+1G > T) in the DOCK1 (dedicator of cytokinesis 1) gene shared by the twins from B10 family would lead to an intron retention, which might cause the truncated protein (remaining only 57 of 1886 (3.0%) amino acid residues) to be non-functional. In the nervous system, the protein (Dock180) of the DOCK1 gene was essentially involved in the regulation of axon guidance and dendritic spine morphogenesis (Li et al., 2008). The brain-specific angiogenesis inhibitor-1 (BAI-1), an interacting protein of Dock180-ELMO module, had been linked to a variety of neuropsychiatric disorders, including schizophrenia, bipolar disorder, and drug addiction (Park et al., 2007; Lanoue et al., 2013). The Dock proteins have been well recognized to be related to neurological diseases (Shi, 2013).

2.5. Inherited SNPs and schizophrenia

We performed a gene-based association analysis on the basis of all variants in 32 individuals from the 8 families that were scored relative to human reference genome (hg19; <http://genome.ucsc.edu/>). As our sample size was relatively small and had limited statistical power, we therefore chose an arbitrary threshold (p -value < 0.01) in order to sample more risk genes even there is a possibility of false positive hit. A total of 55 genes were captured by our analysis (p -value < 0.01) (Table S8). Among the most significant genes (p -value = 9.99×10^{-4}), *KCNQ2* encodes a subunit of KCNQ which is a voltage-gated K⁺ channel regulating the excitability of neurons (Brown et al., 2007) and variants in this gene had been reported to be associated with schizophrenia (Lee et al., 2013). We further screened the recently reported 128 susceptibility SNPs that met the genome-wide significance (p -value $\leq 5 \times 10^{-8}$) (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014) in our samples. A total of 607 hits from 8 families were identified. The polygenic risk scores based on the 607 hits showed that probands in general had substantially higher score than their mothers (p -value = 0.008, R^2 = 0.583), fathers (p -value = 0.353, R^2 = 0.022), and parents (average, p -value = 0.021, R^2 = 0.468). Because these SNPs used for the calculation of polygenic risk score were all common inherited SNPs and were shared by MZ twins, and the DNM in each twin provided no additional information for the calculation, we could not distinguish the polygenic risk score between the proband and his/her sibling (Table S9 and Fig. 3). Furthermore, we focused on those risk alleles that were homozygous in the offspring but heterozygous in their parents, which would increase the risk of schizophrenia in the offspring. As a result, we identified 38 such variants. Eight of these 38 risk alleles occurred in multiple families (Table S10). 8 families all harbored at least 3 homozygous risk alleles, with the highest number in families B5 and B10 (each had 7). One of the homozygous risk variants in B8 family, rs2007044, influenced the expression of *CACNA1C* (encoding the α -1C subunit of the L-type voltage-gated calcium channel) according to the eQTL analysis (Yang et al., 2010). As an important schizophrenia risk gene, *CACNA1C* has been reported to be involved in neuronal development (Green et al., 2010; Nyegaard et al., 2010). Another homozygous risk variant in B10 family, rs10503253 (located in *CSMD1*), was associated with schizophrenia in several previous reports (Donohoe et al., 2013; Rose et al., 2013; Koiliari et al., 2014). Note that in LI family, we also detected a DNM that was located in the intron region of the *CSMD1* gene (Table S4).

Rare genetic variants with a large effect were thought to play a substantial role in causing schizophrenia (McClellan et al., 2007). We detected all the rare (MAF ≤ 0.01 in the 1000 Genomes Project (Sudmant et al., 2015)), inherited LOF (i.e., stop-gain or frameshift) and damaging missense variants (as predicted by at least two of the

five prediction algorithms) in our samples. In total, 9480 rare damaging or LOF variants were found, with an average of 296 variants per individual (Table S11). However, we found no significant p -value for the gene-based rare variant burden test based on all rare damaging or LOF variants in all samples. It was noteworthy that *LAMC1* (laminin subunit gamma 1), with 5 rare damaging variants in 5 families (Table S12), had a marginal significance (p -value = 0.068). We further focused on these schizophrenia associated genes carrying the rare damaging or LOF variants by referring to 3 classes of gene sets (Materials and methods). In total, we identified 5 Class I genes, 17 Class II genes, and 45 Class III genes containing rare damaging or LOF variants (Table S13).

Among the 38 Class I genes, *RELN* (reelin), *GAD1*, *PLXNA2*, *MTHFR*, and *FEZ1* had rare damaging or LOF variants. The *RELN* gene encodes an extracellular matrix protein essential for cell positioning and neuronal migration during brain development (Tissir and Goffinet, 2003), which was previously reported to be involved in schizophrenia (Costa et al., 2002; Shifman et al., 2008), autism (Persico et al., 2001) and Alzheimer's disease (Seripa et al., 2008). The *RELN* variant p.M927V might contribute to schizophrenia-risk in family B11. The variant p.D180V in *FEZ1* (fasciculation and elongation protein zeta-1) was found in LI family. *FEZ1* was involved in nerve growth and fasciculation; together with *DISC1*, it regulates neuronal development that is related to schizophrenia (Kang et al., 2011). Genetic variants in *FEZ1* had been reported to be associated with schizophrenia in Japanese cohort (Yamada et al., 2004). B5 family had 2 Class I genes with rare damaging mutation: *GAD1* (glutamate decarboxylase 1) and *PLXNA2* (plexin A2). Previous study showed that schizophrenic brain had a decreased expression of the gamma-aminobutyric acid (GABA) synthetic enzyme glutamic acid decarboxylase 67 (GAD67), which is encoded by *GAD1* (Addington et al., 2005; Straub et al., 2007). *PLXNA2* is a member of the semaphorin receptor family and plays a role in the development of axonal projections and in neural regeneration (Winberg et al., 1998). Co-expression network analysis showed that *PLXNA2* and *GAD1* had a similar expression pattern and potential interaction, suggesting a potential combined effect on schizophrenia (Fig. S3).

Among the 206 Class II genes, 2 genes (*GSN* and *ITPR1*) had more than one damaging non-synonymous variant. *GSN* variants p.K546R and p.R144Q were found in families B12 and B8, respectively. The *GSN* gene was found to be significantly down-regulated in a genome-wide expression analysis in a set of schizophrenia patients (Hakak et al., 2001). *ITPR1* played a key role in synaptic plasticity by binding with disrupted-in-schizophrenia 1 (*DISC1*) (Tsuboi et al., 2015), a promising susceptibility factor for schizophrenia (Brandon and Sawa, 2011). The two missense variants p.I2030M (family B5) and p.I1421V (family LI) in *ITPR1* were predicted to be damaging. It is plausible that these variants enacted an effect via altered binding activity with *DISC1*. Intriguingly, we found

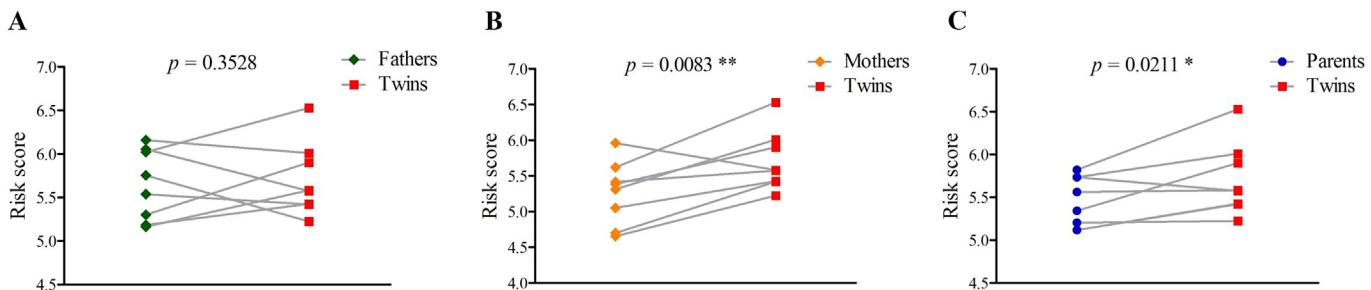


Fig. 3. Polygenic risk score based on 128 schizophrenia-risk SNPs in the comparison of probands and healthy parents. Comparison of the risk score between probands and their fathers (A), mothers (B) and parents (C) was evaluated by using paired t-test. *, $p < 0.05$; **, $p < 0.01$.

that families B5 and B8 appeared to have an enrichment of damaging missense variants (each had 5 variants) in Class II genes, and followed by families B9 and B11 (each had 2 variants) (Table S13).

We found 5 Class III genes that were shared by two families (Table S13). One of these genes was protocadherin alpha 6 (*PCDHA6*), which encodes a potential calcium-dependent cell-adhesion protein and is involved in the establishment and maintenance of specific neuronal connections in the brain (Wu and Maniatis, 1999). The interacting protein of *PCDHA6*, *RELN*, was reported as an important risk gene for schizophrenia (Costa et al., 2002; Shifman et al., 2008).

The enriched rare damaging or LOF variants in schizophrenia-related gene sets may significantly increase the risk of schizophrenia (Fromer et al., 2014). We compared the number of rare damaging variants or LOF variants in 17 core gene sets in parents and offspring (Table S14). Although we did not find a significant *p*-value for the odd ratio (OR) of rare damaging or LOF variants in these gene sets in offspring relative to their parents, partially due to the small sample size, the offspring did show a higher risk than parents in several gene sets (OR > 1.4) (Table S14). In the gene set of the NMDAR network, the offspring had 14 rare damaging variants whereas the parents only had 10 (OR = 1.40; *p*-value = 0.423). The NMDA signaling triggers multiple processes required for inducing synaptic plasticity (Malenka and Nicoll, 1993), and NMDA receptor hypofunction may contribute to the symptomatic features of schizophrenia (Goff and Coyle, 2001).

2.6. Inherited CNVs and schizophrenia

Combining the results from four algorithms (Breakdancer (Chen et al., 2009), CNVnator (Abzyov et al., 2011), Delly (Rausch et al., 2012) and Lumpy (Layer et al., 2014)), we detected 6619 CNVs (length > 1000 bp) in all probands and their parents, with an average of 276 variants per individual (Table S15). There were 4 inherited CNVs (both shared by twin pairs) in the previously reported schizophrenia-risk regions (Table S16) according to the data set that was based on 32 related studies ((Luo et al., 2014) and references therein). The proband from B11 family had a large inherited deletion (16p13.11; 2.9 Mb) contained 10 genes (*MPV17L*, *C16orf45*, *KIAA0430*, *NDE1*, *MYH11*, *C16orf63*, *ABCC1*, *ABCC6*, *NOMO3* and *XYLT1*) (Fig. S4). The *NDE1* (nuclear distribution gene E homolog 1) gene was involved in brain development, neuronal proliferation, migration and synapse formation (Shu et al., 2004). This gene was significantly associated with schizophrenia and had a biologically interaction with *DISC1* and *LIS1* proteins (Burdick et al., 2008; Bradshaw et al., 2011). Moreover, *NDE1*, *MYH11* and *ABCC1* were identified as top schizophrenia candidate genes by using a cumulative scoring to systematically prioritized genes affected by CNVs in schizophrenia (Luo et al., 2014).

3. Discussion

Multiple lines of evidence demonstrated that schizophrenia is a common disease with a complex etiology, involving heterogeneous genetic factors (Saha et al., 2005; Wu et al., 2017). We used WGS as a tool to identify genetic mutations/variants in 8 families with one MZ twin pair discordant for schizophrenia, to provide a comprehensive profile for genetic mutations and variants that might contribute to schizophrenia risk. Theoretically, the choice of identical twins discordant for schizophrenia would offer an extremely rare and valuable opportunity to pinpoint the pathogenic mutation(s) causing schizophrenia. We identified 504 DNMs in offspring and observed a significant positive correlation between father's age at conception of child and the number of DNMs in the offspring,

whereas mother's age at conception was not significant. This finding is consistent with the result of previous studies (Kong et al., 2012; Michaelson et al., 2012; Genome of the Netherlands Consortium, 2014), and could explain the epidemiological observation that the risk of schizophrenia increased significantly with father's age at conception of child in Iceland (Malaspina, 2001).

To pinpoint the potential genetic alterations that would account for schizophrenia in the twin discordant for disease, we focused on DNMs, inherited common risk SNPs, inherited rare damaging or LOF variants in the family members. Among the identified DNMs only in probands, none of them were located in coding region. Nevertheless, whether these DNMs influence the discordance of schizophrenia remains to be elucidated. The missense DNMs shared by twins might also increase the risk of schizophrenia, such as p.V24689I in *TTN*, p.S2506T in *GCN1L1* and IVS3+1G > T in *DOCK1*, to name a few, and functional assays should be performed to validate our speculation.

Previous studies found no discordant DNA variants in MZ twins with discordant disease (Baranzini et al., 2010; Chaiyasadaporn et al., 2014; Petersen et al., 2014), suggesting that DNMs might partially account for disease, together with other genetic factors. All these variants with potentially small effect sizes were combined into one big effect network, rather than few independent variants, to increase the inherited risk of common diseases such as schizophrenia (International Schizophrenia et al., 2009; Kraft and Hunter, 2009). Presently, there were no single mutation that could have enough effect to cause schizophrenia (McCarthy et al., 2008). Given the characteristics of pathogenic complexity and high heritability in schizophrenia, we also analyzed the effect of inherited variants on schizophrenia. The gene-based burden analysis identified many genes that might increase the risk of schizophrenia, such as *KCNQ2*. We also found 38 common inherited variants that were homozygous in offspring but heterozygous in their parents, and these variants might be functional according to data-mining of available datasets. For instance, SNP rs2007044 from B8 family could change the expression of *CACNA1C* during the neuronal development (Green et al., 2010; Nyegaard et al., 2010), and SNP rs10503253 in *CSMD1* from B10 family was associated with schizophrenia (Donohoe et al., 2013; Rose et al., 2013; Koiliari et al., 2014). Further analysis of polygenic risk score based on the 128 schizophrenia susceptible SNPs (Schizophrenia Working Group of the Psychiatric Genomics, 2014) indicated that probands had a higher risk for schizophrenia than their parents.

Rare genetic variants, as well as common variants, are thought to contribute to schizophrenia, but rare variants may exert a larger effect than common variants (McClellan et al., 2007; Sebat et al., 2009). By searching the rare damaging variants or LOF variants from three classes of schizophrenia-related genes, we found some important genetic alterations in these schizophrenia-related genes that might be linked to the onset of schizophrenia, such as *GAD1*, *PLXNA2*, *RELN* and *FEZ1*. Moreover, multiple genetic factors, including DNMs, common inherited variants, rare inherited damaging variants and LOF variants, might increase risk for schizophrenia in these families (Fig. 4). For instance, B5 family not only had two missense DNMs in *TTN* and *GCN1L1*, but also had rare damaging variants in *GAD1* and *PLXNA2*, which suggested a combined effect of DNMs and inherited rare damaging variants on schizophrenia. It is noteworthy that B8 and B12 families shared two schizophrenia-associated genes which had rare damaging variants, *GSN* (p.R144Q mutation in B8 family, and p.K546R mutation in B12 family) and *PCDHA6* (p.N403T mutation in B8 and B12 families). These results indicated that the two families might have been subjected to similar genetic risk factors of schizophrenia.

Furthermore, we systematically investigated the interaction network among proteins encoded by genes harboring *de novo*

missense mutations and inherited highly risky or rare damaging mutations mentioned above (Tables 1, S10, S13 and S16). We found that all genes contained in the GeneMANIA database (Montojo et al., 2014) would constitute a network (Fig. S5). This result suggested these genes are involved in one shared interaction network to increase risk for schizophrenia with small additive effects.

The current study had several limitations. First, we collected only 8 families with MZ twin pairs discordant for schizophrenia. The sample size might have limited statistical power in the context of genome-wide sequence data, but this constitutes the largest sample for WGS of MZ twin pairs discordant for schizophrenia. Second, we analyzed the genetic alterations in blood samples, but we could not exclude a possibility of *in situ* DNMs in the schizophrenia-associated brain regions. Moreover, gene expression and epigenetic changes in these families were not analyzed, which had been reported to be an important factor in the pathogenesis of psychiatric disorders (Roth et al., 2009; Dempster et al., 2011; Zong et al., 2015). Finally, we did not perform functional assays to elucidate the biological implications of these missense DNMs and to validate the combined effect of estimated polygenic factors (Li et al., 2017b).

In short, we analyzed the potential risk factors of schizophrenia by using WGS data in 8 families with one MZ twin pair showing a discordance of schizophrenia. We did not identify any specific genes that played a prevalent role in schizophrenia, but we found a potentially combined effect of many genetic alterations, including DNMs, common inherited variants, rare inherited damaging variants and LOF variants, on schizophrenia. These results confirmed the heterogeneous nature of schizophrenia and might have important implications for clinical diagnosis of patients with schizophrenia. When investigating the genetics of schizophrenia, we need to search for rare and common variants, CNVs, and DNMs.

4. Materials and methods

4.1. Sample collection

Eight families of MZ twins discordant for schizophrenia were recruited in this study. These samples were analyzed for mtDNA heteroplasmy in our recent study (Li et al., 2017a). The

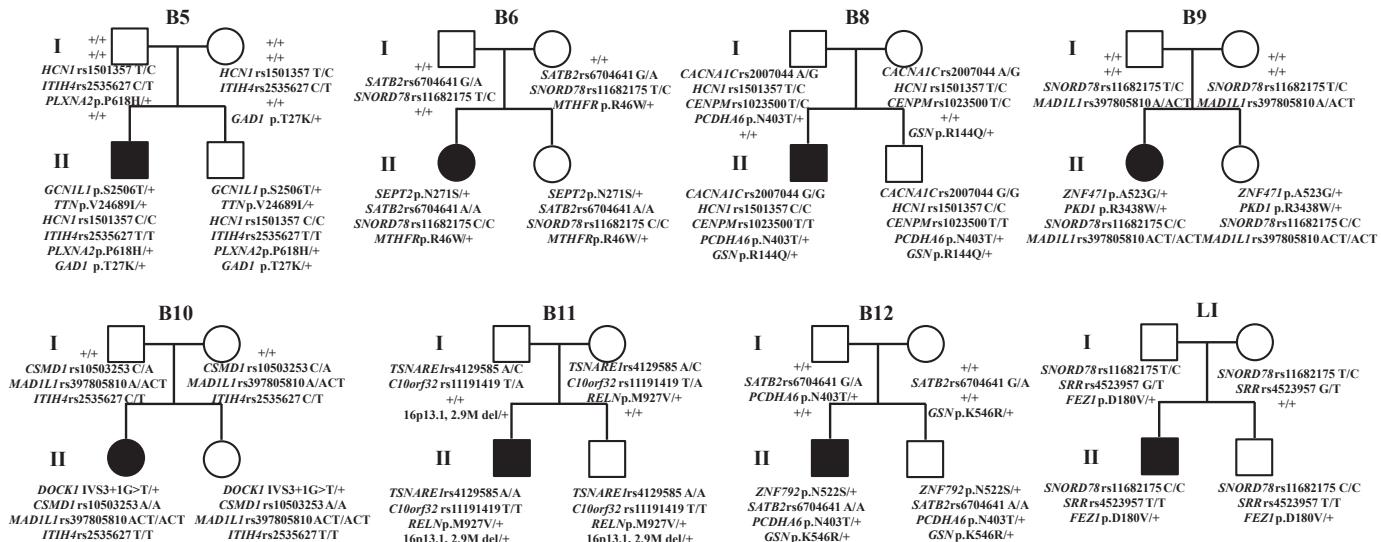
schizophrenia patients were diagnosed independently by two psychiatrists following the criterion of Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition (DSM-IV). Informed consents conforming to the tenets of the Declaration of Helsinki were obtained from each participant or guardian prior to this study. The institutional review board of the Second Xiangya Hospital approved this study.

4.2. Whole genome sequencing

Genomic DNAs from 8 identical twin pairs and their parents were isolated from the peripheral blood by using AxyPrep™ Blood Genomic DNA Miniprep Kit (Axygen, USA) according to the manufacturer's instruction. Deep (~30×) whole genome sequencing was performed at the BGI in Shenzhen using the Illumina HiSeq 2000 Platform (500 bp library, 90 bp reads). One individual (B114) of family B11 was sequenced at the Novogene corporation in Beijing using Illumina HiSeq X Ten Platform (500 bp library, 150 bp reads).

4.3. Alignment and variant calling

Low-quality bases of raw reads were removed using Trimmomatic-0.32 (Bolger et al., 2014) with the parameters as "LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36". Quality-filtered reads were mapped to hg19 reference genome by BWA version 0.79a (Li and Durbin, 2009). The BAM file was then sorted by genomic position and indexed using SortSam in picard-tools-1.107 (<https://github.com/broadinstitute/picard>). The BAM files for each sample were then merged into a single BAM file using MergeSamFiles in picard-tools-1.107. To prevent PCR artifacts from influencing the downstream analysis of our data, we used MarkDuplicates in picard-tools-1.107 to mark the duplicate reads, which were ignored in downstream analysis. GATK version 2.8 (DePristo et al., 2011) was used for SNV discovery by the parameters as recommended (http://www.broadinstitute.org/gsa/wiki/index.php/Best_Practice_Variant_Detection_with_the_GATK_v3). We used the GATK unified genotyper (UG) for SNV discovery and calculation of genotype likelihoods on each family with "trio" mode. We considered all SNVs called by the GATK UG with a Phred-quality score > Q10 as a starting point before filtering in order to



maximize sensitivity. We used the GATK Variant Quality Score Recalibration (VQSR) to filter spurious SNVs due to sequencing errors and mapping artifacts. We annotated all variants according to RefSeq gene transcripts (accessed from the UCSC Genome Browser, <http://genome.ucsc.edu>) using our house script.

4.4. De novo SNPs and indel detection

In order to accurately distinguish true *de novo* SNPs from errors, we used machine-learning classifier called forestDNM (Michaelson et al., 2012) to predict the validation status of the putative DNM in all families. The method was designed to distinguish *de novo* variants from false-positive DNM that arose from errors in sequencing, alignment, or variant calling. We applied following filtering criteria for *de novo* indel detection: a) Phred-scaled quality score (QUAL) of all family members more than Q30; b) QualByDepth (QD: variant confidence from the QUAL field/unfiltered depth) more than 10; c) MappingQualityZero (MQ0: total counts across all samples that have reads with a mapping quality of zero) less than 4; d) The call from the offspring needed to be supported by at least 30% of the reads, and there needed to be at least a 17× read depth; e) The *de novo* indel calls where both parents were genotyped as homozygous reference and at the offspring was genotyped as heterozygous; f) The *de novo* indels were also detected by PINDEL (Ye et al., 2009).

4.5. Detection of CNVs

The CNVs (>1 Kb) were calculated by combining the algorithms of Breakdancer (Chen et al., 2009), CNVnator (Abyzov et al., 2011), Delly (Rausch et al., 2012) and Lumpy (Layer et al., 2014). The putative inherited CNVs were detected by 2 or more algorithms (80% reciprocal overlap) and were present in at least one parent. Putative *de novo* CNVs were selected from calls detected by 4 algorithms (80% reciprocal overlap) and were not present in their parents or in any other individuals. The total list of the SNPs, indels and CNVs has been included in the SZDB dataset (www.szdb.org) (Wu et al., 2017).

4.6. Experimental validation

37 candidate DNM were selected randomly for validation by Sanger sequencing. Primers were designed to span at least 150 bp upstream and downstream of the candidate DNM (Table S5).

4.7. DNMs and father's age at conception of child

We combined the data reported by Michaelson et al. (2012) with our data to detect the potential correlation between the number of DNM and parents' age at conception. As the DNM that we identified had a false-positive variant calling rate around 21.6%, we adjusted the number of DNM by multiplying the positive rate. The age of the fathers and mothers in our study were strongly correlated ($R^2 = 0.83$), therefore it is difficult to distinguish a father's age effect from a mother's age effect. To separate the age effect of the father and/or mother, we used multiple regression to test the relationship between number of DNM and fathers' or/and mother's age at conception. B11 family was excluded from further analysis because we generated the sequencing data from 2 different sequencing platforms.

4.8. Polygenic risk score analysis and comparison

Polygenic risk scores were calculated using the method described by Ahn et al. (2016). We scored our samples using the most recent 128 schizophrenia-risk SNPs (Schizophrenia Working

Group of the Psychiatric Genomics, 2014). The log of odds ratio of an allele was multiplied by 0, 1 or 2 depending on the number of reference alleles. The total polygenic score is a sum of across SNPs. For the comparisons of probands' polygenic risk scores and their mothers, fathers or parents (average), we used the paired *t*-test.

4.9. Identification of rare inherited variants in schizophrenia-related gene sets

Five prediction algorithms (SIFT (Ng, 2003; Kumar et al., 2009), PolyPhen-2 HumDiv, PolyPhen-2 HumVar (Adzhubei et al., 2010, 2013), LRT (Chun and Fay, 2009) and MutationTaster (Schwarz et al., 2014)) were used for predicting potential damaging effect. Missense variants were rated as damaging when at least 2 of the 5 predictions suggested a potential deleterious effect. We followed the prediction of the dbNSFP database (Liu et al., 2016). Rare ($MAF \leq 0.01$ in the 1000 Genomes Project (Sudmant et al., 2015)) and potential damaging variants including nonsense, frameshift, and damaging missense variants were selected for further analysis. All schizophrenia-related genes carrying rare damaging or LOF variants were considered and were grouped into 3 classes:

- Class I – 38 genes listed in the core gene set of SZGR (Jia et al., 2010).
- Class II – 206 genes in schizophrenia-related pathways or complexes, including postsynaptic density protein 95 (PSD-95) complexes, neuronal activity-regulated cytoskeleton-associated protein (ARC) postsynaptic signaling complexes, N-methyl-d-aspartate receptor (NMDAR) network and mGluR5 complexes. The entire list was compiled by Kirov et al. (2012).
- Class III – 352 genes were taken from the 108 schizophrenia susceptibility loci showing a genome-wide significance (Schizophrenia Working Group of the Psychiatric Genomics, 2014).

4.10. Gene-based test

The gene-based burden test was performed by using the PLINK/seq (<https://atgu.mgh.harvard.edu/plinkseq/>) based on all variants in the analyzed subjects relative to human reference sequence (hg19; <http://genome.ucsc.edu/>). We also performed the same test based on rare damaging variants or LOF variants.

4.11. Polygenic burden analysis of rare variants for schizophrenia-related gene sets

We selected the following gene sets for polygenic burden analysis:

- 685 genes from the postsynaptic density (PSD) "human core" gene sets compiled by Kirov et al. (2012), which include NMDAR complex, PSD-95 and mGluR5 subsets.
- 431 genes from pre-synapse gene set compiled by Kirov et al. (2012), which include pre-synaptic active zone and synaptic vesicle.
- 352 genes related to the 108 schizophrenia susceptibility loci of genome-wide significance (Schizophrenia Working Group of the Psychiatric Genomics, 2014).
- 611 genes with a non-synonymous DNM reported by Fromer et al. (2014).
- 1589 genes listed as targets of FMRP were taken from Darnell et al. (2011) and Ascano et al. (2012).
- 432 genes related to axon guidance pathways, 237 genes related to nerve impulse, 1847 genes related to nervous

system development, 309 genes related to neuronal cell body, and 699 genes related to neuron projection were taken from Yuen et al. (2015).

The enrichment analysis of rare variants in each gene set was conducted using the Fisher's exact test.

4.12. Other published datasets for data mining

Distinction of rare variants and common variants depended on the MAF in the 1000 Genomes Project (Sudmant et al., 2015): the cut-off of rare variants is $MAF \leq 0.01$ and the cut-off of common variants is $MAF > 0.01$. The eQTL analysis was performed by Genewvar analysis (Yang et al., 2010). The *p*-values of differential expression were taken from SZDB (Wu et al. (2017) and references therein), which was estimated by using datasets of GSE53987 (Lanz et al., 2015) and GSE35978 (Chen et al., 2013) from Gene Expression Omnibus (GEO) (Clough and Barrett, 2016). *de novo* SNVs had been annotated with known and predicted regulatory elements by RegulomeDB (Boyle et al., 2012), including DNAase hypersensitivity sites, binding sites of transcription factors, and promoter regions.

Acknowledgments

We are grateful to the subjects who donated DNA samples. This study was supported by the Strategic Priority Research Program (B) of the Chinese Academy of Sciences (XDB02020003 and XDB02030002), the Bureau of Frontier Sciences and Education, Chinese Academy of Sciences (QYZDJ-SSW-SMC005) and the National Natural Science Foundation of China (Nos. 81088001, 81271484, 81471361 and 81371480), the Beijing Training Project for the Leading Talents in S & T (Z151100000315020), the National Key Basic Research and Development Program (973) (2012CB517904), the CAS/SAFEA International Partnership Programme for Creative Research Teams (Y2CX131003).

Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.jgg.2017.05.005>.

References

- Abyzov, A., Urban, A.E., Snyder, M., Gerstein, M., 2011. CNVnator: an approach to discover, genotype, and characterize typical and atypical CNVs from family and population genome sequencing. *Genome Res.* 21, 974–984.
- Addington, A.M., Gornick, M., Duckworth, J., Sporn, A., Gogtay, N., Bobb, A., Greenstein, D., Lenane, M., Gochman, P., Baker, N., Balkissoon, R., Vakkalanka, R.K., Weinberger, D.R., Rapoport, J.L., Straub, R.E., 2005. *GAD1* (2q31.1), which encodes glutamic acid decarboxylase (GAD67), is associated with childhood-onset schizophrenia and cortical gray matter volume loss. *Mol. Psychiatry* 10, 581–588.
- Adzhubei, I., Jordan, D.M., Sunyaev, S.R., 2013. Predicting functional effect of human missense mutations using PolyPhen-2. *Cur. Protoc. Hum. Genet. Unit* 7 (Chapter 7), 20.
- Adzhubei, I.A., Schmidt, S., Peshkin, L., Ramensky, V.E., Gerasimova, A., Bork, P., Kondrashov, A.S., Sunyaev, S.R., 2010. A method and server for predicting damaging missense mutations. *Nat. Med.* 7, 248–249.
- Ahn, K., An, S.S., Shugart, Y.Y., Rapoport, J.L., 2016. Common polygenic variation and risk for childhood-onset schizophrenia. *Mol. Psychiatry* 21, 94–96.
- Ascano Jr., M., Mukherjee, N., Bandaru, P., Miller, J.B., Nusbaum, J.D., Corcoran, D.L., Langlois, C., Munschauer, M., Dewell, S., Hafner, M., Williams, Z., Ohler, U., Tuschl, T., 2012. FMRP targets distinct mRNA sequence elements to regulate protein expression. *Nature* 492, 382–386.
- Bae, B.I., Tietjen, I., Atabay, K.D., Ervony, G.D., Johnson, M.B., Asare, E., Wang, P.P., Murayama, A.Y., Im, K., Lisgo, S.N., Overman, L., Sestan, N., Chang, B.S., Barkovich, A.J., Grant, P.E., Topcu, M., Politsky, J., Okano, H., Piao, X., Walsh, C.A., 2014. Evolutionarily dynamic alternative splicing of *GPR56* regulates regional cerebral cortical patterning. *Science* 343, 764–768.
- Baranzini, S.E., Mudge, J., van Velkinburgh, J.C., Khanhhanian, P., Khrebtukova, I., Miller, N.A., Zhang, L., Farmer, A.D., Bell, C.J., Kim, R.W., May, G.D., Woodward, J.E., Caillier, S.J., McElroy, J.P., Gomez, R., Pando, M.J., Clendenen, L.E., Ganusova, E.E., Schilkey, F.D., Ramaraj, T., Khan, O.A., Huntley, J.J., Luo, S., Kwok, P.Y., Wu, T.D., Schroth, G.P., Oksenberg, J.R., Hauser, S.L., Kingsmore, S.F., 2010. Genome, epigenome and RNA sequences of monozygotic twins discordant for multiple sclerosis. *Nature* 464, 1351–1356.
- Bhagwat, M., 2010. Searching NCBI's dbSNP database. *Cur. Protoc. Bioinforma. Unit 1* (Chapter 1), 19.
- Bolger, A.M., Lohse, M., Usadel, B., 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30, 2114–2120.
- Boyle, A.P., Hong, E.L., Hariharan, M., Cheng, Y., Schaub, M.A., Kasowski, M., Karczewski, K.J., Park, J., Hitz, B.C., Weng, S., Cherry, J.M., Snyder, M., 2012. Annotation of functional variation in personal genomes using RegulomeDB. *Genome Res.* 22, 1790–1797.
- Bradshaw, N.J., Soares, D.C., Carlyle, B.C., Ogawa, F., Davidson-Smith, H., Christie, S., Mackie, S., Thomson, P.A., Porteous, D.J., Millar, J.K., 2011. PKA phosphorylation of NDEL1 is DISC1/PDE4 dependent and modulates its interaction with LIS1 and NDEL1. *J. Neurosci.* 31, 9043–9054.
- Brandon, N.J., Sawa, A., 2011. Linking neurodevelopmental and synaptic theories of mental illness through DISC1. *Nat. Rev. Neurosci.* 12, 707–722.
- Brown, D.A., Hughes, S.A., Marsh, S.J., Tinker, A., 2007. Regulation of M(Kv7.2/7.3) channels in neurons by PIP(2) and products of PIP(2) hydrolysis: significance for receptor-mediated inhibition. *J. Physiol.* 582, 917–925.
- Burdick, K.E., Kamiya, A., Hodgkinson, C.A., Lencz, T., DeRosse, P., Ishizuka, K., Elashvili, S., Arai, H., Goldman, D., Sawa, A., Malhotra, A.K., 2008. Elucidating the relationship between DISC1, NDEL1 and NDE1 and the risk for schizophrenia: evidence of epistasis and competitive binding. *Hum. Mol. Genet.* 17, 2462–2473.
- Chaiyaporn, P., Kulawonganunchai, S., Srichomthong, C., Tongsim, S., Suphapeetiporn, K., Shotelersuk, V., 2014. Whole genome and exome sequencing of monozygotic twins with trisomy 21, discordant for a congenital heart defect and epilepsy. *PLoS One* 9, e100191.
- Chen, C., Cheng, L., Grennan, K., Pibiri, F., Zhang, C., Badner, J.A., Members of the Bipolar Disorder Genome Study C, Gershon, E.S., Liu, C., 2013. Two gene co-expression modules differentiate psychotics and controls. *Mol. Psychiatry* 18, 1308–1314.
- Chen, K., Wallis, J.W., McLellan, M.D., Larson, D.E., Kalicki, J.M., Pohl, C.S., McGrath, S.D., Wendt, M.C., Zhang, Q., Locke, D.P., Shi, X., Fulton, R.S., Ley, T.J., Wilson, R.K., Ding, L., Mardis, E.R., 2009. BreakDancer: an algorithm for high-resolution mapping of genomic structural variation. *Nat. Methods* 6, 677–681.
- Chun, S., Fay, J.C., 2009. Identification of deleterious mutations within three human genomes. *Genome Res.* 19, 1553–1561.
- Clough, E., Barrett, T., 2016. The gene expression omnibus database. *Methods Mol. Biol.* 1418, 93–110.
- CNV and Schizophrenia Working Groups of the Psychiatric Genomics Consortium, 2017. Contribution of copy number variants to schizophrenia from a genome-wide study of 41,321 subjects. *Nat. Genet.* 49, 27–35.
- Costa, E., Chen, Y., Davis, J., Dong, E., Noh, J.S., Tremolizzo, L., Veldic, M., Grayson, D.R., Guidotti, A., 2002. REELIN and schizophrenia: a disease at the interface of the genome and the epigenome. *Mol. Interv.* 2, 47–57.
- Darnell, J.C., Van Driesche, S.J., Zhang, C., Hung, K.Y., Mele, A., Fraser, C.E., Stone, E.F., Chen, C., Fak, J.J., Chi, S.W., Licatalosi, D.D., Richter, J.D., Darnell, R.B., 2011. FMRP stalls ribosomal translocation on mRNAs linked to synaptic function and autism. *Cell* 146, 247–261.
- Dempster, E.L., Pidsley, R., Schalkwyk, L.C., Owens, S., Georgiades, A., Kane, F., Kalidindi, S., Picchioni, M., Kravariti, E., Toulopoulou, T., Murray, R.M., Mill, J., 2011. Disease-associated epigenetic changes in monozygotic twins discordant for schizophrenia and bipolar disorder. *Hum. Mol. Genet.* 20, 4786–4796.
- DePristo, M.A., Banks, E., Poplin, R., Garimella, K.V., Maguire, J.R., Hartl, C., Philippakis, A.A., del Angel, G., Rivas, M.A., Hanna, M., McKenna, A., Fennell, T.J., Kernytsky, A.M., Sivachenko, A.Y., Cibulskis, K., Gabriel, S.B., Altshuler, D., Daly, M.J., 2011. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat. Genet.* 43, 491–498.
- Donohoe, G., Walters, J., Hargreaves, A., Rose, E.J., Morris, D.W., Fahey, C., Bellini, S., Cummins, E., Giegling, I., Hartmann, A.M., Moller, H.J., Muglia, P., Owen, M.J., Gill, M., O'Donovan, M.C., Tropea, D., Rujescu, D., Corvin, A., 2013. Neuro-psychological effects of the *CSMD1* genome-wide associated schizophrenia risk variant rs10503253. *Genes Brain Behav.* 12, 203–209.
- Emamian, E.S., Hall, D., Birnbaum, M.J., Karayiorgou, M., Gogos, J.A., 2004. Convergent evidence for impaired AKT1-GSK3beta signaling in schizophrenia. *Nat. Genet.* 36, 131–137.
- Fromer, M., Pocklington, A.J., Kavanagh, D.H., Williams, H.J., Dwyer, S., Gormley, P., Georgieva, L., Rees, E., Palta, P., Ruderfer, D.M., Carrera, N., Humphreys, I., Johnson, J.S., Roussos, P., Barker, D.D., Banks, E., Milanova, V., Grant, S.G., Hannon, E., Rose, S.A., Chambert, K., Mahajan, M., Scolnick, E.M., Moran, J.L., Kirov, G., Palotie, A., McC Carroll, S.A., Holmans, P., Sklar, P., Owen, M.J., Purcell, S.M., O'Donovan, M.C., 2014. *De novo* mutations in schizophrenia implicate synaptic networks. *Nature* 506, 179–184.
- Gai, X., Xie, H.M., Perin, J.C., Takahashi, N., Murphy, K., Wenocur, A.S., D'Arcy, M., O'Hara, R.J., Goldmuntz, E., Grice, D.E., Shaikh, T.H., Hakonarson, H., Buxbaum, J.D., Elia, J., White, P.S., 2012. Rare structural variation of synapse and neurotransmission genes in autism. *Mol. Psychiatry* 17, 402–411.
- Gauthier, J., Siddiqui, T.J., Huashan, P., Yokomaku, D., Hamdan, F.F., Champagne, N., Lapointe, M., Spiegelman, D., Noreau, A., Lafreniere, R.G., Fathall, F., Joober, R., Krebs, M.O., DeLisi, L.E., Mottron, L., Fombonne, E., Michaud, J.L., Drapeau, P., Carbonetto, S., Craig, A.M., Rouleau, G.A., 2011. Truncating mutations in *NRXN2* and *NRXN1* in autism spectrum disorders and schizophrenia. *Hum. Genet.* 130,

- 563–573.
- Genome of the Netherlands Consortium, 2014. Whole-genome sequence variation, population structure and demographic history of the Dutch population. *Nat. Genet.* 46, 818–825.
- Goff, D.C., Coyle, J.T., 2001. The emerging role of glutamate in the pathophysiology and treatment of schizophrenia. *Am. J. Psychiatry* 158, 1367–1377.
- Green, E.K., Grozeva, D., Jones, I., Jones, L., Kirov, G., Caesar, S., Gordon-Smith, K., Fraser, C., Forty, L., Russell, E., Hampshire, M.L., Moskvina, V., Nikолов, I., Farmer, A., McGuffin, P., Wellcome Trust Case Control C., Holmans, P.A., Owen, M.J., O'Donovan, M.C., Craddock, N., 2010. The bipolar disorder risk allele at *CACNA1C* also confers risk of recurrent major depression and of schizophrenia. *Mol. Psychiatry* 15, 1016–1022.
- Hakak, Y., Walker, J.R., Li, C., Wong, W.H., Davis, K.L., Buxbaum, J.D., Haroutunian, V., Fienberg, A.A., 2001. Genome-wide expression analysis reveals dysregulation of myelination-related genes in chronic schizophrenia. *Proc. Natl. Acad. Sci. U. S. A.* 98, 4746–4751.
- Haldane, J.B., 2004. The rate of spontaneous mutation of a human gene. 1935. *J. Genet.* 83, 235–244.
- Hodgkinson, C.A., Goldman, D., Jaeger, J., Persaud, S., Kane, J.M., Lipsky, R.H., Malhotra, A.K., 2004. Disrupted in schizophrenia 1 (DISC1): association with schizophrenia, schizoaffective disorder, and bipolar disorder. *Am. J. Hum. Genet.* 75, 862–872.
- Huang, Q., 2015. Genetic study of complex diseases in the post-GWAS era. *J. Genet. Genomics* 41, 87–98.
- International Schizophrenia Consortium, Purcell, S.M., Wray, N.R., Stone, J.L., Visscher, P.M., O'Donovan, M.C., Sullivan, P.F., Sklar, P., 2009. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature* 460, 748–752.
- Iossifov, I., Ronemus, M., Levy, D., Wang, Z., Hakker, I., Rosenbaum, J., Yamrom, B., Lee, Y.H., Narzisi, G., Leotta, A., Kendall, J., Grabowska, E., Ma, B., Marks, S., Rodgers, L., Stepansky, A., Troge, J., Andrews, P., Bekritsky, M., Pradhan, K., Ghaban, E., Kramer, M., Parla, J., Demeter, R., Fulton, L.L., Fulton, R.S., Magrini, V.J., Ye, K., Darnell, J.C., Darnell, R.B., Mardis, E.R., Wilson, R.K., Schatz, M.C., McCombie, W.R., Wigler, M., 2012. *De novo* gene disruptions in children on the autistic spectrum. *Neuron* 74, 285–299.
- Jia, P., Sun, J., Guo, A.Y., Zhao, Z., 2010. SZGR: a comprehensive schizophrenia gene resource. *Mol. Psychiatry* 15, 453–462.
- Jiang, Y.H., Yuen, R.K., Jin, X., Wang, M., Chen, N., Wu, X., Ju, J., Mei, J., Shi, Y., He, M., Wang, G., Liang, J., Wang, Z., Cao, D., Carter, M.T., Chrysler, C., Drmic, I.E., Howe, J.L., Lau, L., Marshall, C.R., Merico, D., Nalpathamkalam, T., Thiruvahindrapuram, B., Thompson, A., Uddin, M., Walker, S., Luo, J., Agnostou, E., Zwaigenbaum, L., Ring, R.H., Wang, J., Lajonchere, C., Wang, J., Shih, A., Szatmari, P., Yang, H., Dawson, G., Li, Y., Scherer, S.W., 2013. Detection of clinically relevant genetic variants in autism spectrum disorder by whole-genome sequencing. *Am. J. Hum. Genet.* 93, 249–263.
- Kang, E., Burdick, K.E., Kim, J.Y., Duan, X., Guo, J.U., Sailor, K.A., Jung, D.E., Ganeshan, S., Choi, S., Prajidan, D., Lu, B., Avramopoulos, D., Christian, K., Malhotra, A.K., Song, H., Ming, G.L., 2011. Interaction between FEZ1 and DISC1 in regulation of neuronal development and risk for schizophrenia. *Neuron* 72, 559–571.
- Kirop, G., Pocklington, A.J., Holmans, P., Ivanov, D., Ikeda, M., Ruderfer, D., Moran, J., Chambert, K., Toncheva, D., Georgieva, L., Grozeva, D., Fjodorova, M., Wollerton, R., Rees, E., Nikолов, I., van de Lagemaat, L.N., Bayes, A., Fernandez, E., Olason, P.I., Bottcher, Y., Komiya, N.H., Collins, M.O., Choudhary, J., Stefansson, K., Stefansson, H., Grant, S.G., Purcell, S., Sklar, P., O'Donovan, M.C., Owen, M.J., 2012. *De novo* CNV analysis implicates specific abnormalities of postsynaptic signalling complexes in the pathogenesis of schizophrenia. *Mol. Psychiatry* 17, 142–153.
- Koilari, E., Roussos, P., Pasparakis, E., Lencz, T., Malhotra, A., Siever, L.J., Giakoumaki, S.G., Bitsios, P., 2014. The *CSMD1* genome-wide associated schizophrenia risk variant rs10503253 affects general cognitive ability and executive function in healthy males. *Schizophr. Res.* 154, 42–47.
- Kong, A., Frigge, M.L., Masson, G., Besenbacher, S., Sulem, P., Magnusson, G., Gudjonsson, S.A., Sigurdsson, A., Jonasdottir, A., Jonasdottir, A., Wong, W.S., Sigurdsson, G., Walters, G.B., Steinberg, S., Helgason, H., Thorleifsson, G., Gudbjartsson, D.F., Helgason, A., Magnusson, O.T., Thorsteinsdottir, U., Stefansson, K., 2012. Rate of *de novo* mutations and the importance of father's age to disease risk. *Nature* 488, 471–475.
- Koren, A., Polak, P., Nemesh, J., Michaelson, J.J., Sebat, J., Sunyaev, S.R., McCarron, S.A., 2012. Differential relationship of DNA replication timing to different forms of human mutation and variation. *Am. J. Hum. Genet.* 91, 1033–1040.
- Kraft, P., Hunter, D.J., 2009. Genetic risk prediction—are we there yet? *N. Engl. J. Med.* 360, 1701–1703.
- Kumar, P., Henikoff, S., Ng, P.C., 2009. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat. Protoc.* 4, 1073–1081.
- Kwon, E., Wang, W., Tsai, L.H., 2013. Validation of schizophrenia-associated genes *CSMD1*, *C10orf26*, *CACNA1C* and *TCF4* as miR-137 targets. *Mol. Psychiatry* 18, 11–12.
- Lanoue, V., Usardi, A., Sigoillot, S.M., Talleur, M., Iyer, K., Mariani, J., Isope, P., Vodjdani, G., Heintz, N., Selimi, F., 2013. The adhesion-GPCR *BAI3*, a gene linked to psychiatric disorders, regulates dendrite morphogenesis in neurons. *Mol. Psychiatry* 18, 943–950.
- Lanz, T.A., Joshi, J.J., Reinhart, V., Johnson, K., Grantham 2nd, L.E., Volkson, D., 2015. STEP levels are unchanged in pre-frontal cortex and associative striatum in post-mortem human brain samples from subjects with schizophrenia, bipolar disorder and major depressive disorder. *PLoS One* 10, e0121744.
- Layer, R.M., Chiang, C., Quinlan, A.R., Hall, I.M., 2014. LUMPY: a probabilistic framework for structural variant discovery. *Genome Biol.* 15, R84.
- Lee, S.H., DeCandia, T.R., Ripke, S., Yang, J., Schizophrenia Psychiatric Genome-Wide Association Study C., International Schizophrenia C., Molecular Genetics of Schizophrenia C., Sullivan, P.F., Goddard, M.E., Keller, M.C., Visscher, P.M., Wray, N.R., 2012. Estimating the proportion of variation in susceptibility to schizophrenia captured by common SNPs. *Nat. Genet.* 44, 247–250.
- Lee, Y.H., Kim, J.H., Song, G.G., 2013. Pathway analysis of a genome-wide association study in schizophrenia. *Gene* 525, 107–115.
- Li, H., Bi, R., Fan, Y., Wu, Y., Tang, Y., Li, Z., He, Y., Zhou, J., Tang, J., Chen, X., Yao, Y.G., 2017a. mtDNA heteroplasmny in monozygotic twins discordant for schizophrenia. *Mol. Neurobiol.* <http://dx.doi.org/10.1007/s12035-016-9996-x>.
- Li, H., Durbin, R., 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25, 1754–1760.
- Li, J., Cai, T., Jiang, Y., Chen, H., He, X., Chen, C., Li, X., Shao, Q., Ran, X., Li, Z., Xia, K., Liu, C., Sun, Z.S., Wu, J., 2016. Genes with *de novo* mutations are shared by four neuropsychiatric disorders discovered from NPdenovo database. *Mol. Psychiatry* 21, 290–297.
- Li, M., Weinberger, D.R., 2017b. Illuminating the dark road from schizophrenia genetic associations to disease mechanisms. *Natl. Sci. Rev.* 4, 240–251.
- Li, X., Gao, X., Liu, G., Xiong, W., Wu, J., Rao, Y., 2008. Netrin signal transduction and the guanine nucleotide exchange factor DOCK180 in attractive signaling. *Nat. Neurosci.* 11, 28–35.
- Liu, H., Abecasis, G.R., Heath, S.C., Knowles, A., Demars, S., Chen, Y.J., Roos, J.L., Rapoport, J.L., Gogos, J.A., Karayiorgou, M., 2002. Genetic variation in the 22q11 locus and susceptibility to schizophrenia. *Proc. Natl. Acad. Sci. U. S. A.* 99, 16859–16864.
- Liu, X., Wu, C., Li, C., Boerwinkle, E., 2016. dbNSFP v3.0: a one-stop database of functional predictions and annotations for human nonsynonymous and splice-site SNVs. *Hum. Mutat.* 37, 235–241.
- Luo, X., Huang, L., Han, L., Luo, Z., Hu, F., Tieu, R., Gan, L., 2014. Systematic prioritization and integrative analysis of copy number variations in schizophrenia reveal key schizophrenia susceptibility genes. *Schizophr. Bull.* 40, 1285–1299.
- Ma, L., Tang, J., Wang, D., Zhang, W., Liu, W., Wang, D., Liu, X.H., Gong, W., Yao, Y.G., Chen, X., 2013. Evaluating risk loci for schizophrenia distilled from genome-wide association studies in Han Chinese from Central China. *Mol. Psychiatry* 18, 638–639.
- Malaspina, D., 2001. Paternal factors and schizophrenia risk: *de novo* mutations and imprinting. *Schizophr. Bull.* 27, 379–393.
- Malenka, R.C., Nicoll, R.A., 1993. NMDA-receptor-dependent synaptic plasticity: multiple forms and mechanisms. *Trends Neurosci.* 16, 521–517.
- McCarthy, M.I., Abecasis, G.R., Cardon, L.R., Goldstein, D.B., Little, J., Ioannidis, J.P., Hirschhorn, J.N., 2008. Genome-wide association studies for complex traits: consensus, uncertainty and challenges. *Nat. Rev. Genet.* 9, 356–369.
- McClellan, J., King, M.C., 2010. Genetic heterogeneity in human disease. *Cell* 141, 210–217.
- McClellan, J.M., Susser, E., King, M.C., 2007. Schizophrenia: a common disease caused by multiple rare alleles. *Br. J. Psychiatry* 190, 194–199.
- McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytsky, A., Garimella, K., Altshuler, D., Gabriel, S., Daly, M., DePristo, M.A., 2010. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* 20, 1297–1303.
- Michaelson, J.J., Shi, Y., Gujral, M., Zheng, H., Malhotra, D., Jin, X., Jian, M., Liu, G., Greer, D., Bhandari, A., Wu, W., Corominas, R., Peoples, A., Koren, A., Gore, A., Kang, S., Lin, G.N., Estabillo, J., Gadomski, T., Singh, B., Zhang, K., Akshoomoff, N., Corsello, C., McCarron, S., Iakoucheva, L.M., Li, Y., Wang, J., Sebat, J., 2012. Whole-genome sequencing in autism identifies hot spots for *de novo* germline mutation. *Cell* 151, 1431–1442.
- Missler, M., Hammer, R.E., Sudhof, T.C., 1998. Neurexophilin binding to alpha-neurexins: a single LNS domain functions as an independently folding ligand-binding unit. *J. Biol. Chem.* 273, 34716–34723.
- Montojo, J., Zuberi, K., Rodriguez, H., Bader, G.D., Morris, Q., 2014. GeneMANIA: fast gene network construction and function prediction for Cytoscape. *F1000Res.* 3, 153.
- Need, A.C., McEvoy, J.P., Gennarelli, M., Heinzen, E.L., Ge, D., Maia, J.M., Shianna, K.V., He, M., Cirulli, E.T., Gumbs, C.E., Zhao, Q., Campbell, C.R., Hong, L., Rosenquist, P., Putkonen, A., Halilainen, T., Repo-Tiihonen, E., Tiihonen, J., Levy, D.L., Meltzer, H.Y., Goldstein, D.B., 2012. Exome sequencing followed by large-scale genotyping suggests a limited role for moderately rare risk factors of strong effect in schizophrenia. *Am. J. Hum. Genet.* 91, 303–312.
- Ng, P.C., 2003. SIFT: predicting amino acid changes that affect protein function. *Nucleic Acids Res.* 31, 3812–3814.
- Nyegaard, M., Demontis, D., Foldager, L., Hedemand, A., Flint, T.J., Sorensen, K.M., Andersen, P.S., Nordentoft, M., Werge, T., Pedersen, C.B., Hougaard, D.M., Mortensen, P.B., Mors, O., Borglum, A.D., 2010. *CACNA1C* (rs1006737) is associated with schizophrenia. *Mol. Psychiatry* 15, 119–121.
- Park, D., Tosello-Trampont, A.C., Elliott, M.R., Lu, M., Haney, L.B., Ma, Z., Klibanov, A.L., Mandell, J.W., Ravichandran, K.S., 2007. *BAI1* is an engulfment receptor for apoptotic cells upstream of the ELMO/Dock180/Rac module. *Nature* 450, 430–434.
- Persico, A.M., D'Agruma, L., Maiorano, N., Totaro, A., Militerni, R., Bravaccio, C., Wassink, T.H., Schneider, C., Melmed, R., Trillo, S., Montecchi, F., Palermo, M.,

- Pascucci, T., Puglisi-Allegra, S., Reichelt, K.L., Conciatori, M., Marino, R., Quattrochi, C.C., Baldi, A., Zelante, L., Gasparini, P., Keller, F., Collaborative Linkage Study of Autism, 2001. *Reelin* gene alleles and haplotypes as a factor predisposing to autistic disorder. *Mol. Psychiatry* 6, 150–159.
- Petersen, B.S., Spehlmann, M.E., Raedler, A., Stade, B., Thomsen, I., Rabionet, R., Rosenstiel, P., Schreiber, S., Franke, A., 2014. Whole genome and exome sequencing of monozygotic twins discordant for Crohn's disease. *BMC Genomics* 15, 564.
- Rausch, T., Zichner, T., Schlattl, A., Stutz, A.M., Benes, V., Korbel, J.O., 2012. DELLY: structural variant discovery by integrated paired-end and split-read analysis. *Bioinformatics* 28, i333–i339.
- Ripke, S., O'Dushlaine, C., Chambert, K., Moran, J.L., Kahler, A.K., Akterin, S., Bergen, S.E., Collins, A.L., Crowley, J.J., Fromer, M., Kim, Y., Lee, S.H., Magnusson, P.K., Sanchez, N., Stahl, E.A., Williams, S., Wray, N.R., Xia, K., Bettella, F., Borglum, A.D., Bulik-Sullivan, B.K., Cormican, P., Craddock, N., de Leeuw, C., Durmishi, N., Gill, M., Golimbet, V., Hamshere, M.L., Holmans, P., Hougaard, D.M., Kendler, K.S., Lin, K., Morris, D.W., Mors, O., Mortensen, P.B., Neale, B.M., O'Neill, F.A., Owen, M.J., Milovancevic, M.P., Posthuma, D., Powell, J., Richards, A.L., Riley, B.P., Ruderfer, D., Rujescu, D., Sigurdsson, E., Silagadze, T., Smit, A.B., Stefansson, H., Steinberg, S., Suvisaari, J., Tosato, S., Verhage, M., Walters, J.T., , Multicenter Genetic Studies of Schizophrenia C, Levinson, D.F., Gejman, P.V., Kendler, K.S., Laurent, C., Mowry, B.J., O'Donovan, M.C., Owen, M.J., Pulver, A.E., Riley, B.P., Schwab, S.G., Wildenauer, D.B., Dudbridge, F., Holmans, P., Shi, J., Albus, M., Alexander, M., Campion, D., Cohen, D., Dikeos, D., Duan, J., Eichhammer, P., Godard, S., Hansen, M., Lerer, F.B., Liang, K.Y., Maier, W., Mallet, J., Nertney, D.A., Nestadt, G., Norton, N., O'Neill, F.A., Papadimitriou, G.N., Ribble, R., Sanders, A.R., Silverman, J.M., Walsh, D., Williams, N.M., Wormley, B., Psychosis Endophenotypes International C, Arranz, M.J., Bakker, S., Bender, S., Bramon, E., Collier, D., Crespo-Facorro, B., Hall, J., Iyegbe, C., Jablensky, A., Kahn, R.S., Kalaydjieva, L., Lawrie, S., Lewis, C.M., Lin, K., Linszen, D.H., Mata, I., McIntosh, A., Murray, R.M., Ophoff, R.A., Powell, J., Rujescu, D., Van Os, J., Walshe, M., Weisbrod, M., Wiersma, D., , Wellcome Trust Case Control C, Donnelly, P., Barroso, I., Blackwell, J.M., Bramon, E., Brown, M.A., Casas, J.P., Corvin, A.P., Deloukas, P., Duncanson, A., Jankowski, J., Markus, H.S., Mathew, C.G., Palmer, C.N., Plomin, R., Rautanen, A., Sawcer, S.J., Trembath, R.C., Viswanathan, A.C., Wood, N.W., Spencer, C.C., Band, G., Bellenguez, C., Freeman, C., Hellenthal, G., Giannoulatou, E., Pirinen, M., Pearson, R.D., Strange, A., Su, Z., Vukcevic, D., Donnelly, P., Langford, C., Hunt, S.E., Edkins, S., Gwilliam, R., Blackburn, H., Bumpstead, S.J., Dronov, S., Gillman, M., Gray, E., Hammond, N., Jayakumar, A., McCann, O.T., Liddle, J., Potter, S.C., Ravindrarajah, R., Ricketts, M., Tashakkori-Ghanbari, A., Waller, M.J., Weston, P., Widaa, S., Whittaker, P., Barroso, I., Deloukas, P., Mathew, C.G., Blackwell, J.M., Brown, M.A., Corvin, A.P., McCarthy, M.I., Spencer, C.C., Bramon, E., Corvin, A.P., O'Donovan, M.C., Stefansson, K., Scolnick, E., Purcell, S., McCarroll, S.A., Sklar, P., Hultman, C.M., Sullivan, P.F., 2013. Genome-wide association analysis identifies 13 new risk loci for schizophrenia. *Nat. Genet.* 45, 1150–1159.
- Rose, E.J., Morris, D.W., Hargreaves, A., Fahey, C., Greene, C., Garavan, H., Gill, M., Corvin, A., Donohoe, G., 2013. Neural effects of the *CSMD1* genome-wide associated schizophrenia risk variant rs10503253. *Am. J. Med. Genet. B. Neuropsychiatr. Genet.* 162, 530–537.
- Rossin, E.J., Lage, K., Raychaudhuri, S., Xavier, R.J., Tatar, D., Benita, Y., , International Inflammatory Bowel Disease Genetics C, Cotsapas, C., Daly, M.J., 2011. Proteins encoded in genomic regions associated with immune-mediated disease physically interact and suggest underlying biology. *PLoS Genet.* 7, e1001273.
- Roth, T.L., Lubin, F.D., Sodhi, M., Kleinman, J.E., 2009. Epigenetic mechanisms in schizophrenia. *Biochim. Biophys. Acta* 1790, 869–877.
- Rousou, P., Mitchell, A.C., Voloudakis, G., Fullard, J.F., Pothula, V.M., Tsang, J., Stahl, E.A., Georgakopoulos, A., Ruderfer, D.M., Charney, A., Okada, Y., Simonovich, K.A., Worthington, J., Padyukov, L., Klareskog, L., Gregersen, P.K., Plenge, R.M., Raychaudhuri, S., Fromer, M., Purcell, S.M., Brennan, K.J., Robakis, N.K., Schadt, E.E., Akbarian, S., Sklar, P., 2014. A role for noncoding variation in schizophrenia. *Cell Rep.* 9, 1417–1429.
- Saha, S., Chant, D., Welham, J., McGrath, J., 2005. A systematic review of the prevalence of schizophrenia. *PLoS Med.* 2, e141.
- Schizophrenia Psychiatric Genome-Wide Association Study Consortium, 2011. Genome-wide association study identifies five new schizophrenia loci. *Nat. Genet.* 43, 969–976.
- Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 511, 421–427.
- Schwab, S.G., Hoefgen, B., Hanses, C., Hassenbach, M.B., Albus, M., Lerer, B., Trixler, M., Maier, W., Wildenauer, D.B., 2005. Further evidence for association of variants in the *AKT1* gene with schizophrenia in a sample of European sib-pair families. *Biol. Psychiatry* 58, 446–450.
- Schwarz, J.M., Cooper, D.N., Schuelke, M.D.S., 2014. MutationTaster2: mutation prediction for the deep-sequencing age. *Nat. Med.* 21, 361–362.
- Sebat, J., Levy, D.L., McCarthy, S.E., 2009. Rare structural variants in schizophrenia: one disorder, multiple mutations; one mutation, multiple disorders. *Trends Genet.* 25, 528–535.
- Seripa, D., Matera, M.G., Franceschi, M., Daniele, A., Bizzarro, A., Rinaldi, M., Panza, F., Fazio, V.M., Gravina, C., D'Onofrio, G., Solfrizzi, V., Masullo, C., Pilotto, A., 2008. The *RELN* locus in Alzheimer's disease. *J. Alzheimers Dis.* 14, 335–344.
- Shi, L., 2013. Dock protein family in brain development and neurological disease. *Commun. Integr. Biol.* 6, e26839.
- Shi, Y., Li, Z., Xu, Q., Wang, T., Li, T., Shen, J., Zhang, F., Chen, J., Zhou, G., Ji, W., Li, B., Xu, Y., Liu, D., Wang, P., Yang, P., Liu, B., Sun, W., Wan, C., Qin, S., He, G., Steinberg, S., Cichon, S., Werge, T., Sigurdsson, E., Tosato, S., Palotie, A., Nothen, M.M., Rietschel, M., Ophoff, R.A., Collier, D.A., Rujescu, D., Clair, D.S., Stefansson, H., Stefansson, K., Ji, J., Wang, Q., Li, W., Zheng, L., Zhang, H., Feng, G., He, L., 2011. Common variants on 8p12 and 1q24.2 confer risk of schizophrenia. *Nat. Genet.* 43, 1224–1227.
- Shifman, S., Johannesson, M., Bronstein, M., Chen, S.X., Collier, D.A., Craddock, N.J., Kendler, K.S., Li, T., O'Donovan, M., O'Neill, F.A., Owen, M.J., Walsh, D., Weinberger, D.R., Sun, C., Flint, J., Darvasi, A., 2008. Genome-wide association identifies a common variant in the *reelin* gene that increases the risk of schizophrenia only in women. *PLoS Genet.* 4, e28.
- Shu, T., Ayala, R., Nguyen, M.D., Xie, Z., Gleeson, J.G., Tsai, L.H., 2004. Ndel1 operates in a common pathway with Lis1 and cytoplasmic dynein to regulate cortical neuronal positioning. *Neuron* 44, 263–277.
- Stefansson, H., Meyer-Lindenberg, A., Steinberg, S., Magnusdottir, B., Morgen, K., Arnarsdottir, S., Bjornsdottir, G., Walters, G.B., Jonsdottir, G.A., Doyle, O.M., Tost, H., Grimm, O., Kristjansdottir, S., Snorrason, H., Davidsdottir, S.R., Gudmundsson, L.J., Jonsson, G.F., Stefansdottir, B., Helgadottir, I., Haraldsson, M., Jonsdottir, B., Thygesen, J.H., Schwarz, A.J., Didriksen, M., Stensbol, T.B., Brammer, M., Kapur, S., Hallordsson, J.G., Hreidarsson, S., Saemundsen, E., Sigurdsson, E., Stefansson, K., 2014. CNVs conferring risk of autism or schizophrenia affect cognition in controls. *Nature* 505, 361–366.
- Straub, R.E., Lipska, B.K., Egan, M.F., Goldberg, T.E., Callicott, J.H., Mayhew, M.B., Vakkalanka, R.K., Kolachana, B.S., Kleinman, J.E., Weinberger, D.R., 2007. Allelic variation in *GAD1* (*GAD67*) is associated with schizophrenia and influences cortical function and gene expression. *Mol. Psychiatry* 12, 854–869.
- Sudmant, P.H., Rausch, T., Gardner, E.J., Handsaker, R.E., Abyzov, A., Huddleston, J., Zhang, Y., Ye, K., Jun, G., Hsi-Yang Fritz, M., Konkel, M.K., Malhotra, A., Stutz, A.M., Shi, X., Paolo Casale, F., Chen, J., Hormozdiari, F., Dayama, G., Chen, K., Malig, M., Chaisson, M.J., Walter, K., Meiers, S., Kashin, S., Garrison, E., Auton, A., Lam, H.Y., Jasmine Mu, X., Alkan, C., Antaki, D., Bae, T., Cerveira, E., Chines, P., Chong, Z., Clarke, L., Dal, E., Ding, L., Emery, S., Fan, X., Gujral, M., Kahveci, F., Kidd, J.M., Kong, Y., Lameijer, E.W., McCarthy, S., Flicek, P., Gibbs, R.A., Marth, G., Mason, C.E., Menelaou, A., Muzny, D.M., Nelson, B.J., Noor, A., Parrish, N.F., Pendleton, M., Quitadamo, A., Raeder, B., Schadt, E.E., Romanovitch, M., Schlattl, A., Sebra, R., Shabalina, A.A., Untergasser, A., Walker, J.A., Wang, M., Yu, F., Zhang, C., Zhang, J., Zheng-Bradley, X., Zhou, W., Zichner, T., Sebat, J., Batzer, M.A., McCarroll, S.A., , Genomes Project C, Mills, R.E., Gerstein, M.B., Bashir, A., Stegle, O., Devine, S.E., Lee, C., Eichler, E.E., Korbel, J.O., 2015. An integrated map of structural variation in 2,504 human genomes. *Nature* 526, 75–81.
- Sullivan, P.F., Daly, M.J., O'Donovan, M., 2012. Genetic architectures of psychiatric disorders: the emerging picture and its implications. *Nat. Rev. Genet.* 13, 537–551.
- Sullivan, P.F., Kendler, K.S., Neale, M.C., 2003. Schizophrenia as a complex trait: evidence from a meta-analysis of twin studies. *Arch. Gen. Psychiatry* 60, 1187–1192.
- Thomson, P.A., Parla, J.S., McRae, A.F., Kramer, M., Ramakrishnan, K., Yao, J., Soares, D.C., McCarthy, S., Morris, S.W., Cardone, L., Cass, S., Ghiban, E., Hennah, W., Evans, K.L., Rebolini, D., Millar, J.K., Harris, S.E., Starr, J.M., MacIntyre, D.J., Generation, S., McIntosh, A.M., Watson, J.D., Deary, I.J., Visscher, P.M., Blackwood, D.H., McCombie, W.R., Porteous, D.J., 2014. 708 Common and 2010 rare *DISC1* locus variants identified in 1542 subjects: analysis for association with psychiatric disorder and cognitive traits. *Mol. Psychiatry* 19, 668–675.
- Tissir, F., Goffinet, A.M., 2003. Reelin and brain development. *Nat. Rev. Neurosci.* 4, 496–505.
- Tsuboi, D., Kuroda, K., Tanaka, M., Namba, T., Iizuka, Y., Taya, S., Shinoda, T., Hikita, T., Muraoka, S., Iizuka, M., Nimura, A., Mizoguchi, A., Shiina, N., Sokabe, M., Okano, H., Mikoshiba, K., Kaibuchi, K., 2015. Disrupted-in-schizophrenia 1 regulates transport of *ITPR1* mRNA for synaptic plasticity. *Nat. Neurosci.* 18, 698–707.
- Wang, Q., Li, M., Yang, Z., Hu, X., Wu, H.M., Ni, P., Ren, H., Deng, W., Li, M., Ma, X., Guo, W., Zhao, L., Wang, Y., Xiang, B., Lei, W., Sham, P.C., Li, T., 2015. Increased co-expression of genes harboring the damaging *de novo* mutations in Chinese schizophrenic patients during prenatal development. *Sci. Rep.* 5, 18209.
- Ward, L.D., Kelis, M., 2012. Interpreting noncoding genetic variation in complex traits and human disease. *Nat. Biotechnol.* 30, 1095–1106.
- Winberg, M.L., Noordermeer, J.N., Tamagnone, L., Comoglio, P.M., Spriggs, M.K., Tessier-Lavigne, M., Goodman, C.S., 1998. Plexin A is a neuronal semaphorin receptor that controls axon guidance. *Cell* 95, 903–916.
- Wu, Q., Maniatis, T., 1999. A striking organization of a large family of human neural cadherin-like cell adhesion genes. *Cell* 97, 779–790.
- Wu, Y., Yao, Y.G., Luo, X.J., 2017. SZDB: a database for schizophrenia genetic research. *Schizophr. Bull.* 43, 459–471.
- Xiao, X., Chang, H., Li, M., 2017. Molecular mechanisms underlying noncoding risk variations in psychiatric genetic studies. *Mol. Psychiatry* 22, 497–511.
- Xu, B., Roos, J.L., Levy, S., van Rensburg, E.J., Gogos, J.A., Karayiorgou, M., 2008. Strong association of *de novo* copy number mutations with sporadic schizophrenia. *Nat. Genet.* 40, 880–885.
- Yamada, K., Nakamura, K., Minabe, Y., Iwayama-Shigeno, Y., Takao, H., Toyota, T., Hattori, E., Takei, N., Sekine, Y., Suzuki, K., Iwata, Y., Miyoshi, K., Honda, A., Baba, K., Katayama, T., Tohyama, M., Mori, N., Yoshikawa, T., 2004. Association analysis of *FEZ1* variants with schizophrenia in Japanese cohorts. *Biol.*

- Psychiatry 56, 683–690.
- Yang, T.P., Beazley, C., Montgomery, S.B., Dimas, A.S., Gutierrez-Arcelus, M., Stranger, B.E., Deloukas, P., Dermitzakis, E.T., 2010. Genevar: a database and Java application for the analysis and visualization of SNP-gene associations in eQTL studies. *Bioinformatics* 26, 2474–2476.
- Ye, K., Schulz, M.H., Long, Q., Apweiler, R., Ning, Z., 2009. Pindel: a pattern growth approach to detect break points of large deletions and medium sized insertions from paired-end short reads. *Bioinformatics* 25, 2865–2871.
- Yue, W.H., Wang, H.F., Sun, L.D., Tang, F.L., Liu, Z.H., Zhang, H.X., Li, W.Q., Zhang, Y.L., Zhang, Y., Ma, C.C., Du, B., Wang, L.F., Ren, Y.Q., Yang, Y.F., Hu, X.F., Wang, Y., Deng, W., Tan, L.W., Tan, Y.L., Chen, Q., Xu, G.M., Yang, G.G., Zuo, X.B., Yan, H., Ruan, Y.Y., Lu, T.L., Han, X., Ma, X.H., Wang, Y., Cai, L.W., Jin, C., Zhang, H.Y., Yan, J., Mi, W.F., Yin, X.Y., Ma, W.B., Liu, Q., Kang, L., Sun, W., Pan, C.X., Shuang, M., Yang, F.D., Wang, C.Y., Yang, J.L., Li, K.Q., Ma, X., Li, L.J., Yu, X., Li, Q.Z., Huang, X., Lv, L.X., Li, T., Zhao, G.P., Huang, W., Zhang, X.J., Zhang, D., 2011. Genome-wide association study identifies a susceptibility locus for schizophrenia in Han Chinese at 11p11.2. *Nat. Genet.* 43, 1228–1231.
- Yuen, R.K., Thiruvahindrapuram, B., Merico, D., Walker, S., Tammimies, K., Hoang, N., Chrysler, C., Nalpathamkalam, T., Pellecchia, G., Liu, Y., Gazzellone, M.J., D'Abate, L., Deneault, E., Howe, J.L., Liu, R.S., Thompson, A., Zarrei, M., Uddin, M., Marshall, C.R., Ring, R.H., Zwaigenbaum, L., Ray, P.N., Weksberg, R., Carter, M.T., Fernandez, B.A., Roberts, W., Szatmari, P., Scherer, S.W., 2015. Whole-genome sequencing of quartet families with autism spectrum disorder. *Nat. Med.* 21, 185–191.
- Zong, X.F., Hu, M.L., Li, Z.C., Cao, H.B., Chen, X.G., Tang, J.S., 2015. DNA methylation in schizophrenia: progress and challenges. *Sci. Bull.* 60, 149–155.
- Zuk, O., Schaffner, S.F., Samocha, K., Do, R., Hechter, E., Kathiresan, S., Daly, M.J., Neale, B.M., Sunyaev, S.R., Lander, E.S., 2014. Searching for missing heritability: designing rare variant association studies. *Proc. Natl. Acad. Sci. U. S. A.* 111, E455–E464.

Supplemental Figures

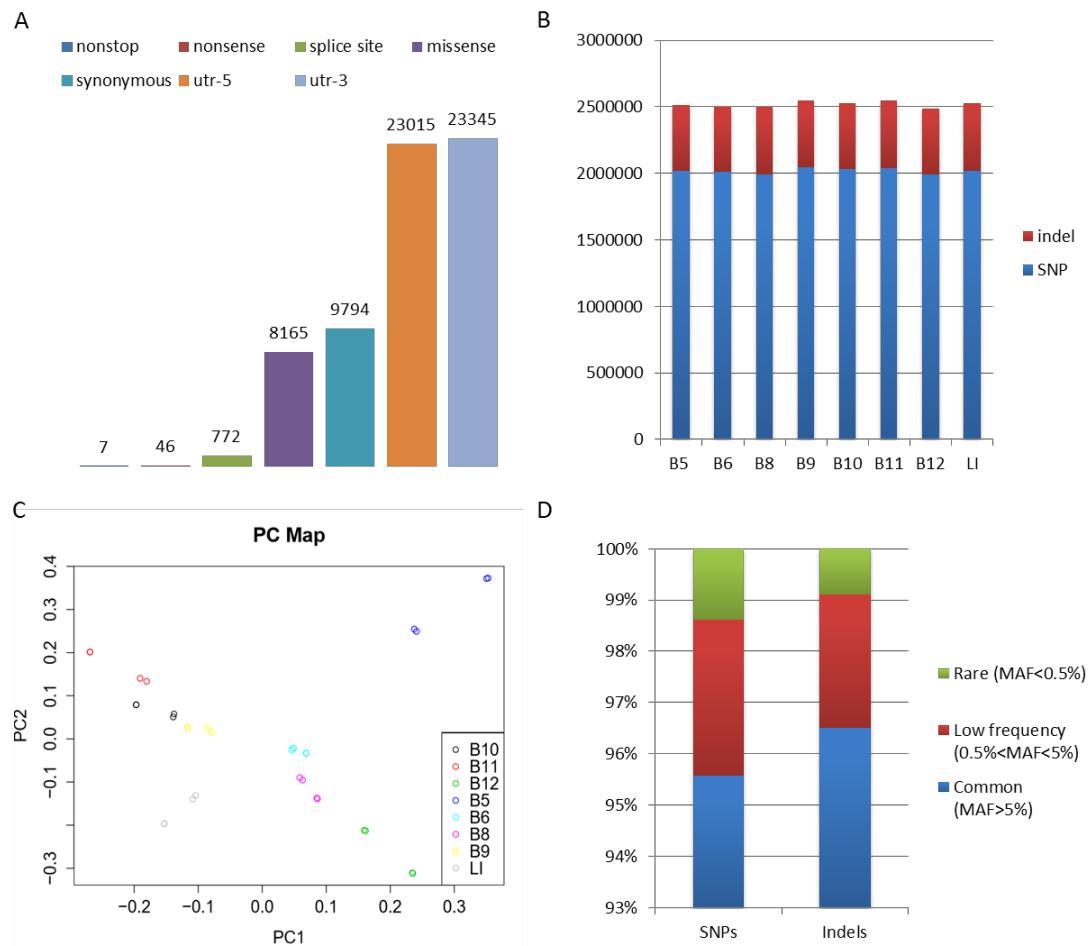


Fig. S1. The average distribution of SNVs in eight families (32 samples). The average number of various types of variations (A) and frequency (B) per individual; C: Principal component analysis (PCA) plot (the first and the second principal components) for 32 samples. The PCA was implemented by GCTA (Yang et al., 2011). D: The plot of shared SNVs in each family.

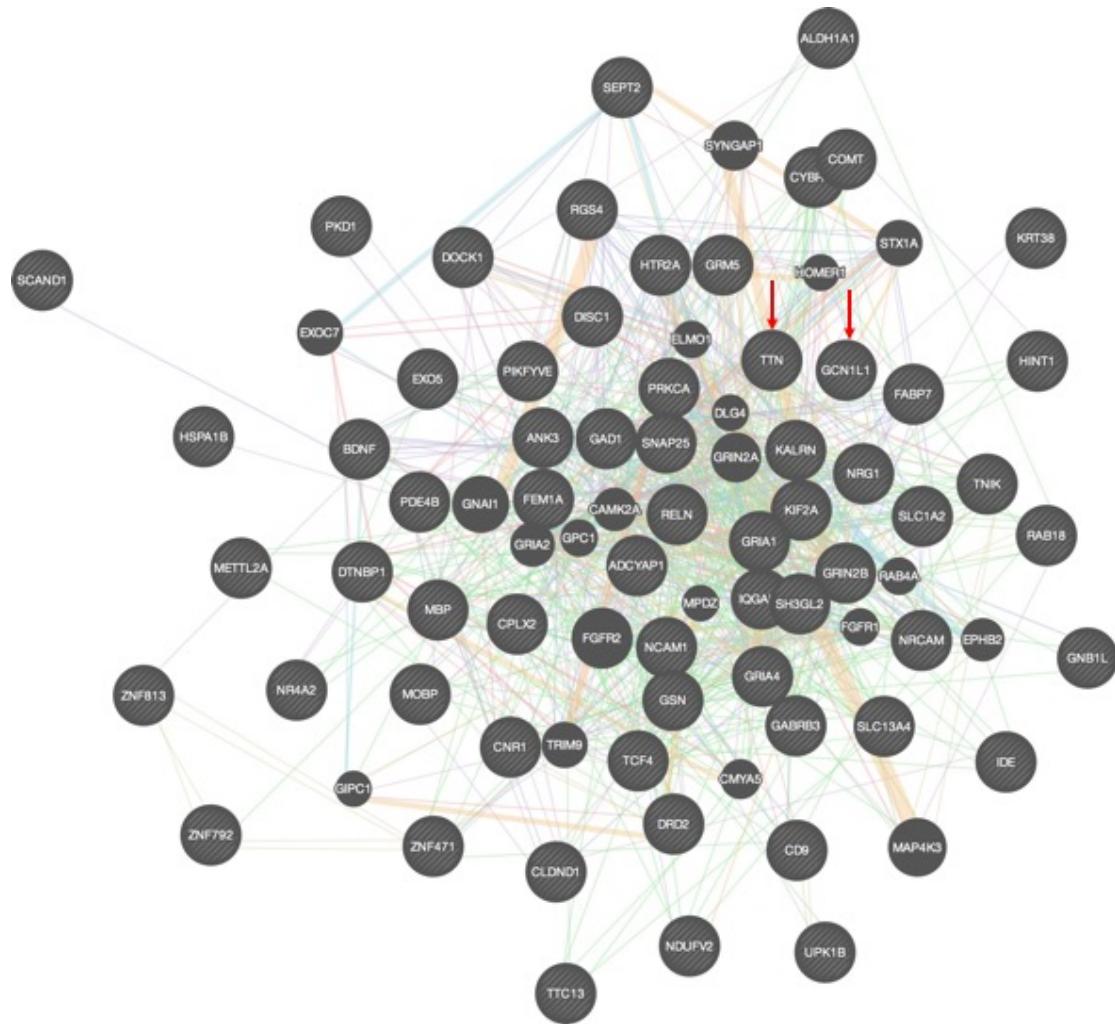


Fig. S2. Protein–protein interaction (PPI) network of 23 genes with DNM_s and 42 schizophrenia top genes. The network was based on the genes retrieved from GeneMANIA (Montojo et al., 2014) by inputting 23 genes hit by *de novo* mutations in the coding region, UTR region, 3'-downstream region or 5'-upstream region, and 42 SCZ top genes (identified by convergent functional genomics [CFG] score) of the SZDB data set (www.szdb.org). The *TTN* (titin; p.V24689I) and *GCN1L1* (GCN1 eIF2 alpha kinase activator homolog; p.S2506T) genes identified in family B5 were marked by arrows.

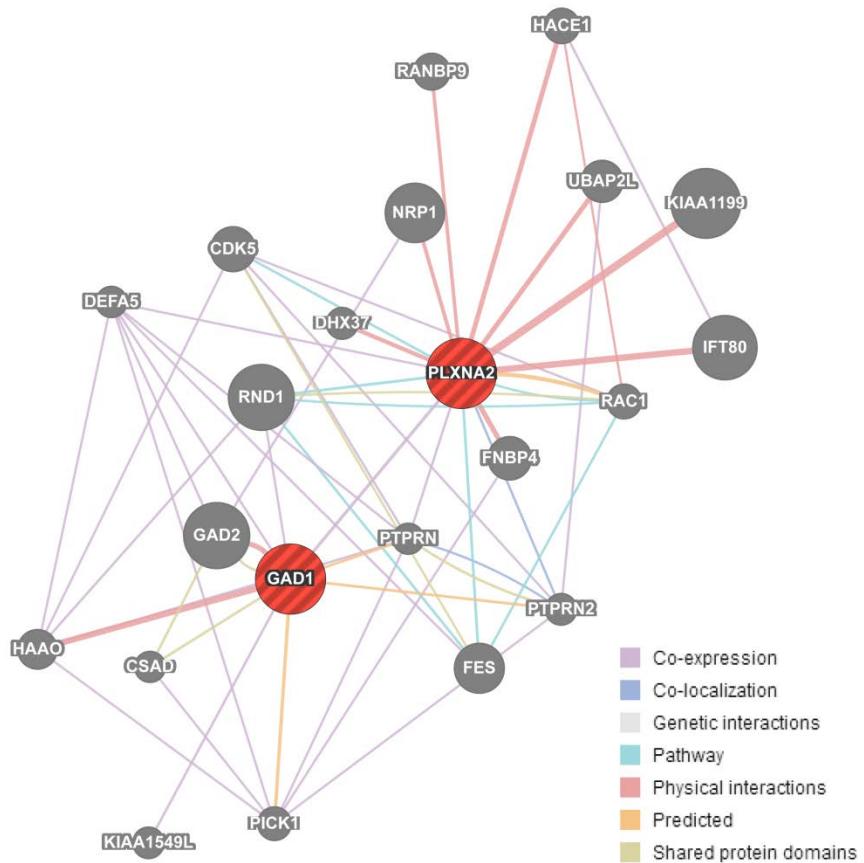


Fig. S3. GeneMANIA network showing potential interactions with GAD1 and PLXNA2. The network was based on the genes retrieved from GeneMANIA (Montojo et al., 2014) by inputting GAD1 and PLXNA2. GAD1 and PLXNA2 are marked with red color, and the other genes that interacted with GAD1 or PLXNA2 are marked with grey color.

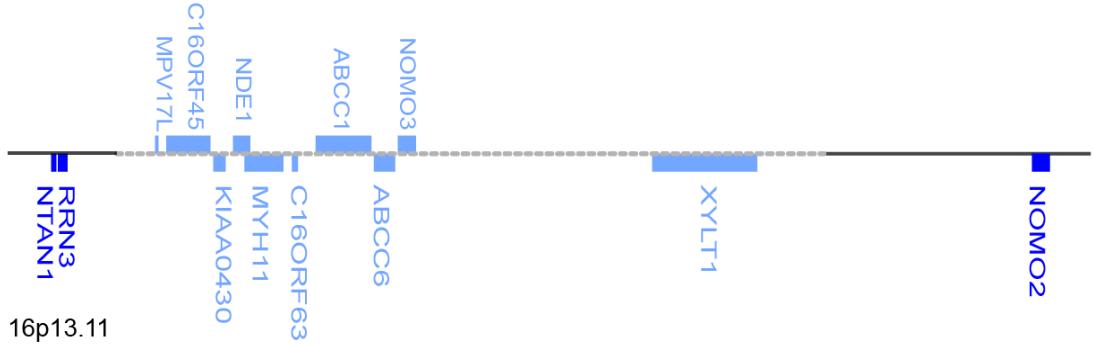


Fig. S4. Schematic profile of the large deletion (16p13.11; 2.9M) in proband of B11 family. The deletion contains 10 genes (*MPV17L*, *C16orf45*, *KIAA0430*, *NDE1*, *MYH11*, *C16orf63*, *ABCC1*, *ABCC6*, *NOMO3* and *XYLT1*) marked with lighter color and grey dotted line.

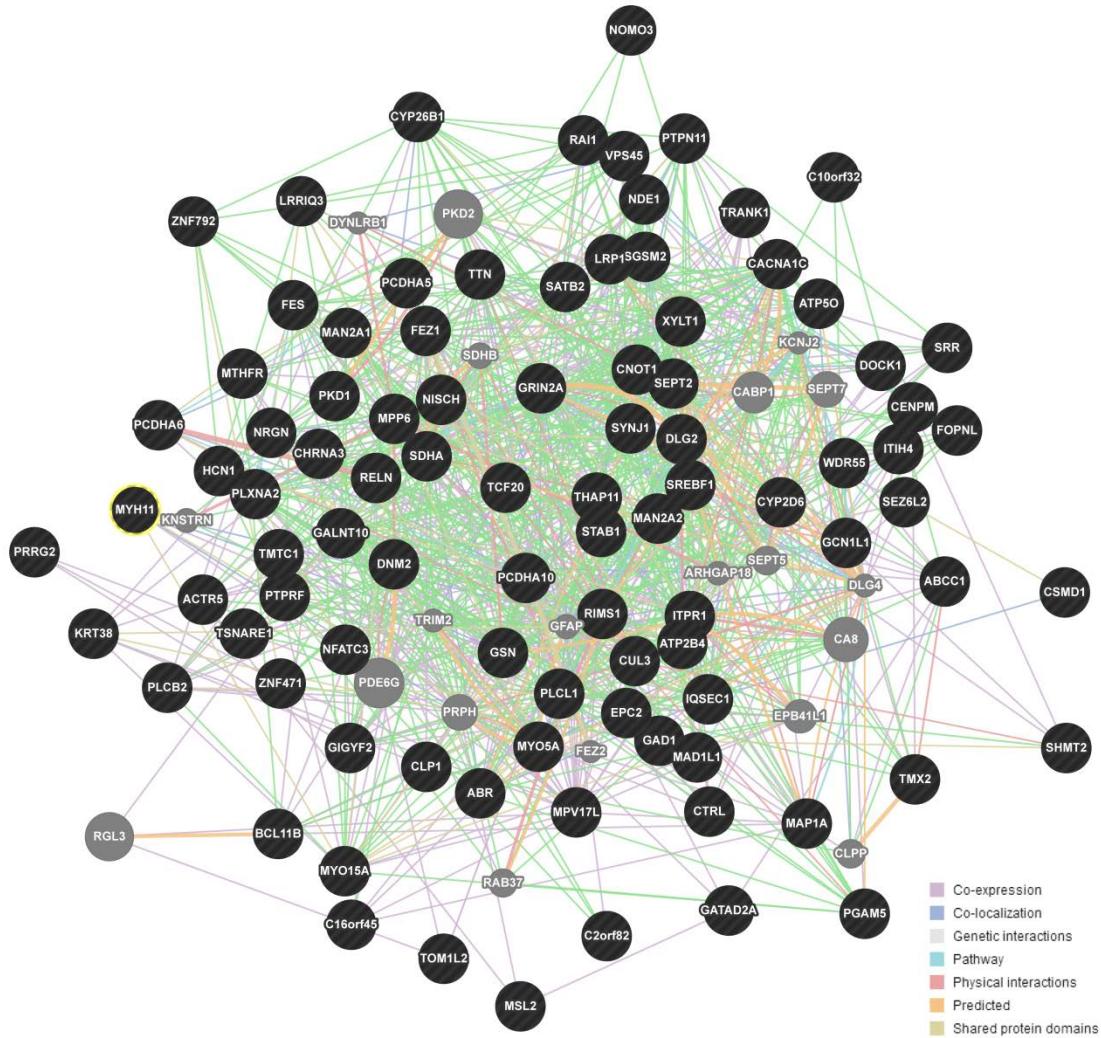


Fig. S5. GeneMANIA network categories. The network analysis was based on genes from the GeneMANIA (Montojo et al., 2014) by inputting genes harbored *de novo* missense mutations and inherited highly risky or rare damaging mutations (Tables 1, S10, S13 and S16). Inputted genes are marked with black color, and the other genes that interacted with the inputted genes are marked with grey color.

Supplemental Tables

Table S1. Demographical information and sequencing depth for each member of the 8 families with MZ twins

Family ID	Sample	Original sample ID	Sex ^a	Age	Kinship	Onset age	Duration of psychosis (year)	Average sequencing depth (X)
B5	B5-F	B51	M	45	Father	-	-	35.77
	B5-M	B52	F	47	Mother	-	-	31.91
	B5-T1	B53	M	24	Son	-	-	30.78
	B5-T2	B54	M	24	Son	23	1	31.4
B6	B6-F	B64	M	52	Father	-	-	30.81
	B6-M	B61	F	50	Mother	-	-	31.61
	B6-T1	B62	F	25	Daughter	-	-	32.31
	B6-T2	B63	F	25	Daughter	21	4	32.92
B8	B8-F	B81	M	44	Father	-	-	31.44
	B8-M	B82	F	42	Mother	-	-	31.45
	B8-T1	B83	M	22	Son	-	-	31.37
	B8-T2	B84	M	22	Son	12	10	33.27
B9	B9-F	B91	M	62	Father	-	-	33.92
	B9-M	B92	F	60	Mother	-	-	33.66
	B9-T1	B94	F	26	Daughter	-	-	30.13
	B9-T2	B93	F	26	Daughter	25	1	31.2
B10	B10-F	B101	M	50	Father	-	-	31.55
	B10-M	B102	F	50	Mother	-	-	30.91
	B10-T1	B103	F	24	Daughter	-	-	32.13
	B10-T2	B104	F	24	Daughter	21	3	31.22
B11	B11-F	B111	M	76	Father	-	-	32
	B11-M	B112	F	68	Mother	-	-	31.79
	B11-T1	B114	M	40	Son	-	-	31.11
	B11-T2	B113	M	40	Son	32	8	33.6
B12	B12-F	B121	M	46	Father	-	-	31.7
	B12-M	B122	F	46	Mother	-	-	31.58
	B12-T1	B123	M	19	Son	-	-	34.88
	B12-T2	B124	M	19	Son	17.5	1.5	32.29
LI	LI-F	LF	M	49	Father	-	-	28.46
	LI-M	LM	F	46	Mother	-	-	29.75
	LI-T1	LS	M	26	Son	-	-	31.51
	LI-T2	LJ	M	26	Son	24	2	28.3

The demographic information for each sample was described in our recent study (Li et al., 2016). ^a M - male; F - female.

Table S2. The number of shared SNVs in each family

Family ID	Shared SNPs	Shared indels
B5	2021376	495686
B6	2010400	489202
B8	1995282	504487
B9	2049020	497705
B10	2036478	491321
B11	2040822	505594
B12	1995286	494165
LI	2020038	504579
Mean	2021087.8	497842
SD	20190.49	6380.7

All variants in 32 individuals from the 8 families were scored relative to human reference genome (hg19; <http://genome.ucsc.edu/>).

Table S3. The number of DNMs in 8 pairs of MZ twins

Family ID	T1 unique SNPs ^a	T2 unique SNVs ^a	Shared SNVs	Shared indels
B5	0	1	54	1
B6	3	2	67	4
B8	5	7	49	1
B9	0	0	79	2
B10	0	1	66	1
B11	8	5	36	1
B12	2	3	58	1
LI	2	2	52	1
Total	20	21	461	12

^a T1 - twin 1; T2 - twin 2; Twin 2 of each family had schizophrenia.

Table S4. The distribution of DNMs in 8 pairs of MZ twins.

This table was listed as a separate Excel file, because the table is too big. The 8 DNMs that were not validated in this study were excluded.

Table S5. Validation of DNMs in 8 pairs of MZ twins

Sample	SNP	Allele	Gene	Predicted function	Validation	Primer Sequence (5'-3')	Sequencing region & Product length
B53 B54	chr2:179436794	C/T	<i>TTN</i>	p.V24689I	Validated	U: TCTTTATGCCAGGTTACAGC L: TACTATTCAGGGTCTCGC	chr2:179436604-179437094; 491bp
B53 B54	chr1:40982533	A/G	<i>EXO5</i>	3'-downstream	Validated	U: CAATAGCAGAACGCTAGAAGC L: CTTCTAAGATGGCTTGGGATT	chr1:40982362-40982602; 241bp
B53 B54	chr4:3769798	T/C	<i>ADRA2C</i>	3'-utr	False positive	U: TTCGTGCTCTGCTGGTCCC L: ACCCTGACCTCACAGCCACA	chr4:3769504-3770011; 508bp
B53 B54	chr12:120569035	C/G	<i>GCN1L1</i>	p.S2506T	Validated	U: CAAGTGCTGGTCCAAGGG L: GCCAAGATAGCAGGGAGG	chr12:120568834-120569295; 462bp
B53 B54	chr15:90200559	A/G	<i>KIF7</i>	5'-upstream	False positive	U: AAACCCATCACTCTAGGAGG L: ACATCCACCAGAAAGAGCAA	chr15:90200283-90200730; 448bp
B53 B54	chr17:39595049	G/T	<i>KRT38</i>	p.P265H	Validated	U: ACATCCTGGCGGTTGGTC L: TGAGGTGGGTCTGAGGGTC	chr17:39594971-39595341; 371bp
B53 B54	chr17:41382051	T/C	<i>LINC00854</i>	5'-upstream	Unknown	U: CAGTTTAGAAGCAGGAG L: TTTAGTACCGCATAGAGCC	
B103 B104	chr9:17578171	A/C	<i>SH3GL2</i>	5'-upstream	Validated	U: CGGATGGAGGACCCTT L: GGAGCAGAGCAGCACTAA	chr9:17577961-17578276; 316bp
B103 B104	chr3:118923152	T/C	<i>UPK1B</i>	3'-utr	Validated	U: CTTACATTGCCAAGTCAGA L: GGCATCAAGAGAACATCACC	chr3:118922923-118923330; 408bp
B103 B104	chr11:69923089	A/C	<i>ANO1</i>	5'-upstream	False positive	U: CCAGCCTGAGAACCAAAT L: ACAGCCGATGCTCAATAA	chr11:69922865-69923308; 444bp
B123 B124	chr6:56978734	T/G	RP11_203B9.4	3'-downstream	Validated	U: CATGTTAGCCAGGATGCTC L: TTCTTGAACAATTCACTCTCG	chr6:56978620-56978821; 202bp
B123 B124	chr2:172411647	G/C	<i>CYBRD1</i>	3'-utr	Validated	U: TGCTTCTCCTATTAGCCATAT L: ATTGTCTCCCAGAATTAACAG	chr2:172411394-172411722; 329bp

B123	B124	chr4:80994100	C/A	<i>ANTXR2</i>	5'-utr	False positive	U: GCCGGAACCTTGACGAATC L: CGGGAGGAAAGTTCCGGAGT	chr4:80993933-80994251; 319bp
B123	B124	chr17:60527119	A/G	<i>METTL2A</i>	3'-utr	Unknown	U: CTGGGCAACAAAGCAAGACTCT L: CTCGATCTCCTGACCTCGTGA U: TGATCGTGGTCTTCGCTTA L: GACGGAGTCTTCCTCTGTTGC	
B123	B124	chr19:35449194	T/C	<i>ZNF792</i>	p.N522S	Validated	U: CAGGCCCTGATAAGGTCTG L: CACGGGTGTTGAGTGTG	chr19:35448916-35449411; 496bp
B84		chr17:41382052	T/C	<i>LINC00854</i>	5'-upstream	Unknown	U: AGAGGGTCCCATTGAACT L: TGCTCTTAGTACCGCATAG	
B83	B84	chr2:209224680	C/T	<i>PIKFYVE</i>	3'-downstream	Validated	U: GGATAGAGCGTACCTGA L: GGCCTCTCTTACCTGT	chr2:209224567-209224976; 410bp
B83	B84	chr8:21966891	C/A	<i>NUDT18</i>	5'-utr	False positive	U: TCACCTGCTCGCTGAGGAAC L: ACTGACAGGCAGGCAGAAC	chr8:21966646-21967128; 483bp
B83	B84	chr20:9966603	G/A	<i>LOC101929371</i>	5'-upstream	Validated	U: CCATTACACTGGACTGGCAC L: AAATGTGGGCAGGAATAAGGA	chr20:9966366-9966703; 338bp
B83	B84	chr19:4796824	G/A	<i>FEMIA</i>	3'-downstream	Validated	U: AACGTCCGGTTAACCTCG L: AACCCACTCCTTGATGCA	chr19:4796713-4796954; 242bp
B93	B94	chr10:94297199	C/A	<i>IDE</i>	synonymous	Validated	U: TACCAATAGCTTACGAGGGT L: GAATAATCCAGCCATCAAGAG	chr10:94296973-94297280; 308bp
B93	B94	chr16:2147410	G/A	<i>PKD1</i>	p.R3438W	Validated	U: TGTGGGTGTCTGGGTAGG L: GGGAACGCTCAGTTGGC	chr16:2147174-2147480; 307bp
B93	B94	chr7:135414085	A/G	<i>SLC13A4</i>	5'-upstream	Validated	U: GAGCAGAGTCAAAGGGTTA L: AAATGAGGCCAGAATGTAG	chr7:135413881-135414168; 288bp
B93	B94	chr19:57037004	C/G	<i>ZNF471</i>	p.A523G	Validated	U: AAACCGTATGAATGCAAGGA L: ATTGGAAGTTGGCTGAAGG	chr19:57036805-57037107; 303bp

LJ LS	chr12:88176981	C/A	<i>MKRN9P</i>	non-coding	False positive	U: AGCAGACCAGTTCACACGC L: GAAATACAAGGAGGCAATGAGC	chr12:88176864-88177267; 404bp
LJ LS	chr7:31375247	C/G	<i>NEUROD6</i>	3'-downstream	False positive	U: CCATTGCATTTTGCATCC L: TCAGAACATCTGGTTCAGAC	chr7:31375077-31375366; 290bp
B62 B63	chr15:90929885	T/C	<i>IQGAPI</i>	5'-upstream	Validated	U: GGAGTCAGGCTATGATTGGC L: CCTCTCCGTTTATTCTTG	chr15:90929609-90930097; 489bp
B62 B63	chr2:242283282	A/G	<i>SEPT</i>	p.N271S	Validated	U: TAATGTTCTCGTGTTGCCAGTG L: ACTGAGGGCGGCAATGAA	chr2:242283093-242283431; 339bp
B62 B63	chr4:10042236	C/T	<i>SLC2A9</i>	5'-upstream	False positive	U: ATCATCGGGTAATGCCAA L: AAGCTGACCCCTCCAATGTT	chr4:10041979-10042430; 452bp
B62 B63	chr1:231116417	A/G	<i>TTCI3</i>	5'-upstream	Validated	U: TCTGCCTCCCAAGTTCAAGCAA L: TGCGGGTGGGAATGTGACTAAA	chr1:231116166-231116682; 517bp
B63	chr18:55144502	A/T	<i>ONECUT2</i>	3'-utr	False positive	U: CCTTCCTGGAGCGAGTTA L: GAGCGCAAGTGAAGTGA	chr18:55144240-55144714; 475bp
B113 B114	chr3:98242563	T/C	<i>CLDND1</i>	5'-upstream	Validated	U: CTGGCAGACAGTATCCATCAA L: TAAGATGTCCTGCTTCCTGT	chr3:98242291-98242659; 369bp
B113 B114	chr3:13974618	C/G	<i>FGD5P1</i>	non-coding	False positive	U: CAAAATTTCCCTTGTGGCG L: GGAGTGGAGGGCAAGGGT	chr3:13974381-13974894; 514bp
B113 B114	chr19:53997821	C/T	<i>ZNF813</i>	3'-downstream	Validated	U: AGGCACCCAGAAGGATAAG L: GAGTTTCGCTCTTGTGGACC	chr19:53997571-53997891; 321bp
B84	chr13:45830708	C/T	<i>GTF2F2</i>	intronic	Validated	U: ACCCCTGGTGAATAAAGTAAGT L: GTTCGTTTATGTTCTCAGTCT	chr13:45830487-45830883; 397bp
B84	chr2:167878685	A/G	<i>XIRP2</i>	intronic	Validated	U: TCAGATTTATTTCTGTATTCACC L: ATTATGCAATTGATATGTTCTGAGA	chr2:167878435-167878940; 506bp
B84	chr6:149959476	T/A	<i>KATNA1</i>	intronic	Validated	U: GTCTTGCTATGTTGTAGGCT L: CTCTGCGATGGTCTATTATCA	chr6:149959302-149959615; 314bp

Table S6. The genomic locations of DNMs in 8 pairs of MZ twins.

Genomic region	Number
CDS (missense)	7
CDS (synonymous)	1
3'-UTR	3
5'-UTR	0
3'-upstream	5
5'-upstream	9
Splice site	1
Intronic	223
Intergenic	255

Table S7. The regulatory elements hit by *de novo* SNVs in 8 pairs of MZ twins.

This table was listed as a separate Excel file, because the table is too big.

Table S8. Gene-based associations with schizophrenia

Gene symbol	Transcript ID	Position	NVAR^a	P^b	I^c	DESC^d
<i>KCNQ2</i>	ENST00000360480	chr20:62038277..62103437	564	0.000999	0.001	3245/2910
<i>LIMK2</i>	ENST00000331728	chr22:31608618..31674200	235	0.000999	0.001	1256/1054
<i>COPS4</i>	ENST00000503682	chr4:83956665..83995802	146	0.000999	0.001	712/588
<i>PHYKPL</i>	ENST00000308158	chr5:177639942..177659519	112	0.000999	0.001	658/475
<i>AIF1L</i>	ENST00000372298	chr9:133972131..133996503	102	0.000999	0.001	622/420
<i>ALKBH4</i>	ENST00000292566	chr7:102098010..102104491	34	0.000999	0.002	232/172
<i>POLD2</i>	ENST00000223361	chr7:44154865..44160926	18	0.001998	0.001	109/83
<i>SNAP47</i>	ENST00000366759	chr1:227923986..227968107	132	0.001998	0.001	749/579
<i>LTV1</i>	ENST00000367576	chr6:144164990..144182969	62	0.001998	0.001	286/200
<i>HCK</i>	ENST00000375852	chr20:30640592..30688137	121	0.001998	0.001	861/573
<i>CCDC125</i>	ENST00000396499	chr5:68579047..68616331	171	0.001998	0.001	1182/833
<i>KRT7</i>	ENST00000543899	chr12:52627215..52661714	143	0.001998	0.001	892/781
<i>BATF3</i>	ENST00000243440	chr1:212861303..212873096	60	0.002997	0.001	284/218
<i>EPHA1</i>	ENST00000275815	chr7:143088823..143105830	53	0.002997	0.001	268/218
<i>RGL4</i>	ENST00000382833	chr22:24034288..24040735	10	0.002997	0.001	104/74
<i>MET</i>	ENST00000397752	chr7:116339672..116436110	208	0.002997	0.001	1048/869
<i>EPC2</i>	ENST00000258484	chr2:149402806..149543718	523	0.003996	0.001	3275/2425
<i>ATP5C1</i>	ENST00000335698	chr10:7830319..7849341	69	0.003996	0.001	475/352
<i>ANXA1</i>	ENST00000376911	chr9:75773632..75784327	27	0.003996	0.001	188/122
<i>PSMG1</i>	ENST00000380900	chr21:40547848..40555160	33	0.003996	0.001	224/180
<i>TMEM106B</i>	ENST00000396667	chr7:12254545..12271452	146	0.003996	0.001	1487/937
<i>C5orf45</i>	ENST00000403396	chr5:179264564..179285752	66	0.003996	0.001	474/333
<i>C11orf65</i>	ENST00000525729	chr11:108181511..108332126	489	0.003996	0.001	3693/2618
<i>BPI</i>	ENST00000262865	chr20:36932660..36964414	178	0.004995	0.001	1201/1019
<i>TAPBPL</i>	ENST00000266556	chr12:6561603..6571298	38	0.004995	0.001	321/217
<i>C14orf182</i>	ENST00000399206	chr14:50459315..50472405	45	0.004995	0.001	386/240
<i>FEZF1</i>	ENST00000427185	chr7:121942621..121943610	11	0.004995	0.001	54/33
<i>ATM</i>	ENST00000452508	chr11:108098459..108235433	359	0.004995	0.001	2758/1947
<i>SQSTM1</i>	ENST00000510187	chr5:179248642..179263515	75	0.004995	0.001	599/396
<i>HEATR5A</i>	ENST00000543095	chr14:31763142..31871390	427	0.004995	0.001	2069/1567
<i>LRWD1</i>	ENST00000292616	chr7:102105852..102113256	50	0.005994	0.001	404/306
<i>PLA2G4E</i>	ENST00000399518	chr15:42276176..42342824	378	0.005994	0.001	2229/1658
<i>VAMP1</i>	ENST00000400911	chr12:6572048..6579565	43	0.005994	0.001	256/181
<i>C12orf28</i>	ENST00000547771	chr12:70220331..70345690	485	0.005994	0.001	2456/2052
<i>DEGS1</i>	ENST00000323699	chr1:224371225..224379317	31	0.006993	0.001	172/142
<i>KRT81</i>	ENST00000327741	chr12:52680766..52685213	23	0.006993	0.002	136/107
<i>C4orf34</i>	ENST00000511809	chr4:39554679..39606533	227	0.006993	0.001	1320/1042
<i>THAP9</i>	ENST00000536314	chr4:83822334..83839836	64	0.006993	0.002	374/276
<i>GDA</i>	ENST00000238018	chr9:74764627..74865422	279	0.007992	0.001	1801/1356
<i>RIN1</i>	ENST00000311320	chr11:66099987..66103046	14	0.007992	0.001	85/64
<i>SMAD5</i>	ENST00000545620	chr5:135489891..135513085	63	0.007992	0.002	671/438

<i>CHRNA10</i>	ENST00000250699	chr11:3687985..3692121	18	0.008991	0.001	216/147
<i>SPC25</i>	ENST00000282074	chr2:169728069..169745594	78	0.008991	0.001	464/375
<i>PEX11A</i>	ENST00000300056	chr15:90226947..90233145	17	0.008991	0.001	200/114
<i>CTC1</i>	ENST00000315684	chr17:8132763..8150923	55	0.008991	0.003	454/297
<i>INSR</i>	ENST00000341500	chr19:7117426..7293898	1310	0.008991	0.001	6621/5679
<i>UBE2D3</i>	ENST00000357194	chr4:103719434..103789810	223	0.008991	0.001	2192/1542
<i>FXYD5</i>	ENST00000423817	chr19:35647174..35660508	71	0.008991	0.001	412/302
<i>SYT8</i>	ENST00000535046	chr11:1853107..1857270	20	0.008991	0.001	128/101
<i>SLC39A2</i>	ENST00000298681	chr14:21467913..21469268	6	0.00999	0.001	68/48
<i>SP3</i>	ENST00000310015	chr2:174775562..174829798	181	0.00999	0.001	860/563
<i>TMPRSS11F</i>	ENST00000356291	chr4:68919659..68995529	313	0.00999	0.001	2174/1881
<i>CHI3L2</i>	ENST00000369748	chr1:111770423..111784816	79	0.00999	0.001	551/460
<i>CDKN2AIP</i>	ENST00000504169	chr4:184366145..184367749	8	0.00999	0.001	46/32
<i>CCDC149</i>	ENST00000504487	chr4:24810094..24914338	359	0.00999	0.001	2029/1650

^a NVAR refers to the number of variants in all 32 individuals from the 8 families when compared to the reference genome (hg19). ^b P-value is based on permutations, the empirical significance.

^c I-value indicates the proportion of null replicates for which the best test statistic was tied. ^d DESC contains the number of variants in cases/controls.

Table S9. The polygenic risk score and summary of results in the comparison of probands and healthy parents

Family ID	Father	Mother	Mean ^a	Proband	Proband vs mother increase (%) ^b	Proband vs mean increase (%) ^c
B5	5.1847	5.05077	5.117735	5.42641	7.44	6.03
B6	6.05225	5.41625	5.73425	5.57301	2.89	-2.81
B8	5.30071	5.3839	5.342305	5.90051	9.60	10.45
B9	6.02009	5.61828	5.819185	6.52813	16.19	12.18
B10	5.53655	4.69908	5.117815	5.41896	15.32	5.88
B11	5.16369	5.95777	5.56073	5.58196	-6.31	0.386
B12	6.15814	5.31027	5.734205	6.0097	13.17	4.80
LI	5.75368	4.65429	5.203985	5.22316	12.22	0.37
p-value	0.3528	0.0083	0.0211	-	-	-
R ²	0.02164	0.5834	0.4678	-	-	-

^aThe mean score is calculated as the average of mother's score and father's score. ^bThe increase of risk score is calculated as (the proband score – mother's score)/mother's score x 100%. ^cThe increase of risk score is calculated as (the proband score – the mean score of mother and father)/mean score x 100%.

Table S10. The genotypes of the SCZ-risk alleles in MZ twins

Family ID	dbSNP ID	MAF ^a	Genotype F ^b	Genotype M ^b	Genotype T ^b	Gene	Transcript ID	Function	<i>p</i> -value ^c
B8	rs2007044	G=0.4964	A/G	A/G	G/G	<i>CACNA1C</i>	NM_000719	intron	3.22E-18
B8	rs2973155	T=0.3728	T/C	T/C	C/C	Intergenic			1.11E-10
B8	rs6434928	G=0.2821	G/A	G/A	G/G	Intergenic			2.06E-11
B8	rs1023500	C=0.2969	T/C	T/C	T/T	<i>CENPM</i>	NM_001002876	intron	3.43E-08
B10	rs1023500	C=0.2969	T/C	T/C	T/T	<i>CENPM</i>	NM_001002876	intron	3.43E-08
B10	rs10503253	A=0.2270	C/A	C/A	A/A	<i>CSMD1</i>	NM_033225	intron	1.06E-08
B10	rs397805810	NA	A/ACT	A/ACT	ACT/ACT	<i>MADIL1</i>	NM_001013836.1.	intron	8.2E-15
B10	rs59979824	A=0.267	C/A	C/A	C/C	<i>AC013401.4</i>	ENST00000434525	intron	8.41E-09
B10	rs8042374	G=0.4495	A/G	A/G	A/A	<i>CHRNA3</i>	CCDS10305.1	intron	2.44E-13
B10	rs1501357	C=0.3992	T/C	T/C	C/C	<i>HCN1</i>	CCDS3952.1	intron	5.05E-09
B10	rs2535627	T=0.4665	T/C	T/C	T/T	<i>ITIH4</i>	NM_001166449	3downstream	4.26E-11
B5	rs2535627	T=0.4665	T/C	T/C	T/T	<i>ITIH4</i>	NM_001166449	3downstream	4.26E-11
B5	rs1501357	C=0.3992	T/C	T/C	C/C	<i>HCN1</i>	CCDS3952.1	intron	5.05E-09
B5	rs11139497	T=0.4467	T/A	T/A	A/A	<i>RP11-388B24.3</i>	ENST00000417796	intron	3.61E-09
B5	rs2905426	T=0.4097	G/T	G/T	G/G	Intergenic			3.63E-10
B5	rs4702	G=0.3538	G/A	G/A	G/G	<i>FES</i>	NM_001143783	5upstream	8.30E-14
B5	rs55661361	A=0.4058	G/A	G/A	G/G	<i>NRGN</i>	NM_001126181	intron	2.80E-12
B5	rs6704768	G=0.4619	G/A	G/A	G/G	<i>GIGYF2</i>	NM_001103146	intron	2.32E-12
B12	rs11685299	A=0.2534	C/A	C/A	C/C	<i>CUL3</i>	NM_001257197.1	intron	1.12E-08
B12	rs5825114	G=0.3948	GA/G	GA/G	GA/GA	Intergenic			8.03E-11
B12	rs6704641	G=0.3327	G/A	G/A	A/A	<i>SATB2</i>	CCDS2327.1	intron	8.33E-09
B6	rs6704641	G=0.3327	G/A	G/A	A/A	<i>SATB2</i>	CCDS2327.1	intron	8.33E-09

B6	rs12148337	C=0.4331	T/C	T/C	T/T	Intergenic			1.79E-08
B6	rs56873913	G=0.3472	T/G	T/G	T/T	<i>PRRG2</i>	NM_000951	intron	4.69E-08
B6	rs679087	A=0.1877	C/A	C/A	C/C	<i>TMTC1</i>	NM_001193451	intron	3.91E-08
B6	rs9420	A=0.247	A/G	A/G	A/A	<i>TMX2</i>	NM_001144012	3downstream	2.24E-09
B6	rs11682175	C=0.3508	T/C	T/C	C/C	<i>SNORD78</i>	ENST00000390866	intron	1.47E-11
B9	rs11682175	C=0.3508	T/C	T/C	C/C	<i>SNORD78</i>	ENST00000390866	intron	1.47E-11
B9	rs2909457	G=0.3117	G/A	G/A	G/G	Intergenic			4.62E-08
B9	rs397805810	NA	A/ACT	A/ACT	ACT/ACT	<i>MADIL1</i>	NM_001013836.1	intron	8.2E-15
B9	rs59979824	A=0.267	C/A	C/A	C/C	<i>AC013401.4</i>	ENST00000434525	intron	8.41E-09
LI	rs11682175	C=0.3508	T/C	T/C	C/C	<i>SNORD78</i>	ENST00000390866	intron	1.47E-11
LI	rs11210892	A=0.4455	G/A	G/A	G/G	Intergenic			3.39E-10
LI	rs2973155	T=0.3728	T/C	T/C	C/C	Intergenic			1.11E-10
LI	rs4523957	G=0.4770	G/T	G/T	T/T	<i>SRR</i>	NM_021947	intron	2.86E-10
B11	rs11191419	A=0.3906	T/A	T/A	T/T	<i>C10orf32</i>	NM_001136200	5upstream	6.20E-19
B11	rs1498232	C=0.4667	T/C	T/C	T/T	<i>AL645944.1</i>	ENST00000410855	intron	2.86E-09
B11	rs4129585	A=0.2388	A/C	A/C	A/A	<i>TSNARE1</i>	CCDS6384.1	intron	1.74E-15

^aThe minor allele frequency (MAF) from the 1000 Genomes Project (Sudmant et al., 2015). ^b F-father; M-mother; T-twins. ^c p-value from the original paper (Schizophrenia Working Group of the Psychiatric Genomics, 2014).

Table S11. The number of rare inherited loss-of-function and damaging missense variants in 8 families with MZ twins

Sample	Damaging missense ^a		Frameshift ^b		Nonsense ^b		Total	
	MAF<0.05 ^c	MAF<0.01 ^c	MAF<0.05	MAF<0.01	MAF<0.05	MAF<0.01	MAF<0.05	MAF<0.01
B10_father	322	236	48	44	7	6	377	286
B10_mother	273	199	49	47	10	6	332	252
B11_father	305	222	47	41	13	12	365	275
B11_mother	320	225	52	48	10	9	382	282
B12_father	325	226	44	42	13	11	382	279
B12_mother	255	192	50	47	7	5	312	244
B5_father	350	255	48	45	16	12	414	312
B5_mother	324	241	56	55	11	10	391	306
B6_mother	329	244	44	40	8	7	381	291
B6_father	357	263	47	43	7	4	411	310
B8_father	353	258	57	53	7	5	417	316
B8_mother	366	270	42	41	6	6	414	317
B9_father	351	250	42	40	15	12	408	302
B9_mother	328	229	48	45	6	5	382	279
LI_father	347	258	47	45	14	10	408	313
LI_mother	321	240	56	51	15	13	392	304
Average	326.63	238.00	48.56	45.44	10.31	8.31	385.50	291.75
B10_twin	357	264	51	47	9	6	417	317
B11_twin	316	229	53	49	14	14	383	292
B12_twin	316	222	49	48	10	7	375	277
B5_twin	352	265	56	54	18	14	426	333
B6_twin	343	261	39	37	5	3	387	301
B8_twin	359	256	54	53	6	4	419	313
B9_twin	320	229	42	40	12	11	374	280
LI_twin	332	235	51	48	11	10	394	293
Average	336.88	245.13	49.38	47.00	10.63	8.63	396.88	300.75
p-value^d								
twin vs mother	0.1034	0.1316	0.9064	0.8957	0.3358	0.3419	0.0329	0.1017
twin vs father	0.7992	0.8855	0.3483	0.1418	0.3042	0.5983	0.9209	0.82
twin vs parents	0.2644	0.3365	0.5715	0.2663	0.7429	0.7173	0.262	0.2598

^a Missense variants are rated as damaging when at least two of five prediction algorithms (SIFT (Ng, 2003; Kumar et al., 2009), PolyPhen2 HumDiv, PolyPhen2 HumVar (Adzhubei et al., 2010, 2013), LRT (Chun and Fay, 2009) and MutationTaster (Schwarz JM et al., 2014)) suggesting a potential deleterious effect.

^b Frameshift and nonsense variants are grouped as loss-of-function variants. ^c MAF- minor allele frequency. The cut-off of the MAF was based on the 1000 Genomes Project (Sudmant et al., 2015). ^d p-value of the Student's t-test.

Table S12. The rare damaging variants in *LAMC1*

Family ID	Mutant	Type ^b	MAF	SIFT ^a	Polyphen2	Polyphen2	LRT ^d	MutationTaster ^e
				HDIV ^b	HVAR ^c			
B5	p.G170R	PI	NA	D	D	D	D	D
B10	p.D325N	PI	NA	D	D	D	D	D
B8	p.F386Y	PI	0.0002	T	B	B	D	D
LI	p.D532E	PI	0.003395	T	B	B	D	D
B10	p.R557W	MI	0.005391	T	P	B	D	D
B6	p.R557W	PI	0.005391	T	P	B	D	D

We used five programs to predict the potential deleterious effect of the variant. ^a SIFT - D: Deleterious (sift <= 0.05); T: tolerated (sift > 0.05). ^b PolyPhen 2 HDIV - D: Probably damaging (>= 0.957), P: possibly damaging (0.453 ~ 0.956); B: benign (<= 0.452). ^c PolyPhen 2 HVar - D: Probably damaging (>= 0.909), P: possibly damaging (0.447 ~ 0.909); B: benign (<= 0.446). ^d LRT - D: Deleterious; N: Neutral; U: Unknown. ^e MutationTaster - A: disease causing automatic; D: disease causing; N: polymorphism; P: polymorphism automatic..

Table S13. Summary of the rare inherited loss-of-function or damaging variants in SCZ-associated genes in 8 pairs of MZ twins

Family ID	Gene	Mutant	RVIS (%ile) ^a	Type ^b	Class ^c	1000genome MAF	ExAC_EAS MAF
B11	RELN	p.M927V	-2.15(1.46)	MI	I	0.0002	0.00155
B6	MTHFR	p.R46W	0.09(60.71)	MI	I	0.000399	0.00242
B5	PLXNA2	p.P618H	-1.48(3.64)	PI	I	0.001997	0.00827
LI	FEZ1	p.D180V	-0.01(53.51)	PI	I	NA	0.00013
B5	GAD1	p.T27K	-0.51(21.56)	MI	I	0.001797	0.00915
B8	PTPRF	p.R216H	-3.92(0.20)	MI	II	NA	NA
B9	ATP2B4	p.G288V	-0.83(11.51)	PI	II	0.003994	0.01072
B5	DLG2	p.P395S	-1.60(3.04)	MI	II	NA	NA
B5	PTPN11	p.K131R	-0.43(25.15)	PI	II	NA	0.00038
B8	PGAM5	p.R118P	0.17(65.76)	PI	II	NA	NA
B5	MAP1A	p.P2127R	1.16(92.61)	MI	II	NA	NA
B11	MYO5A	p.R609C	-1.41(4.15)	PI	II	0.000599	0.0014
B8	ABR	p.N431S	-1.04(7.80)	MI	II	0.0002	0.00165
B10	DNM2	p.A253V	-1.33(4.71)	MI	II	NA	NA
B9	SYNJ1	p.A512V	0.50(79.67)	MI	II	0.000599	0.00571
B8	ATP5O	p.S91R	-0.05(50.01)	MI	II	NA	0.00051
B5	ITPR1	p.I2030M	-3.35(0.40)	MI	II	0.005591	0.0512
LI	ITPR1	p.I1421V	-3.35(0.40)	MI	II	NA	
B11	IQSEC1	p.R457Q	-1.26(5.34)	MI	II	0.002596	0.0187
B5	SDHA	p.Y55H	-0.86(10.95)	MI	II	0.003195	0.02124
B8	GSN	p.R144Q	0.61(82.94)	MI	II	0.0002	0.00242
B12	GSN	p.K546R	0.61(82.94)	MI	II	0.0002	0.00051
B6	LRRIQ3	p.R525C	1.38(94.60)	MI	III	0.004393	0.00013
B9	LRRIQ3	p.R180*	1.38(94.60)	PI	III	0.000399	0.00013
LI	VPS45	p.Y395C	-0.54(20.54)	MI	III	NA	NA
B5	CLP1	p.R143C	-0.05(50.01)	PI	III	NA	NA
B9	LRP1	p.S2934L	-7.28(0.02)	PI	III	0.000799	0.00064
B6	SHMT2	p.R437H	-0.11(45.26)	MI	III	0.002396	0.02708
B9	BCL11B	p.R319Q	NA	PI	III	NA	NA
LI	PLCB2	p.V356M	-0.70(14.81)	PI	III	0.0002	NA
B12	MAN2A2	p.D891G	-1.45(3.93)	MI	III	0.004193	0.01996
LI	MAN2A2	p.D891G	-1.45(3.93)	PI	III	0.004193	0.01996
B5	GRIN2A	p.R856G	-1.46(3.89)	MI	III	0.000399	0.00013
B5	SEZ6L2	p.A242V	-1.12(6.58)	PI	III	NA	NA
B5	CNOT1	p.N2204S	-1.90(1.97)	MI	III	0.001198	0.00264
B11	THAP11	p.L157V	-0.30(32.62)	MI	III	0.000799	0.00191
B6	CTRL	p.H75Y	0.60(82.66)	MI	III	0.003794	0.01375
B12	NFATC3	p.P297S	0.16(64.96)	MI	III	NA	0.00102
B5	SGSM2	p.G533E	-0.74(13.80)	MI	III	0.001597	0.01329

B10	RAI1	p.P1432L	-3.68(0.25)	PI	III	0.000998	0.00581
B6	SREBF1	p.V610M	-0.79(12.60)	MI	III	0.007388	0.00013
LI	TOM1L2	p.R65T	-1.16(6.17)	MI	III	NA	NA
B6	MYO15A	p.A2248V	-1.26(5.35)	PI	III	0.004393	NA
B12	GATAD2A	p.L207V	-0.95(9.27)	MI	III	0.0002	0.00293
B10	PRRG2	p.S73C	0.66(84.35)	MI	III	0.002396	0.01398
B11	CYP26B1	p.W172R	0.00(54.03)	PI	III	NA	NA
B12	EPC2	p.I461V	-0.29(33.20)	MI	III	NA	NA
B5	PLCL1	p.A809T	-0.24(36.28)	PI	III	NA	NA
B8	C2orf82	p.I98M	NA	PI	III	NA	NA
B9	ACTR5	p.P580L	0.27(70.58)	MI	III	0.001997	0.01659
LI	CYP2D6	p.G169R	1.77(96.77)	PI	III	0.001997	0.01531
B9	TCF20	p.P1240A	-2.55(0.85)	MI	III	NA	0.00102
B9	TRANK1	p.A1422T	-3.56(0.31)	MI	III	0.002196	0.00242
LI	TRANK1	p.V601L	-3.56(0.31)	MI	III	0.005591	0.02346
LI	NISCH	p.R930H	-2.31(1.20)	PI	III	0.00619	0.02747
B8	NISCH	p.Q1463H	-2.31(1.20)	PI	III	0.004593	0.02175
B12	STAB1	p.R2071H	-4.12(0.16)	MI	III	NA	NA
B6	MSL2	p.P25L	-0.53(20.70)	PI	III	0.0002	0.00102
B12	MAN2A1	p.I459M	-1.57(3.20)	PI	III	0.0002	
B6	WDR55	p.S246N	0.46(78.59)	MI	III	0.0002	0.00432
B11	PCDHA5	frameshift	0.10(60.76)	PI	III	NA	NA
B8	PCDHA6	p.N403T	-0.76(13.18)	PI	III	0.00599	0.02365
B12	PCDHA6	p.N403T	-0.76(13.18)	PI	III	0.00599	0.02365
LI	PCDHA10	p.G627V	1.61(95.93)	PI	III	0.000399	0.0028
B5	GALNT10	p.K433R	-1.20(5.79)	PI	III	0.002596	0.00878
B10	RIMS1	p.L1314P	-1.54(3.32)	PI	III	0.001997	0.00106
B10	MPP6	p.L343M	-0.09(46.74)	MI	III	NA	NA

^aFrom a distribution of 16,956 genes with pre-calculated respective residual variation intolerance score (RVIS) (Petrovski et al., 2013). ^b MI – inherited from mother; PI – inherited from father. ^c Class I - 38 genes in the SZGR data set (Jia et al., 2010); Class II - 206 genes in SCZ-related pathways or complexes, including PSD-95, ARC, NMDAR network, mGluR5, which were reported in Kirov et al. (2012); class III - 352 genes targeted by the 108 SCZ susceptibility loci showing a genome-wide significance (Schizophrenia Working Group of the Psychiatric Genomics, 2014). ^dThe minor allele frequency (MAF) from the 1000 Genomes Project (Sudmant et al., 2015). ^e MAF from Exome Aggregation Consortium (ExAC)'s East Asian Studies (EAS) (Lek et al., 2016).

Table S14. Polygenic burden analysis of rare damaging and loss of function variants in schizophrenia-related gene sets

Gene set	Number of genes	Number of mutations		<i>p</i> -value ⁱ	Odd ratio (OR)
		Twins	Parents		
Class I gene					
SZGR core gene ^a	38	10	8	0.647	1.25287
Class II gene					
PSD human core ^b	685	148	147	0.953	1.00926
NMDAR network	61	14	10	0.423	1.40497
mGluR5	39	12	11	0.838	1.09342
PSD-95	65	14	18	0.596	0.77794
Pre-synapse ^b	431	68	54	0.203	1.26922
Pre-synaptic active zone	173	26	21	0.469	1.24304
Synaptic vesicle	344	48	38	0.281	1.27093
Class III gene					
108 schizophrenia-risk loci ^c	352	92	72	0.115	1.29131
Other gene sets					
schizophrenia <i>de novo</i> mutations ^d	613	282	282	1.000	1.00212
FMR1 targets ^e	1589	608	584	0.439	1.04926
Axon guidance pathways ^f	432	128	128	1.000	1.00198
Nerve impulse ^f	237	30	37	0.462	0.80996
Nervous system development ^f	1874	506	497	0.712	1.02534
Neuronal cell body ^f	309	66	71	0.731	0.92938
Neuron projection ^f	699	160	150	0.564	1.07355
Cancer pathway ^g	398	104	95	0.520	1.09893
CFG analysis identification ^h	42	16	15	0.860	1.06900
Linkage and association study ^h	223	54	52	0.846	1.04100
Pascal Gene-based test identification ^h	343	54	49	0.622	1.10500
Sherlock analysis identification ^h	12	2	3	1.000	0.66778

^a Gene set of the SZGR database (Jia et al., 2010). ^b Gene sets in Kirov et al. (2012). ^c Gene set of 108 schizophrenia-associated loci showing a genome-wide significance (Schizophrenia Working Group of the Psychiatric Genomics, 2014). ^d Gene set of nonsynonymous *de novo* mutations reported by Fromer et al. (2014). ^e Genes listed as targets of FMRP (fragile X mental retardation protein) by Darnell et al. (2011) and Ascano et al. (2012). ^f Gene sets reported by Yuen et al. (2015). ^g Gene set of the KEGG database (2000). ^h Gene sets of the SZDB database (<http://www.szdb.org/>) (Wu et al., 2017). ⁱ *p*-value of the enrichment of rare variants in each gene set was estimated by the Fisher's exact test.

Table S15. The number of CNVs (> 1 Kb) in 8 families with MZ twins

Family	Father	Mother	Proband	Mean	Total
B10	337	287	292	305.333	916
B11	202	306	248	252	756
B12	198	216	253	222.333	667
B5	307	328	352	329	987
B6	269	266	260	265	795
B8	322	318	273	304.333	913
B9	290	240	280	270	810
LI	212	282	281	258.333	775
Mean	267.125	280.375	279.875	275.792	-
Total	2137	2243	2239	-	6619

Table S16. Summary of the inherited CNV in schizophrenia-associated genes in 8 pairs of MZ twins

Family	CNVs	Gene	Type ^a	Prediction ^b	Reference
B5	chr16:16,934,363-16,940414; Deletion	intergenic	MI or PI	Delly; Lumpy; CNVnator	(Ingason et al., 2011)
B6	chr3:196934573-196939255; Deletion	DLG1 intron	MI	Delly; Lumpy; CNVnator	(Levinson et al., 2011; Walsh et al., 2008)
B11	chr16: 15273991-18185846; Deletion	<i>MPVI7L, C16orf45, KIAA0430, NDE1, MYH11, C16orf63, ABCC1, ABCC6, NOMO3 and XYLT1</i>	PI	Delly; Lumpy; BreakDancer	(Ingason et al., 2011)
LI	chr3:195198880-195223975; Deletion	intergenic	MI	Lumpy; CNVnator	(Levinson et al., 2011; Walsh et al., 2008)

^a MI – inherited from mother; PI – inherited from father. ^b The CNVs (length > 1000 bp) was calculated by combining the results from four algorithms (Breakdancer (Chen et al., 2009), CNVnator (Abzyzov et al., 2011), Delly (Rausch et al., 2012) and Lumpy (Layer et al., 2014)).

Supplemental References

- Abyzov A., Urban A.E., Snyder M., Gerstein M., 2011. CNVnator: an approach to discover, genotype, and characterize typical and atypical CNVs from family and population genome sequencing. *Genome Res.* 21:974-984.
- Adzhubei I., Jordan D.M., Sunyaev S.R., 2013. Predicting functional effect of human missense mutations using PolyPhen-2. *Curr. Protoc. Hum. Genet.* Chapter. 7:Unit7 20.
- Adzhubei I.A., Schmidt S., Peshkin L., Ramensky V.E., Gerasimova A., Bork P., Kondrashov A.S., Sunyaev S.R., 2010. A method and server for predicting damaging missense mutations. *Nature Methods* 7:248-249.
- Ascano M., Jr., Mukherjee N., Bandaru P., Miller J.B., Nusbaum J.D., Corcoran D.L., Langlois C., Munschauer M., Dewell S., Hafner M., Williams Z., Ohler U., Tuschl T., 2012. FMRP targets distinct mRNA sequence elements to regulate protein expression. *Nature* 492:382-386.
- Chen K., Wallis J.W., McLellan M.D., Larson D.E., Kalicki J.M., Pohl C.S., McGrath S.D., Wendl M.C., Zhang Q., Locke D.P., Shi X., Fulton R.S., Ley T.J., Wilson R.K., Ding L., Mardis E.R., 2009. BreakDancer: an algorithm for high-resolution mapping of genomic structural variation. *Nat. Methods* 6:677-681.
- Chun S., Fay J.C., 2009. Identification of deleterious mutations within three human genomes. *Genome Res.* 19:1553-1561.
- Darnell J.C., Van Driesche S.J., Zhang C., Hung K.Y., Mele A., Fraser C.E., Stone E.F., Chen C., Fak J.J., Chi S.W., Licatalosi D.D., Richter J.D., Darnell R.B., 2011. FMRP stalls ribosomal translocation on mRNAs linked to synaptic function and autism. *Cell* 146:247-261.
- Fromer M., Pocklington A.J., Kavanagh D.H., Williams H.J., Dwyer S., Gormley P., Georgieva L., Rees E., Palta P., Ruderfer D.M., Carrera N., Humphreys I., Johnson J.S., Roussos P., Barker D.D., Banks E., Milanova V., Grant S.G., Hannon E., Rose S.A., Chambert K., Mahajan M., Scolnick E.M., Moran J.L., Kirov G., Palotie A., McCarroll S.A., Holmans P., Sklar P., Owen M.J., Purcell S.M., O'Donovan M.C., 2014. *De novo* mutations in schizophrenia implicate synaptic networks. *Nature* 506:179-184.
- Ingason A., Rujescu D., Cichon S., Sigurdsson E., Sigmundsson T., Pietilainen O.P., Buizer-Voskamp J.E., Strengman E., Francks C., Muglia P., Gylfason A., Gustafsson O., Olason P.I., Steinberg S., Hansen T., Jakobsen K.D., Rasmussen H.B., Giegling I., Moller H.J., Hartmann A., Crombie C., Fraser G., Walker N., Lonnqvist J., Suvisaari J., Tuulio-Henriksson A., Bramon E., Kiemeneij L.A., Franke B., Murray R., Vassos E., Toulopoulou T., Muhleisen T.W., Tosato S., Ruggeri M., Djurovic S., Andreassen O.A., Zhang Z., Werge T., Ophoff R.A., Investigators G., Rietschel M., Nothen M.M., Petursson H., Stefansson H., Peltonen L., Collier D., Stefansson K., St Clair D.M., 2011. Copy number variations of chromosome 16p13.1 region associated with

- schizophrenia. Mol. Psychiatry 16:17-25.
- Jia P., Sun J., Guo A.Y., Zhao Z., 2010. SZGR: a comprehensive schizophrenia gene resource. Mol. Psychiatry 15:453-462.
- Kanehisa M., Goto S., 2000. KEGG: kyoto encyclopedia of genes and genomes. Nucleic Acids Res. 28:27-30.
- Kirov G., Pocklington A.J., Holmans P., Ivanov D., Ikeda M., Ruderfer D., Moran J., Chambert K., Toncheva D., Georgieva L., Grozeva D., Fjodorova M., Wollerton R., Rees E., Nikolov I., van de Lagemaat L.N., Bayes A., Fernandez E., Olason P.I., Bottcher Y., Komiyama N.H., Collins M.O., Choudhary J., Stefansson K., Stefansson H., Grant S.G., Purcell S., Sklar P., O'Donovan M.C., Owen M.J., 2012. De novo CNV analysis implicates specific abnormalities of postsynaptic signalling complexes in the pathogenesis of schizophrenia. Mol. Psychiatry 17:142-153.
- Kumar P., Henikoff S., Ng P.C., 2009. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. Nat. Protoc. 4:1073-1081.
- Layer R.M., Chiang C., Quinlan A.R., Hall I.M., 2014. LUMPY: a probabilistic framework for structural variant discovery. Genome Biol. 15:R84.
- Lek M., Karczewski K.J., Minikel E.V., Samocha K.E., Banks E., Fennell T., O'Donnell-Luria A.H., Ware J.S., Hill A.J., Cummings B.B., Tukiainen T., Birnbaum D.P., Kosmicki J.A., Duncan L.E., Estrada K., Zhao F., Zou J., Pierce-Hoffman E., Berghout J., Cooper D.N., Deflaux N., DePristo M., Do R., Flannick J., Fromer M., Gauthier L., Goldstein J., Gupta N., Howrigan D., Kiezun A., Kurki M.I., Moonshine A.L., Natarajan P., Orozco L., Peloso G.M., Poplin R., Rivas M.A., Ruano-Rubio V., Rose S.A., Ruderfer D.M., Shakir K., Stenson P.D., Stevens C., Thomas B.P., Tiao G., Tusie-Luna M.T., Weisburd B., Won H.H., Yu D., Altshuler D.M., Ardiissino D., Boehnke M., Danesh J., Donnelly S., Elosua R., Florez J.C., Gabriel S.B., Getz G., Glatt S.J., Hultman C.M., Kathiresan S., Laakso M., McCarroll S., McCarthy M.I., McGovern D., McPherson R., Neale B.M., Palotie A., Purcell S.M., Saleheen D., Scharf J.M., Sklar P., Sullivan P.F., Tuomilehto J., Tsuang M.T., Watkins H.C., Wilson J.G., Daly M.J., MacArthur D.G., Exome Aggregation C., 2016. Analysis of protein-coding genetic variation in 60,706 humans. Nature 536:285-291.
- Levinson D.F., Duan J., Oh S., Wang K., Sanders A.R., Shi J., Zhang N., Mowry B.J., Olincy A., Amin F., Cloninger C.R., Silverman J.M., Buccola N.G., Byerley W.F., Black D.W., Kendler K.S., Freedman R., Dudbridge F., Pe'er I., Hakonarson H., Bergen S.E., Fanous A.H., Holmans P.A., Gejman P.V., 2011. Copy number variants in schizophrenia: confirmation of five previous findings and new evidence for 3q29 microdeletions and VIPR2 duplications. Am. J. Psychiatry 168:302-316.
- Li H., Bi R., Fan Y., Wu Y., Tang Y., Li Z., He Y., Zhou J., Tang J., Chen X., Yao Y.G., 2016. mtDNA heteroplasmy in monozygotic twins discordant for schizophrenia. Mol. Neurobiol. doi: 10.1007/s12035-016-9996-x
- Montojo J., Zuberi K., Rodriguez H., Bader G.D., Morris Q., 2014. GeneMANIA:

- Fast gene network construction and function prediction for Cytoscape. F1000Res 3:153.
- Ng P.C., 2003. SIFT: predicting amino acid changes that affect protein function. Nucleic. Acids. Res. 31:3812-3814.
- Petrovski S., Wang Q., Heinzen E.L., Allen A.S., Goldstein D.B., 2013. Genic intolerance to functional variation and the interpretation of personal genomes. PLoS Genet. 9:e1003709.
- Rausch T., Zichner T., Schlattl A., Stutz A.M., Benes V., Korbel J.O., 2012. DELLY: structural variant discovery by integrated paired-end and split-read analysis. Bioinformatics 28:i333-i339.
- Schizophrenia Working Group of the Psychiatric Genomics C., 2014. Biological insights from 108 schizophrenia-associated genetic loci. Nature 511:421-427.
- Schwarz JM, Cooper DN, Schuelke M, D. S., 2014. MutationTaster2: mutation prediction for the deep-sequencing age. Nat. Methods 11:361-362.
- Sudmant P.H., Rausch T., Gardner E.J., Handsaker R.E., Abyzov A., Huddleston J., Zhang Y., Ye K., Jun G., Hsi-Yang Fritz M., Konkel M.K., Malhotra A., Stutz A.M., Shi X., Paolo Casale F., Chen J., Hormozdiari F., Dayama G., Chen K., Malig M., Chaisson M.J., Walter K., Meiers S., Kashin S., Garrison E., Auton A., Lam H.Y., Jasmine Mu X., Alkan C., Antaki D., Bae T., Cerveira E., Chines P., Chong Z., Clarke L., Dal E., Ding L., Emery S., Fan X., Gujral M., Kahveci F., Kidd J.M., Kong Y., Lameijer E.W., McCarthy S., Flück P., Gibbs R.A., Marth G., Mason C.E., Menelaou A., Muzny D.M., Nelson B.J., Noor A., Parrish N.F., Pendleton M., Quitadamo A., Raeder B., Schadt E.E., Romanovitch M., Schlattl A., Sebra R., Shabalin A.A., Untergasser A., Walker J.A., Wang M., Yu F., Zhang C., Zhang J., Zheng-Bradley X., Zhou W., Zichner T., Sebat J., Batzer M.A., McCarroll S.A., Genomes Project C., Mills R.E., Gerstein M.B., Bashir A., Stegle O., Devine S.E., Lee C., Eichler E.E., Korbel J.O., 2015. An integrated map of structural variation in 2,504 human genomes. Nature 526:75-81.
- Walsh T., McClellan J.M., McCarthy S.E., Addington A.M., Pierce S.B., Cooper G.M., Nord A.S., Kusenda M., Malhotra D., Bhandari A., Stray S.M., Rippey C.F., Rocanova P., Makarov V., Lakshmi B., Findling R.L., Sikich L., Stromberg T., Merriman B., Gogtay N., Butler P., Eckstrand K., Noory L., Gochman P., Long R., Chen Z., Davis S., Baker C., Eichler E.E., Meltzer P.S., Nelson S.F., Singleton A.B., Lee M.K., Rapoport J.L., King M.C., Sebat J., 2008. Rare structural variants disrupt multiple genes in neurodevelopmental pathways in schizophrenia. Science 320:539-543.
- Wu Y., Yao Y.G., Luo X.J., 2017. SZDB: A database for schizophrenia genetic research. Schizophr. Bull. 43:459-471.
- Yang J., Lee S.H., Goddard M.E., Visscher P.M., 2011. GCTA: a tool for genome-wide complex trait analysis. Am. J. Hum. Genet. 88:76-82.
- Yuen R.K., Thiruvahindrapuram B., Merico D., Walker S., Tammimies K., Hoang N., Chrysler C., Nalpathamkalam T., Pellecchia G., Liu Y., Gazzellone M.J., D'Abate L., Deneault E., Howe J.L., Liu R.S., Thompson A., Zarrei M., Uddin

M., Marshall C.R., Ring R.H., Zwaigenbaum L., Ray P.N., Weksberg R., Carter M.T., Fernandez B.A., Roberts W., Szatmari P., Scherer S.W., 2015. Whole-genome sequencing of quartet families with autism spectrum disorder. Nat. Med. 21:185-191.

Supplemental Table S4. The distribution of DNMs in 8 pairs of MZ twins

Sample	Chromosome	chromPosition	Allele1	Allele2	Band	dbSNP	Gene	Transcript	Predicted function
B103 B104	chr3	118923152	T	C	q13.32		UPK1B	NM_006952	3utr
B103 B104	chr9	17578171	A	C	p22.2		SH3GL2	NM_003026	5upstream
B103 B104	chr10	71169085	T	G	q22.1		TACR2	NM_001057	intronic
B103 B104	chr11	126855973	C	A	q24.2	rs11220689	KIRREL3	NM_001161707	intronic
B103 B104	chr12	11507171	G	A	p13.2		PRB1	NM_005039	intronic
B103 B104	chr12	104900233	A	G	q23.3		CHST11	NM_001173982	intronic
B103 B104	chr12	133397352	G	A	q24.33		GOLGA3	NM_001172557	intronic
B103 B104	chr13	43170170	G	A	q14.11		TNFSF11	NM_003701	intronic
B103 B104	chr13	74635919	C	T	q22.1	rs114949718	KLF12	NM_007249	intronic
B103 B104	chr15	56398367	T	C	q21.3		RFX7	NM_022841	intronic
B103 B104	chr15	78321335	T	G	q25.1		TBC1D2B	NM_015079	intronic
B103 B104	chr16	19266444	C	T	p12.3		SYT17	NM_016524	intronic
B103 B104	chr17	20145251	C	T	p11.2		SPECC1	NM_001033553	intronic
B103 B104	chr17	66580685	C	T	q24.2		FAM20A	NM_001243746	intronic
B103 B104	chr18	64205104	G	A	q22.1		CDH19	NM_001271028	intronic
B103 B104	chr2	185753314	C	A	q32.1		ZNF804A	NM_194250	intronic
B103 B104	chr2	231640380	T	G	q37.1		CAB39	NM_001130849	intronic
B103 B104	chr20	19700562	G	A	p11.23		SLC24A3	NM_020689	intronic
B103 B104	chr20	34319685	T	G	q11.22		RBM39	NM_001242599	intronic
B103 B104	chr20	34752676	C	T	q11.23		EPB41L1	NM_001258329	intronic
B103 B104	chr20	37200837	G	T	q11.23		RALGAPB	NM_001282917	intronic
B103 B104	chr3	68974336	T	C	p14.1		FAM19A4	NM_001005527	intronic
B103 B104	chr3	89209126	T	G	p11.1		EPHA3	NM_005233	intronic
B103 B104	chr3	178375930	C	T	q26.32		KCNMB2	NM_001278911	intronic
B103 B104	chr5	63649074	T	A	q12.3		RNF180	NM_001113561	intronic
B103 B104	chr8	674731	A	G	p23.3		ERICH1	NM_207332	intronic
B103 B104	chr8	118537055	G	A	q24.11		MED30	NM_001282986	intronic
B103 B104	chr8	121565361	A	G	q24.12		SNTB1	NM_021021	intronic
B103 B104	chr10	128776253	G	T	q26.2		DOCK1	NM_001290223	intronic (splice_site) IVS3+1G>T
B103 B104	chr13	101552351	C	A	q32.3		NALCN_AS1	NR_047687	non-coding intronic
B103 B104	chr14	104320775	G	A	q32.33		LINC00637	NR_038436	non-coding intronic
B103 B104	chr11	94241750	C	A	q21				intergenic
B103 B104	chr12	42352274	A	C	q12				intergenic
B103 B104	chr12	58842837	C	G	q14.1				intergenic
B103 B104	chr12	79083015	G	A	q21.2				intergenic
B103 B104	chr13	72813354	G	T	q21.33				intergenic
B103 B104	chr14	78571385	T	C	q24.3	rs177181			intergenic
B103 B104	chr14	85591083	T	C	q31.3				intergenic
B103 B104	chr16	63843879	C	T	q21				intergenic
B103 B104	chr16	69210370	A	C	q22.1				intergenic
B103 B104	chr16	69210371	C	A	q22.1				intergenic
B103 B104	chr17	2421858	C	T	p13.3				intergenic
B103 B104	chr17	41633492	A	C	q21.31	rs112683677			intergenic
B103 B104	chr18	57721409	A	G	q21.32				intergenic
B103 B104	chr2	21646773	C	T	p24.1				intergenic
B103 B104	chr2	82569412	T	C	p12				intergenic
B103 B104	chr2	88639215	C	T	p11.2				intergenic
B103 B104	chr3	95967254	T	G	q11.2				intergenic
B103 B104	chr3	140390293	C	T	q23				intergenic
B103 B104	chr4	53162451	C	T	q12				intergenic
B103 B104	chr4	125864692	C	T	q28.1				intergenic
B103 B104	chr4	162128420	C	T	q32.2				intergenic
B103 B104	chr5	172158355	G	A	q35.1				intergenic
B103 B104	chr7	21283412	G	A	p15.3	rs116816019			intergenic
B103 B104	chr7	152835331	A	G	q36.2				intergenic
B103 B104	chr8	5214640	G	A	p23.2				intergenic
B103 B104	chr8	25459305	G	T	p21.2				intergenic
B103 B104	chr8	47649158	A	G	q11.1				intergenic
B103 B104	chr8	66834673	A	G	q13.1				intergenic
B103 B104	chr8	106280832	T	G	q23.1				intergenic
B103 B104	chr9	11417777	A	T	p23				intergenic
B103 B104	chr9	12145048	G	A	p23				intergenic
B103 B104	chrX	20489848	T	C	p22.12				intergenic
B103 B104	chrX	42471130	T	G	p11.3				intergenic
B103 B104	chrX	81626563	A	G	q21.1				intergenic
B103 B104	chr6	78082285	TA	T					intergenic
B104	chr2	153664053	G	A	q23.3	rs142683553			intergenic
B113	chr12	26742696	A	G	p11.23	rs11048597	ITPR2	NM_002223	intronic
B113	chr9	127918364	G	A	q33.3		PPP6C	NM_001123355	intronic
B113	chr20	52177664	A	G	q13.2		RP4_724E16.2	NR_110051	non-coding intronic
B113	chr15	96155017	C	A	q26.2				intergenic
B113	chr4	122576011	T	C	q27				intergenic
B113 B114	chr19	53997821	C	T	q13.42	rs10423700	ZNF813	NM_001004301	3downstream

B113 B114	chr20	34539772	A	G	q11.23	SCAND1	NM_016558	3downstream
B113 B114	chr3	98242563	T	C	q11.2	CLDND1	NM_001040181	5upstream
B113 B114	chr10	117458502	T	A	q25.3	ATRNL1	NM_207303	intronic
B113 B114	chr11	16923047	T	A	p15.1	PLEKHA7	NM_175058	intronic
B113 B114	chr12	80731159	G	A	q21.31	OTOGL	NM_173591	intronic
B113 B114	chr13	28557037	C	T	q12.2	URAD	NM_001105577	intronic
B113 B114	chr15	62958974	T	C	q22.2	TLN2	NM_015059	intronic
B113 B114	chr16	89961140	C	A	q24.3 rs35518096	TCF25	NM_014972	intronic
B113 B114	chr18	6348505	T	C	p11.31	L3MBTL4	NM_173464	intronic
B113 B114	chr18	32683856	G	C	q12.1	MAPRE2	NM_001143826	intronic
B113 B114	chr2	10036253	C	T	p25.1	TAF1B	NM_005680	intronic
B113 B114	chr3	171864901	G	C	q26.31	FNDC3B	NM_001135095	intronic
B113 B114	chr4	89304876	G	A	q22.1	HERC6	NM_001165136	intronic
B113 B114	chr5	127441266	A	G	q23.3	SLC12A2	NM_001046	intronic
B113 B114	chr9	124373251	T	C	q33.2	DAB2IP	NM_032552	intronic
B113 B114	chr1	101012039	G	T	p21.2			
B113 B114	chr10	123074966	T	C	q26.12			
B113 B114	chr11	27959616	C	G	p14.1			
B113 B114	chr12	103506236	C	T	q23.2			
B113 B114	chr13	65852160	C	A	q21.32			
B113 B114	chr16	24377252	A	G	p12.1			
B113 B114	chr16	86024734	T	C	q24.1 rs12596402			
B113 B114	chr17	26144055	A	G	q11.2			
B113 B114	chr2	207501019	T	C	q33.3			
B113 B114	chr2	220870844	T	C	q35			
B113 B114	chr22	49740309	G	C	q13.33			
B113 B114	chr3	174392522	T	C	q26.31			
B113 B114	chr5	31616208	A	G	p13.3			
B113 B114	chr5	118782822	G	A	q23.1 rs56372726			
B113 B114	chr6	34401949	G	A	p21.31			
B113 B114	chr6	142436831	C	G	q24.1			
B113 B114	chr6	145886359	G	C	q24.3			
B113 B114	chr7	108858722	C	A	q31.1			
B113 B114	chr7	108858732	G	T	q31.1			
B113 B114	chr11	33429549	G	GT				
B114	chr12	67837996	G	A	q15 rs35359961			
B114	chr13	21816875	A	G	q12.11 rs79194363			
B114	chr13	21816893	T	A	q12.11 rs78730531			
B114	chr13	21816900	G	A	q12.11 rs79353765			
B114	chr13	21816903	A	G	q12.11 rs76326863			
B114	chr6	154215825	T	C	q25.2 rs5003108			
B114	chr9	66861389	T	C	q13 rs76206935			
B114	chr9	66861395	T	C	q13 rs78501402			
B123	chr1	63702337	A	C	p31.3	LINC00466	NR_038252	non-coding intronic
B123	chr14	43121265	G	A	q21.1			
B123 B124	chr17	60527119	A	G	q23.2	METTL2A	NM_181725	3utr
B123 B124	chr2	172411647	G	C	q31.1	CYBRD1	NM_001127383	3utr
B123 B124	chr19	35449194	T	C	q13.11	ZNF792	NM_175872	nonsyn p.N522S
B123 B124	chr10	15589618	C	T	p13	ITGA8	NM_003638	intronic
B123 B124	chr10	87386677	A	C	q23.1	GRID1	NM_017551	intronic
B123 B124	chr11	47691714	A	G	p11.2	AGBL2	NM_024783	intronic
B123 B124	chr12	19654480	A	T	p12.3	AEBP2	NM_001114176	intronic
B123 B124	chr21	42082429	G	A	q22.2 rs17000228	DSCAM	NM_001271534	intronic
B123 B124	chr3	7391648	G	A	p26.1	GRM7	NM_000844	intronic
B123 B124	chr3	13085170	C	T	p25.2 rs138660003	IQSEC1	NM_001134382	intronic
B123 B124	chr3	51651550	C	T	p21.2	RAD54L2	NM_015106	intronic
B123 B124	chr3	60760326	C	G	p14.2	FHIT	NM_001166243	intronic
B123 B124	chr3	139887457	G	T	q23 rs78952941	CLSTN2	NM_022131	intronic
B123 B124	chr4	2463556	C	T	p16.3	RP11_503N18	NM_001193282	intronic
B123 B124	chr4	15798530	G	T	p15.32	CD38	NM_001775	intronic
B123 B124	chr5	34827544	A	C	p13.2	RAI14	NM_001145520	intronic
B123 B124	chr5	64235781	C	A	q12.3	CWC27	NM_005869	intronic
B123 B124	chr5	78999514	T	C	q14.1	CMYA5	NM_153610	intronic
B123 B124	chr5	79459636	T	C	q14.1	SERINC5	NM_001174071	intronic
B123 B124	chr6	56419279	C	T	p12.1	DST	NM_183380	intronic
B123 B124	chr6	56978734	T	G	p12.1	ZNF451	NM_001031623	intronic
B123 B124	chr8	1779112	G	A	p23.3 rs185594803	ARHGEF10	NM_014629	intronic
B123 B124	chr8	23211573	T	G	p21.3	LOXL2	NM_002318	intronic
B123 B124	chr8	100386318	A	G	q22.2	VPS13B	NM_017890	intronic
B123 B124	chr9	15437524	C	T	p22.3	SNAPC3	NM_001039697	intronic
B123 B124	chr9	99136093	G	C	q22.32	SLC35D2	NM_001286990	intronic
B123 B124	chr9	139787912	G	T	q34.3	TRAF2	NM_021138	intronic
B123 B124	chr2	67362239	G	A	p14	AC078941.1	NR_038844	non-coding intronic
B123 B124	chr3	168345596	T	G	q26.2	EGFEM1P	NR_021485	non-coding intronic
B123 B124	chr3	181670965	C	T	q26.33	LINC01206	NR_104146	non-coding intronic
B123 B124	chr6	150229677	G	T	q25.1	RAET1E_AS1	NR_045126	non-coding intronic

B123	B124	chr10	20845041	C	T	p12.31	rs192503538	intergenic	
B123	B124	chr12	75146156	T	C	q21.1		intergenic	
B123	B124	chr12	129314669	A	G	q24.33		intergenic	
B123	B124	chr13	76971899	G	C	q22.2		intergenic	
B123	B124	chr14	46455725	T	G	q21.2		intergenic	
B123	B124	chr14	104390533	C	T	q32.33		intergenic	
B123	B124	chr16	35013471	T	A	p11.1		intergenic	
B123	B124	chr16	77170825	G	A	q23.1		intergenic	
B123	B124	chr19	31507732	T	G	q12		intergenic	
B123	B124	chr2	41332857	C	T	p22.1		intergenic	
B123	B124	chr20	55421177	T	G	q13.31		intergenic	
B123	B124	chr21	40959364	T	C	q22.2		intergenic	
B123	B124	chr3	3361010	G	C	p26.2		intergenic	
B123	B124	chr3	21088283	C	T	p24.3		intergenic	
B123	B124	chr3	111154558	G	T	q13.13		intergenic	
B123	B124	chr4	33398722	C	T	p15.1		intergenic	
B123	B124	chr4	55065295	T	A	q12		intergenic	
B123	B124	chr4	128289126	C	T	q28.1		intergenic	
B123	B124	chr5	97928714	T	G	q15		intergenic	
B123	B124	chr6	138054222	T	C	q23.3		intergenic	
B123	B124	chr7	125495096	C	T	q31.33		intergenic	
B123	B124	chr9	11027822	C	T	p23		intergenic	
B123	B124	chr9	24903909	G	A	p21.3		intergenic	
B123	B124	chr9	77915529	G	A	q21.13		intergenic	
B123	B124	chr9	82729515	A	G	q21.31		intergenic	
B123	B124	chrX	144020346	G	A	q27.3		intergenic	
B123	B124	chr2	181977397	TC	T		AC104820.2	ENST0000041475 intron	
B124	chr1	230530315	T	C	q42.13		PGBD5	NM_001258311 intronic	
B124	chr17	12638660	T	C	p12	rs7217163	MYOCD	NM_001146312 intronic	
B124	chr17	12638661	A	G	p12		MYOCD	NM_001146312 intronic	
B53	B54	chr1	40982533	A	G	p34.2		EXO5	NM_022774 3downstream
B53	B54	chr17	41382051	T	C	q21.31	rs77050562	LINC00854	NR_047479 5upstream
B53	B54	chr12	120569035	C	G	q24.23		GCN1L1	NM_006836 nonsyn p.S2506T
B53	B54	chr17	39595049	G	T	q21.2		KRT38	NM_006771 nonsyn p.P265H
B53	B54	chr2	179436794	C	T	q31.2		TTN	NM_001256850 nonsyn p.V23048I
B53	B54	chr1	182887203	C	A	q25.3	rs13373923	SHCBP1L	NM_030933 intronic
B53	B54	chr11	57175782	C	T	q12.1		SLC43A3	NM_001278201 intronic
B53	B54	chr12	634274	T	G	p13.33	rs200275288	B4GALNT3	NM_173593 intronic
B53	B54	chr13	36215362	A	C	q13.3		NBEA	NM_001204197 intronic
B53	B54	chr14	37701455	T	C	q13.3		MIPOL1	NM_001195296 intronic
B53	B54	chr15	68647297	T	A	q23		ITGA11	NM_001004439 intronic
B53	B54	chr15	77314816	G	A	q24.3		PSTPIP1	NM_003978 intronic
B53	B54	chr2	68543768	A	T	p14		CNRIP1	NM_001111101 intronic
B53	B54	chr2	230096243	G	A	q36.3		PID1	NM_001100818 intronic
B53	B54	chr2	230274367	C	T	q36.3		DNER	NM_139072 intronic
B53	B54	chr2	241555733	C	A	q37.3		GPR35	NM_001195381 intronic
B53	B54	chr3	57476826	A	C	p14.3	rs74933745	DNAH12	NM_178504 intronic
B53	B54	chr4	81742124	C	T	q21.21		C4orf22	NM_001206997 intronic
B53	B54	chr5	19663794	A	G	p14.3		CDH18	NM_001167667 intronic
B53	B54	chr5	61635205	C	T	q12.1	rs2034240	KIF2A	NM_001098511 intronic
B53	B54	chr5	177563147	C	T	q35.3		RMND5B	NM_001288794 intronic
B53	B54	chr6	123917045	A	G	q22.31		TRDN	NM_001251987 intronic
B53	B54	chr8	56129326	G	A	q12.1		XKR4	NM_052898 intronic
B53	B54	chr9	16205585	C	T	p22.3		C9orf92	NM_001271829 intronic
B53	B54	chr9	34617204	C	T	p13.3		DCTN3	NM_001281425 intronic
B53	B54	chr11	101016266	T	C	q22.1		LOC10105452	NR_073144 non-coding intronic
B53	B54	chr5	16385105	T	C	p15.1		RP1_167G20.1	NR_104625 non-coding intronic
B53	B54	chr6	140126330	A	G	q24.1		LOC10013273	NR_038399 non-coding intronic
B53	B54	chr7	20169410	C	T	p21.1		AC005062.2	NR_110114 non-coding intronic
B53	B54	chr1	14607191	C	T	p36.21			intergenic
B53	B54	chr1	43588564	C	T	p34.2	rs9661826		intergenic
B53	B54	chr1	191364591	T	G	q31.2			intergenic
B53	B54	chr11	30052740	G	C	p14.1			intergenic
B53	B54	chr11	31215844	C	G	p13			intergenic
B53	B54	chr13	32042443	G	A	q12.3			intergenic
B53	B54	chr13	42917019	C	G	q14.11			intergenic
B53	B54	chr2	124712603	A	C	q14.3	rs375668524		intergenic
B53	B54	chr22	43077904	T	C	q13.2	rs130363		intergenic
B53	B54	chr3	76877788	A	G	p12.3			intergenic
B53	B54	chr4	12439530	C	A	p15.33			intergenic
B53	B54	chr4	114760417	T	A	q26			intergenic
B53	B54	chr4	155058899	C	A	q31.3			intergenic
B53	B54	chr5	118144221	T	C	q23.1			intergenic
B53	B54	chr6	17265790	C	T	p22.3			intergenic
B53	B54	chr6	133263812	G	C	q23.2			intergenic
B53	B54	chr7	2384889	C	G	p22.3			intergenic

B53 B54	chr7	24594488	T	C	p15.3		intergenic	
B53 B54	chr7	113264399	A	G	q31.1	rs7804693	intergenic	
B53 B54	chr8	43297461	T	G	p11.1		intergenic	
B53 B54	chr8	57645596	T	A	q12.1		intergenic	
B53 B54	chr8	111879636	T	A	q23.2		intergenic	
B53 B54	chr3	74615676	TC	T			intergenic	
B53 B54	chr17	41382052	T	C	q21.31	rs76863987	LINC00854	NR_047479
B54	chr12	76731688	G	T	rs7804693	q21.2	intergenic	5upstream
B62	chr7	27657582	G	A	p15.2		HIBADH	NM_152740
B62	chr11	35864072	A	T	p13	rs56352615	intergenic	intronic
B62	chr13	87882896	A	C	q31.2		intergenic	
B62 B63	chr1	231116417	A	G	q42.2		TTC13	NM_001122835
B62 B63	chr15	90929885	T	C	q26.1		IQGAP1	NM_003870
B62 B63	chr2	242283282	A	G	q37.3		SEPT2	NM_001008491
B62 B63	chr1	23120852	C	T	p36.12		EPHB2	NM_004442
B62 B63	chr10	24095479	T	C	p12.2		KIAA1217	NM_001098500
B62 B63	chr10	31767758	G	A	p11.22		ZEB1	NM_001128128
B62 B63	chr10	95127010	C	T	q23.33		MYOF	NM_013451
B62 B63	chr10	127420577	G	A	q26.13		EDRF1	NM_001202438
B62 B63	chr11	34949538	A	G	p13		PDHX	NM_001135024
B62 B63	chr11	60294628	C	T	q12.2	rs185033225	MS4A13	NM_001012417
B62 B63	chr12	28411429	G	T	p11.22		CCDC91	NM_018318
B62 B63	chr14	64473123	C	T	q23.2		SYNE2	NM_015180
B62 B63	chr16	81489957	A	C	q23.2	rs9930117	CMIP	NM_198390
B62 B63	chr2	43609005	G	C	p21		THADA	NM_001083953
B62 B63	chr2	149183277	G	A	q23.1		MBD5	NM_018328
B62 B63	chr2	188360514	A	G	q32.1		TFPI	NM_001032281
B62 B63	chr20	13918102	A	G	p12.1		SEL1L2	NM_001271539
B62 B63	chr20	30972541	G	C	q11.21		ASXL1	NM_015338
B62 B63	chr22	45313501	A	G	q13.31		PHF21B	NM_001135862
B62 B63	chr3	99546146	G	A	q12.1		CMSS1	NM_032359
B62 B63	chr4	1883359	A	G	p16.3		WHSC1	NM_001042424
B62 B63	chr4	102822941	A	T	q24		BANK1	NM_001083907
B62 B63	chr5	166938981	A	G	q34		TENM2	NM_001122679
B62 B63	chr6	64923693	C	T	q12	rs192052327	EYS	NM_001142800
B62 B63	chr6	163579077	G	A	q26		PACRG	NM_001080378
B62 B63	chr7	127503274	G	A	q32.1		SND1	NM_014390
B62 B63	chr8	62392971	G	A	q12.3		CLVS1	NM_173519
B62 B63	chrX	96306108	C	T	q21.33		DIAPH2	NM_006729
B62 B63	chrX	111417043	G	A	q23		ZCCHC16	NM_001004308
B62 B63	chrX	123121415	C	T	q25		STAG2	NM_001042749
B62 B63	chr2	47527796	G	A	p21		AC073283.4	NR_110207
B62 B63	chr1	81565925	A	G	q31.1			non-coding intronic
B62 B63	chr1	199011594	G	A	q32.1			
B62 B63	chr11	30391709	A	C	p14.1	rs66485912		
B62 B63	chr11	71038704	G	A	q13.4			
B62 B63	chr11	104908704	C	T	q22.3			
B62 B63	chr12	17246962	A	G	p12.3			
B62 B63	chr12	41970530	A	G	q12			
B62 B63	chr13	20923983	G	A	q12.11	rs114706614		
B62 B63	chr13	85569294	C	T	q31.1			
B62 B63	chr14	54397605	T	C	q22.2			
B62 B63	chr15	95452112	T	A	q26.2	rs199823837		
B62 B63	chr17	12179178	A	G	p12			
B62 B63	chr18	53310663	A	G	q21.2			
B62 B63	chr19	20630678	T	C	p12	rs201031945		
B62 B63	chr2	17073960	C	T	p24.2			
B62 B63	chr2	57552402	T	A	p16.1			
B62 B63	chr2	68293158	A	G	p14			
B62 B63	chr2	122634222	A	G	q14.3			
B62 B63	chr2	183452479	C	T	q32.1			
B62 B63	chr20	52707477	A	G	q13.2	rs371335191		
B62 B63	chr21	21885587	G	A	q21.1			
B62 B63	chr22	22003746	T	A	q11.21	rs73877639		
B62 B63	chr3	16887859	C	T	p24.3			
B62 B63	chr3	24933640	C	T	p24.2			
B62 B63	chr3	76252246	C	T	q12.3			
B62 B63	chr4	46434769	G	A	p12			
B62 B63	chr4	106901329	G	A	q24	rs115374603		
B62 B63	chr5	92461001	A	C	q15			
B62 B63	chr5	113042944	C	T	q22.2			
B62 B63	chr6	18908561	G	A	p22.3			
B62 B63	chr7	64318208	C	T	q11.21			
B62 B63	chr9	25628943	T	C	p21.2			
B62 B63	chrX	58380768	A	C	p11.1			
B62 B63	chrX	108826763	C	T	q23			

B62 B63	chrX	150711814	G	T	q28		intergenic		
B62 B63	chr5	90898204	TA	T		rs538545087	CTD-2281D19 ENST0000050536	intron	
B62 B63	chr8	133669256	TATG	T			LRRC6	CCDS6365.1	intron
B62 B63	chr12	41102610	TCTG	T			CNTN1	NM_001843	intron
B62 B63	chr9	71827884	CCTT	C		rs67229633	TJP2	CCDS6627.1	intron
B63	chr4	1331141	T	C	p16.3	rs28612202	MAEA	NM_001017405	intronic
B83	chr4	119231662	C	T	q26		PRSS12	NM_003619	intronic
B83	chr8	10745154	T	C	p23.1	rs7388643			
B83	chr8	16483051	T	C	p22				
B83	chr8	50040053	G	A	q11.21				
B83	chr8	94260080	G	T	q22.1				
B83 B84	chr19	4796824	G	A	p13.3		FEM1A	NM_018708	3downstream
B83 B84	chr2	209224680	C	T	q34		PIKFYVE	NM_015040	3downstream
B83 B84	chr20	9966603	G	A	p12.2		LOC10192937	NR_109861	5upstream
B83 B84	chr1	150338972	A	G	q21.3		RPRD2	NM_015203	intronic
B83 B84	chr10	21149815	G	C	p12.31		NEBL	NM_001173484	intronic
B83 B84	chr10	76753997	G	A	q22.2		KAT6B	NM_001256468	intronic
B83 B84	chr10	118876496	A	G	q25.3		KIAA1598	NM_001258300	intronic
B83 B84	chr12	21661853	A	C	p12.1	rs11046094	GOLT1B	NM_016072	intronic
B83 B84	chr15	48506290	T	C	q21.1		SLC12A1	NM_000338	intronic
B83 B84	chr15	75719027	T	C	q24.2		SIN3A	NM_001145357	intronic
B83 B84	chr19	4421191	C	T	p13.3		CHAF1A	NM_005483	intronic
B83 B84	chr2	137786478	C	T	q22.1	rs72985565	THSD7B	NM_001080427	intronic
B83 B84	chr3	53325297	C	T	p21.1		DCP1A	NM_001290204	intronic
B83 B84	chr4	57295012	G	A	q12		PPAT	NM_002703	intronic
B83 B84	chr4	128589038	T	A	q28.1		INTU	NM_015693	intronic
B83 B84	chr5	11250985	G	A	p15.2		CTNND2	NM_001288715	intronic
B83 B84	chr5	53365411	A	C	q11.2	rs10067481	ARL15	NM_019087	intronic
B83 B84	chr6	40413089	G	A	p21.2		LRFN2	NM_020737	intronic
B83 B84	chr7	16224899	A	G	p21.2		ISPD	NM_001101417	intronic
B83 B84	chr8	125660326	T	C	q24.13		MTSS1	NM_001282971	intronic
B83 B84	chr9	139573616	C	T	q34.3		AGPAT2	NM_001012727	intronic
B83 B84	chr14	30014118	A	G	q12		MIR548AI	NR_039672	non-coding
B83 B84	chr21	17868492	A	G	q21.1		LINC00478	NR_027790	non-coding
B83 B84	chr1	105309724	G	A	p21.1				intronic
B83 B84	chr1	218352394	A	G	q41				intronic
B83 B84	chr11	87164141	C	T	q14.2				intronic
B83 B84	chr12	66449758	T	C	q14.3				intronic
B83 B84	chr13	64665172	G	T	q21.31	rs269607			intronic
B83 B84	chr14	35915225	C	T	q13.2				intronic
B83 B84	chr14	43204114	A	G	q21.1				intronic
B83 B84	chr17	13734863	G	A	p12				intronic
B83 B84	chr17	31223286	C	T	q11.2				intronic
B83 B84	chr2	109609615	G	A	q12.3				intronic
B83 B84	chr2	172987977	C	T	q31.1	rs7572413			intronic
B83 B84	chr2	230145171	G	A	q36.3				intronic
B83 B84	chr21	16181595	A	G	q11.2				intronic
B83 B84	chr21	25258273	G	C	q21.2				intronic
B83 B84	chr3	31020364	A	G	p23				intronic
B83 B84	chr3	164228118	A	C	q26.1				intronic
B83 B84	chr3	189327482	C	T	q28				intronic
B83 B84	chr4	29361176	T	A	p15.1				intronic
B83 B84	chr4	132378202	C	T	q28.3				intronic
B83 B84	chr5	5609554	A	C	p15.32				intronic
B83 B84	chr5	54356988	C	G	q11.2				intronic
B83 B84	chr5	116550935	G	A	q23.1				intronic
B83 B84	chr7	138001305	T	C	q33				intronic
B83 B84	chr8	138597674	G	C	q24.23				intronic
B83 B84	chr9	82567879	T	C	q21.31				intronic
B83 B84	chr19	58613970	CAA	C			ZSCAN18	CCDS46214.1	intron
B84	chr13	45830708	C	T	q14.13		GTF2F2	NM_004128	intronic
B84	chr2	167878685	A	G	q24.3		XIRP2	NM_001079810	intronic
B84	chr6	149959476	T	A	q25.1		KATNA1	NM_001204076	intronic
B84	chr7	8681356	T	C	p21.3		NXPH1	NM_152745	intronic
B84	chr9	94497890	T	C	q22.31		ROR2	NM_004560	intronic
B84	chr4	36920104	A	C	p14				intronic
B84	chr17	41382052	T	C	q21.31	rs76863987	LINC00854	NR_047479	5upstream
B93 B94	chr7	135414085	A	G	q33		SLC13A4	NM_012450	5upstream
B93 B94	chr10	94297199	C	A	q23.33		IDE	NM_004969	syn
B93 B94	chr16	2147410	G	A	p13.3	rs374486955	PKD1	NM_000296	nonsyn p.R3438W
B93 B94	chr19	57037004	C	G	q13.43		ZNF471	NM_020813	nonsyn p.A523G
B93 B94	chr1	43646521	C	T	p34.2		WDR65	NM_001167965	intronic
B93 B94	chr1	186835728	A	G	q31.1		PLA2G4A	NM_024420	intronic
B93 B94	chr1	224422130	T	C	q42.11		NVL	NM_001243146	intronic
B93 B94	chr10	21979949	C	G	p12.31		MLLT10	NM_001195626	intronic
B93 B94	chr10	131456911	C	T	q26.3		MGMT	NM_002412	intronic

B93 B94	chr11	36428450	G	C	p12	PRR5L	NM_001160167	intronic
B93 B94	chr11	36428499	C	A	p12	PRR5L	NM_001160167	intronic
B93 B94	chr11	83191270	C	A	q14.1	DLG2	NM_001142699	intronic
B93 B94	chr11	131293441	T	C	q25	NTM	NM_001048209	intronic
B93 B94	chr15	39934086	G	A	q14	FSIP1	NM_152597	intronic
B93 B94	chr15	39986654	C	G	q14	FSIP1	NM_152597	intronic
B93 B94	chr16	81098864	G	A	q23.2	C16orf46	NM_001100873	intronic
B93 B94	chr17	49168444	T	C	q21.33	SPAG9	NM_001130527	intronic
B93 B94	chr17	54433363	A	G	q22	ANKFN1	NM_153228	intronic
B93 B94	chr17	54433372	T	C	q22	ANKFN1	NM_153228	intronic
B93 B94	chr17	58787185	G	A	q23.2	BCAS3	NM_001099432	intronic
B93 B94	chr17	65581782	G	A	q24.2	PITPN1C	NM_012417	intronic
B93 B94	chr18	72619462	C	T	q22.3	ZNF407	NM_001146189	intronic
B93 B94	chr2	32706159	A	G	p22.3	BIRC6	NM_016252	intronic
B93 B94	chr2	73229215	T	A	p13.2	SFXN5	NM_144579	intronic
B93 B94	chr2	182975989	A	T	q31.3	PPP1R1C	NM_001080545	intronic
B93 B94	chr2	241957475	G	A	q37.3	SNED1	NM_001080437	intronic
B93 B94	chr3	113657145	G	A	q13.31	GRAMD1C	NM_001172105	intronic
B93 B94	chr4	66240782	A	G	q13.1	EPHA5	NM_001281765	intronic
B93 B94	chr5	65445453	C	G	q12.3	SREK1	NM_001077199	intronic
B93 B94	chr5	65445884	C	G	q12.3	SREK1	NM_001077199	intronic
B93 B94	chr6	139472949	A	G	q24.1	HECA	NM_016217	intronic
B93 B94	chr6	146456876	G	A	q24.3	GRM1	NM_001278064	intronic
B93 B94	chr6	163212623	C	A	q26	PACRG	NM_001080378	intronic
B93 B94	chr7	17914832	C	A	p21.1	SNX13	NM_015132	intronic
B93 B94	chr7	97860074	G	A	q21.3	TECP1	NM_015395	intronic
B93 B94	chr7	132653469	G	A	q33	CHCHD3	NM_017812	intronic
B93 B94	chr8	22898794	T	C	p21.3	TNFRSF10B	NM_003842	intronic
B93 B94	chr9	77343121	T	A	q21.13	TRPM6	NM_001177310	intronic
B93 B94	chr6	21747135	C	T	p22.3	CASC15	NR_015410	non-coding intronic
B93 B94	chr1	64191122	G	A	p31.3			
B93 B94	chr1	151829054	G	C	q21.3			
B93 B94	chr10	72842217	A	G	q22.1			
B93 B94	chr11	90849241	A	G	q14.3			
B93 B94	chr11	116421503	A	T	q23.3			
B93 B94	chr12	20304160	A	G	p12.2			
B93 B94	chr13	79350867	A	G	q31.1			
B93 B94	chr14	64047343	A	G	q23.2			
B93 B94	chr15	50440752	G	T	q21.2			
B93 B94	chr15	70747621	C	G	q23 rs10220813			
B93 B94	chr18	508117	A	C	p11.32			
B93 B94	chr18	10325164	C	T	p11.22			
B93 B94	chr18	12057323	A	C	p11.21			
B93 B94	chr18	73917867	G	A	q23			
B93 B94	chr19	21062165	C	T	p12			
B93 B94	chr2	6263593	C	T	p25.2			
B93 B94	chr2	16519005	G	C	p24.3			
B93 B94	chr2	79194279	T	A	p12			
B93 B94	chr2	138486744	A	G	q22.1			
B93 B94	chr2	156799239	A	G	q24.1			
B93 B94	chr2	164338918	G	A	q24.3			
B93 B94	chr2	212168526	A	G	q34			
B93 B94	chr3	30351562	T	C	p24.1			
B93 B94	chr3	46151596	A	G	p21.31			
B93 B94	chr3	50574244	T	C	p21.31			
B93 B94	chr3	70268930	A	C	p13			
B93 B94	chr3	88598299	A	G	p11.1			
B93 B94	chr4	5962283	A	G	p16.2			
B93 B94	chr4	103002276	C	T	q24			
B93 B94	chr4	154787026	A	G	q31.3			
B93 B94	chr5	3908422	C	T	p15.33			
B93 B94	chr6	165600454	T	C	q27			
B93 B94	chr7	19081637	C	A	p21.1			
B93 B94	chr7	85163682	G	A	q21.11			
B93 B94	chr7	103763484	C	T	q22.1			
B93 B94	chr8	41046737	A	T	p11.21			
B93 B94	chr9	25091094	G	C	p21.3			
B93 B94	chr9	89456759	C	T	q21.33			
B93 B94	chrX	55365659	C	A	p11.21			
B93 B94	chrX	55595693	C	A	p11.21			
B93 B94	chr11	72159268	CT	C	rs141487620			
B93 B94	chr16	81278499	TG	T		BCMO1	CCDS10934.1	intron
LJ	chr6	12749681	C	G	p24.1	PHACTR1	NM_001242648	intronic
LJ	chr19	3354318	T	C	p13.3			
LJ LS	chr1	7523207	G	A	p36.23	CAMTA1	NM_015215	intronic
LJ LS	chr1	118539438	T	C	p12	SPAG17	NM_206996	intronic

LJ LS	chr10	24822488	T	C	p12.1	KIAA1217	NM_001098500	intronic	
LJ LS	chr10	53069364	T	C	q21.1	PRKG1	NM_001098512	intronic	
LJ LS	chr10	64995156	A	G	q21.3	JMJD1C	NM_001282948	intronic	
LJ LS	chr10	118484331	C	T	q25.3	HSPA12A	NM_025015	intronic	
LJ LS	chr11	84078617	G	T	q14.1	DLG2	NM_001142699	intronic	
LJ LS	chr11	100687399	G	A	q22.1	ARHGAP42	NM_152432	intronic	
LJ LS	chr16	71264288	C	T	q22.2	HYDIN	NM_001198542	intronic	
LJ LS	chr19	8468273	C	T	p13.2	RAB11B	NM_004218	intronic	
LJ LS	chr19	57113990	G	T	q13.43	ZNF71	NM_021216	intronic	
LJ LS	chr2	167170722	C	T	q24.3	SCN9A	NM_002977	intronic	
LJ LS	chr3	120315393	G	A	q13.33	NDUFB4	NM_001168331	intronic	
LJ LS	chr4	39119053	G	T	p14	KLHL5	NM_001007075	intronic	
LJ LS	chr4	115909880	T	A	q26	NDST4	NM_022569	intronic	
LJ LS	chr4	115909881	C	A	q26	NDST4	NM_022569	intronic	
LJ LS	chr6	37921491	G	A	p21.2	ZFAND3	NM_021943	intronic	
LJ LS	chr6	124510527	C	T	q22.31	NKAIN2	NM_001040214	intronic	
LJ LS	chr6	124515065	A	T	q22.31	NKAIN2	NM_001040214	intronic	
LJ LS	chr7	20427755	C	T	p21.1	ITGB8	NM_002214	intronic	
LJ LS	chr7	50434049	G	A	p12.2	IKZF1	NM_001220765	intronic	
LJ LS	chr8	3381974	C	A	p23.2	CSMD1	NM_033225	intronic	
LJ LS	chr9	105381829	C	T	q31.1	LINC00587	NR_103830	non-coding intronic	
LJ LS	chr1	79265388	C	T	p31.1				
LJ LS	chr10	73143664	G	A	q22.1				
LJ LS	chr11	97265884	G	C	q22.1				
LJ LS	chr12	105193487	A	G	q23.3	rs77594953			
LJ LS	chr13	73798380	G	A	q22.1				
LJ LS	chr14	39073589	G	C	q21.1				
LJ LS	chr16	47797029	T	C	q12.1				
LJ LS	chr16	76707882	C	T	q23.1				
LJ LS	chr17	13733612	C	T	p12				
LJ LS	chr18	38732423	A	G	q12.3				
LJ LS	chr2	146609689	G	A	q22.3				
LJ LS	chr2	184856690	T	C	q32.1				
LJ LS	chr2	186167711	T	G	q32.1				
LJ LS	chr2	186167712	A	T	q32.1				
LJ LS	chr2	218660590	C	G	q35				
LJ LS	chr2	238513142	C	T	q37.3	rs59237394			
LJ LS	chr20	50688780	G	T	q13.2				
LJ LS	chr3	36418949	C	T	p22.3				
LJ LS	chr3	155766601	G	A	q25.31				
LJ LS	chr4	27144101	G	A	p15.2				
LJ LS	chr4	70243771	G	C	q13.2				
LJ LS	chr4	129175380	T	G	q28.2				
LJ LS	chr7	36863766	C	T	p14.2				
LJ LS	chr7	54008241	C	T	p11.2				
LJ LS	chr9	15398550	G	C	p22.3				
LJ LS	chr9	134281048	C	T	q34.13				
LJ LS	chrY	17459241	A	G	q11.221				
LJ LS	chrX	93019398	CAG	C					
LS	chr12	123034737	A	T	q24.31	rs12581072	KNTC1	NM_014708	intronic
LS	chr2	174321128	C	G	q31.1	rs13021431			

Supplemental Table S7. The regulatory elements hit by *de novo* SNVs in 8 pairs of MZ twins

Sample	Chromosome	chromPosition	Allele1	Allele2	Transition/Transversion	Gene	Predicted function	CpG_site	CpG_island	Score*	Regulatory elements hits
B93 B94	chr15	50440752	G	T	transversion	intergenic		CpG_site	NA	2a	Motifs PWM Stra13, Motifs Footprinting AosmcSerumfree Stra13, Motifs Footprinting AosmcSerumfree USF, Motifs Footprinting Mcf7Hypoxlac Stra13, Motifs Footprinting A549 Stra13, Motifs Footprinting Mcf7 USF, Motifs Footprinting Mcf7Hypoxlac USF, Motifs Footprinting H9es Stra13, Motifs Footprinting A549 Motifs Footprinting SkMC , Motifs Footprinting K562 , Motifs PWM CREB3L1, Motifs PWM USF, Motifs Footprinting Mcf7 Stra13, Motifs Footprinting A549 USF, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Nb4, Chromatin_Structure Diffa14d DNase- seq H7es, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase- seq Htr8, Chromatin_Structure DNase-seq Hpaec, Chromatin_Structure DNase- seq Hpdlf, Chromatin_Structure Znfb34a8 DNase-seq K562, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase- seq Hmvecdneo, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Hrpe, Chromatin_Structure Znfe103c6 DNase-seq K562, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase- seq Ag09319, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Prec, Chromatin_Structure DNase- seq Fibroblgm03348, Chromatin_Structure Est100nm1h DNase-seq Mcf7, Chromatin_Structure Znf4c50c4 DNase-seq K562, Chromatin_Structure DNase- seq Hpe, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure DNase- seq Rptec, Chromatin_Structure DNase-seq Gm12864, Chromatin_Structure DNase-seq M059j, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase- seq K562G2mphase, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure DNase-seq Hs5, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Hmveclbl, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure Ohurea FAIRE K562, Chromatin_Structure DNase-seq Hvfm, Chromatin_Structure DNase-seq Hrgec, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Hmvecdlyneo, Chromatin_Structure DNase-

B93 B94	chr16	2147410	G	A	transition	PKD1	nonsyn p.R3438W	CpG_site	NA	2b	Motifs Footprinting Huh75 TLX1::NFIC, Motifs Footprinting AosmcSerumfree TLX1::NFIC, Motifs Footprinting Melano CTF1, Motifs Footprinting Myometr myogenin/NF-1, Motifs Footprinting H1hesc TLX1::NFIC, Motifs Footprinting Htr8 CTF1, Motifs Footprinting Panislets CTF1, Motifs Footprinting Myometr CTF1, Motifs Footprinting Gm19239 CTF1, Motifs Footprinting Osteobl CTF1, Motifs Footprinting Phte TLX1::NFIC, Motifs PWM myogenin/NF-1, Motifs Footprinting Mcf7 TLX1::NFIC, Motifs Footprinting Gm12891 myogenin/NF-1, Motifs Footprinting Chorion CTF1, Motifs Footprinting Gm18507 TLX1::NFIC, Motifs Footprinting K562 TLX1::NFIC, Motifs Footprinting Panislets myogenin/NF-1, Motifs Footprinting Gm12892 myogenin/NF-1, Motifs Footprinting Hmec TLX1::NFIC, Motifs Footprinting Fibrop TLX1::NFIC, Motifs Footprinting A549 CTF1, Motifs Footprinting H1hesc CTF1, Motifs Footprinting Fibrobl TLX1::NFIC, Motifs Footprinting Gm18507 CTF1, Motifs Footprinting Panislets TLX1::NFIC, Motifs Footprinting Mcf7Hypoxlac TLX1::NFIC, Motifs Footprinting Huvec CTF1, Motifs Footprinting Gm12892 TLX1::NFIC, Motifs Footprinting Huh7 TLX1::NFIC, Motifs Footprinting Lncap myogenin/NF-1, Motifs Footprinting Huh75 CTF1, Motifs Footprinting Mcf7 CTF1, Motifs Footprinting Hepatocytes CTF1, Motifs Footprinting Hsmmm myogenin/NF-1, Motifs Footprinting Helas3 TLX1::NFIC, Motifs Footprinting Chorion TLX1::NFIC, Motifs Footprinting Gliobla CTF1, Motifs Footprinting AosmcSerumfree CTF1, Motifs Footprinting Hpde6e6e7 CTF1, Motifs Footprinting Hepg2 TLX1::NFIC, Motifs PWM NFIIX, Motifs Footprinting LncapAndro myogenin/NF-1, Motifs Footprinting Gm19239 TLX1::NFIC, Motifs Footprinting Medullo TLX1::NFIC, Motifs Footprinting Gm12892 CTF1, Motifs Footprinting Lncap CTF1, Motifs Footprinting Cll TLX1::NFIC, Motifs Footprinting Hsmmt myogenin/NF-1, Motifs Footprinting Hsmmm TLX1::NFIC, Motifs Footprinting A549 TLX1::NFIC, Motifs Footprinting Myometr TLX1::NFIC, Motifs Footprinting Mcf7Hypoxlac CTF1, Motifs Footprinting Hmec CTF1, Motifs PWM CTF1, Motifs Footprinting 8988t myogenin/NF-1, Motifs PWM NFIIB, Motifs Footprinting Hepg2 CTF1, Motifs Footprinting Phte CTF1, Motifs Footprinting Huh7 CTF1,
---------	-------	---------	---	---	------------	------	-----------------	----------	----	----	---

B83	B84	chr19	4796824	G	A	transition	FEM1A	3downstream	CpG_site	NA	2b	Motifs Footprinting Medullo Olf-1, Motifs PWM CAC-bindingprotein, Motifs Footprinting H1hesc Olf-1, Motifs Footprinting Gm12891 Olf-1, Motifs PWM Olf-1, Motifs Footprinting Gm12892 Olf-1, Chromatin_Structure DNase-seq Hrpe, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Skmc, Chromatin_Structure DNase-seq Osteob1, Chromatin_Structure Diff4d DNase-seq Lhcnm2, Chromatin_Structure DNase-seq Lhcnm2, Chromatin_Structure DNase-seq Medullo, Protein_Binding ChIP-seq GM12878 POLR2A, Protein_Binding ChIP-seq K562 POLR2A, Protein_Binding ChIP-seq GM12892 POLR2A, Protein_Binding ChIP-
B83	B84	chr20	9966603	G	A	transition	LOC101929371	5upstream	NA	NA	2b	Motifs PWM IRF-7, Motifs Footprinting Hpde6e6e7 IRF-7, Chromatin_Structure DNase-seq Saec, Chromatin_Structure Estctrl0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure Est100nm1h DNase-seq Mcf7, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hvmf, Chromatin_Structure DNase-seq Hcm, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Hpdlf, Chromatin_Structure DNase-seq Hee, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Skmc, Chromatin_Structure DNase-seq Hgf, Protein_Binding 01pct ChIP-seq MCF10A-Er-Src STAT3, Protein_Binding 01pct ChIP-seq MCF10A-Er-Src MYC, Protein_Binding 4ohtam_1um_4hr ChIP-seq MCF10A-Er-Src FOS, Protein_Binding 01pct_4hr ChIP-seq MCF10A-Er-Src STAT3, Protein_Binding 4ohtam_1um_36hr ChIP-seq MCF10A-Er-Src STAT3 Motifs Footprinting Hepg2 STAT5A(homotetramer), Motifs PWM STAT5A(homotetramer), Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure FAIRE Hepg2, Protein_Binding forskolin ChIP-seq HepG2 POLR2A, Protein_Binding ChIP-seq HepG2 HDAC2, Protein_Binding ChIP-seq HepG2 SP1, Protein_Binding ChIP-seq HepG2 HNF4A, Protein_Binding ChIP-seq HepG2 YY1, Protein_Binding forskolin ChIP-seq HepG2 HNF4A, Protein_Binding ChIP-seq NB4 MYC, Protein_Binding ChIP-seq HepG2 HNF4G, Protein_Binding ChIP-seq HepG2 POLR2A,
B62		chr7	27657582	G	A	transition	HIBADH	intronic	CpG_site	NA	2b	Motifs Footprinting Hepg2 STAT5A(homotetramer), Motifs PWM STAT5A(homotetramer), Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure FAIRE Hepg2, Protein_Binding forskolin ChIP-seq HepG2 POLR2A, Protein_Binding ChIP-seq HepG2 HDAC2, Protein_Binding ChIP-seq HepG2 SP1, Protein_Binding ChIP-seq HepG2 HNF4A, Protein_Binding ChIP-seq HepG2 YY1, Protein_Binding forskolin ChIP-seq HepG2 HNF4A, Protein_Binding ChIP-seq NB4 MYC, Protein_Binding ChIP-seq HepG2 HNF4G, Protein_Binding ChIP-seq HepG2 POLR2A,

B53 B54	chr17	41382052	T	C	transition	LINC00854	5upstream	NA	NA	2b	Motifs Footprinting K562, Motifs PWM EBF1, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Hpdif, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure FAIRE UrotsaUt189, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Cd4naivewb11970640, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure fng4h FAIRE Helas3, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure FAIRE Gm12878, Chromatin_Structure FAIRE Nhbe, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure FAIRE Htr8, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure Ohurea FAIRE K562, Chromatin_Structure DNase-seq Hrgec, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure Ctfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Medullo, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Th2, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Tregwb78495824, Chromatin_Structure DNase-seq Cd4naivewb78495824, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq Saec,
---------	-------	----------	---	---	------------	-----------	-----------	----	----	----	---

B84	chr17	41382052	T	C	transition	LINC00854	5upstream	NA	NA	2b	Motifs Footprinting K562, Motifs PWM EBF1, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Hpdif, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure FAIRE UrotsaUt189, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Cd4naivewb11970640, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure fng4h FAIRE Helas3, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure FAIRE Gm12878, Chromatin_Structure FAIRE Nhbe, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure FAIRE Htr8, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure Ohurea FAIRE K562, Chromatin_Structure DNase-seq Hrgec, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure Ctfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Medullo, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Th2, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Tregwb78495824, Chromatin_Structure DNase-seq Cd4naivewb78495824, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq Saec,
-----	-------	----------	---	---	------------	-----------	-----------	----	----	----	---

B103	B104	chr18	57721409	A	G	transition	intergenic	NA	NA	2b	Motifs Footprinting Hpde6e6e7 Foxq1, Motifs Footprinting AosmcSerumfree Foxq1, Motifs Footprinting Phte Foxq1, Motifs PWM Foxq1, Motifs Footprinting Helas3 Ifna4h Foxq1, Motifs Footprinting Helas3 Foxq1, Motifs Footprinting Panisd Foxq1, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure Ctcfshrna DNase-seq Mcf7, Chromatin_Structure FAIRE Medullo, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure FAIRE Gliobla, Chromatin_Structure DNase-seq K562, Chromatin_Structure FAIRE Helas3, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase- seq Helas3, Chromatin_Structure DNase-seq Nha, Chromatin_Structure FAIRE K562, Chromatin_Structure DNase-seq Phte, Chromatin_Structure DNase-seq Lhcnm2, Chromatin_Structure DNase- seq Hsmmemb, Chromatin_Structure DNase-seq Fibropag20443, Chromatin_Structure DNase-seq Nhlf, Chromatin_Structure Znfg54a11 DNase- seq K562, Chromatin_Structure DNase-seq Aoaf, Protein_Binding ChIP-seq HeLa-
------	------	-------	----------	---	---	------------	------------	----	----	----	---

B53	B54	chr15	68647297	T	A	transversion	ITGA11	intronic	NA	NA	2b	Motifs Footprinting Helas3 Ifna4h Ik-1, Motifs Footprinting Fibrop Ik-1, Motifs Footprinting Hpde6e6e7 Ik-1, Motifs Footprinting AosmcSerumfree Ik-1, Motifs PWM Ik-1, Motifs Footprinting Htr8 Ik-1, Motifs Footprinting Hsmm Ik-1, Motifs Footprinting Fibrobl Ik-1, Motifs Footprinting Hmec Ik-1, Motifs Footprinting Panisd Ik-1, Motifs Footprinting 8988t Ik-1, Motifs Footprinting Gliobla Ik-1, Motifs Footprinting A549 Ik-1, Motifs Footprinting Myometr Ik-1, Motifs Footprinting Huvec Ik-1, Motifs Footprinting Helas3 Ik-1, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Ipsnih7, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure FAIRE UrotsaUt189, Chromatin_Structure Znfe103c6 DNase-seq K562, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure Est100nm1h DNase-seq Mcf7, Chromatin_Structure FAIRE Nhbe, Chromatin_Structure DNase-seq Hs5, Chromatin_Structure DNase-seq Hmvecad, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure Serumfree DNase-seq Aosmc, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure Lenticon DNase-seq Fibroblgm03348, Chromatin_Structure Ifna4h FAIRE Helas3, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq K562, Chromatin_Structure FAIRE Urotsa, Chromatin_Structure DNase-seq Hl60, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Nhek, Chromatin_Structure DNase-seq Gliobla, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure Lentimyod DNase-seq Fibroblgm03348, Chromatin_Structure FAIRE Astrocy, Chromatin_Structure DNase-seq Hsmmfshd, Chromatin_Structure DNase-seq Hsmmemb, Chromatin_Structure DNase-seq SkMC, Motifs PWM RREB1, Motifs Footprinting AosmcSerumfree RREB1, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Ag09319, Chromatin_Structure DNase-seq Rptec, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Hgf, Protein_Binding ChIP-seq AG04450 CTCF, Protein_Binding 4ohtam_1um_12hr ChIP-seq MCF10A-Er-Src STAT3,
B103	B104	chr16	69210370	A	C	transversion		intergenic	NA	NA	2b	

B103	B104	chr16	69210371	C	A	transversion	intergenic	NA	NA	2b	Motifs Footprinting SkMC, Motifs PWM RREB1, Motifs Footprinting AosmcSerumfree RREB1, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Ag09319, Chromatin_Structure DNase-seq Rptec, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Hgf, Protein_Binding ChIP-seq AG04450 CTCF, Protein_Binding 4ohtam_1um_12hr ChIP-seq MCF10A-Er-Src STAT3,
B83	B84	chr8	125660326	T	C	transition	MTSS1	intronic	NA	NA	2b Motifs Footprinting Mcf7 IPF1, Motifs Footprinting Htr8 IPF1, Motifs PWM IPF1, Motifs Footprinting Gm12878 IPF1, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure FAIRE Htr8, Protein_Binding ChIP-seq HFF-Myc CTCF, Protein_Binding ChIP-seq Octachl CTCF_Protein_Binding ChIP-seq K562 CTCF

B123	B124	chr12	129314669	A	G	transition	intergenic	NA	NA	2b	Motifs Footprinting Gm18507 Pax-8, Motifs Footprinting Cll Pax-8, Motifs Footprinting Gm12878 Pax-8, Motifs PWM Pax-8, Motifs Footprinting Medullo Pax-8, Motifs PWM Duxl, Motifs Footprinting Gm19238 Pax-8, Motifs Footprinting Gm12892 Pax-8, Motifs Footprinting Gm12891 Pax-8, Motifs Footprinting Hsmmt Pax-8, Motifs PWM FOXJ2, Motifs Footprinting Hsmmt Pax-8, Motifs Footprinting Gm19239 Pax-8, Chromatin_Structure DNase-seq Nb4, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase- seq Cd20ro01794, Chromatin_Structure DNase-seq Prec, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure FAIRE Gm12878, Chromatin_Structure DNase- seq Cd34mobilized, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase- seq Wi38, Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase- seq Gm10248, Chromatin_Structure FAIRE Gm18507, Chromatin_Structure DNase-seq Th2, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase- seq Gm20000, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase- seq Hmf, Chromatin_Structure DNase-seq Skmc, Chromatin_Structure DNase- seq Th2wb54553204, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure DNase- seq Fibrobl, Chromatin_Structure DNase-seq Th1wb33676984, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase- seq Gbcell, Chromatin_Structure DNase-seq Werirb1, Chromatin_Structure DNase-seq Cll, Chromatin_Structure DNase-seq Hl60, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase- seq T47d, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase- seq Gm19238, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure DNase- seq Gm19239, Chromatin_Structure DNase-seq Naivecell, Chromatin_Structure DNase-seq Hee, Chromatin_Structure Ohtam DNase- seq Wi38, Chromatin_Structure Diff4d DNase-seq Lhcnm2, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure Lentimyod DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Lhcnm2, Chromatin_Structure DNase-
------	------	-------	-----------	---	---	------------	------------	----	----	----	--

B93	B94	chr10	131456911	C	T	transition	MGMT	intronic	NA	NA	2b	Motifs Footprinting Huh7 Gfi1, Motifs Footprinting Mcf7 Gfi1, Motifs Footprinting Huh7 Gfi1b, Motifs PWM Gfi1b, Motifs Footprinting Mcf7 Gfi-1, Motifs PWM Gfi1, Motifs PWM Gfi, Motifs Footprinting Huh7 Gfi, Motifs Footprinting Cll Gfi1, Motifs Footprinting Cll Gfi1b, Motifs PWM Gfi-1, Motifs Footprinting Mcf7 Gfi-1, Motifs Footprinting Mcf7 Gfi1b, Motifs Footprinting Huh7 Gfi-1, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Ipsnih17, Chromatin_Structure DNase-seq Ag09319, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Nhdfad, Chromatin_Structure DNase-seq Hgf,
B124		chr1	230530315	T	C	transition	PGBD5	intronic	NA	NA	2b	Motifs PWM SP4, Motifs Footprinting A549 SP1, Motifs PWM SP1, Motifs PWM Zfp410, Motifs PWM Bcl6b, Motifs PWM ZNF219, Motifs Footprinting Lncap ZNF219, Motifs PWM Sp1, Motifs Footprinting Lncap CAC-bindingprotein, Motifs PWM Zfp740, Motifs PWM CAC-bindingprotein, Motifs PWM Zfp281, Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq Ipsnih11, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Ipsnih17, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure Sahactrl DNase-seq K562, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-
B103	B104	chr4	125864692	C	T	transition		intergenic	CpG_site	NA	2c	Motifs PWM IRF-1, Motifs PWM IRF1, Motifs PWM ICSBP, Motifs PWM IRF-2, Motifs PWM IRF2, Motifs PWM IRF, Motifs PWM ISGF-3, Motifs PWM STAT1, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure Hypoxlac FAIRE Mcf7, Chromatin_Structure FAIRE Gliobla, Protein_Binding ChIP-seq MCF-7 ZNF217, Protein_Binding ChIP-seq HeLa-S3 CHD2, Protein_Binding ChIP-seq HeLa-S3 JUN, Protein_Binding ChIP-seq HeLa-S3 STAT3, Protein_Binding ChIP-seq HeLa-S3 MAX, Protein_Binding ChIP-seq SK-N-SH SIN3A, Protein_Binding ChIP-seq A549 CEPB, Protein_Binding ChIP-seq HeLa-S3 MYC, Protein_Binding ChIP-seq HeLa-S3 RFX5, Protein_Binding fng30 ChIP-

LJ	chr6	12749681	C	G	transversion	PHACTR1	intronic	NA	NA	3a	Motifs PWM Irf3, Chromatin_Structure DNase-seq Ipsnih17, Chromatin_Structure DNase-seq Cd20ro1794, Chromatin_Structure DNase-seq Ipsnih11, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure Lenticon DNase-seq Fibroblgm03348, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure DNase-seq Hff, Chromatin_Structure Diffa9d DNase-seq H7es, Chromatin_Structure DNase-seq Werirb1, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Cerebellumoc, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq H9es, Chromatin_Structure Diff4d DNase-seq Lhcnm2, Chromatin_Structure Lentimyod DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Lhcnm2, Chromatin_Structure DNase-seq Hsmmfshd, Chromatin_Structure DNase-seq Medullo, Protein_Binding ChIP-seq H1-hESC TAF1, Protein_Binding ChIP-seq H1-hESC REST, Protein_Binding ChIP-seq H1-hESC RFX5, Protein_Binding ChIP-seq H1-hESC SP4, Protein_Binding ChIP-seq H1-hESC YY1, Protein_Binding ChIP-seq H1-hESC SIN3A, Protein_Binding ChIP-seq H1-hESC RC1_11A
LJ LS	chr7	20427755	C	T	transition	ITGB8	intronic	NA	NA	3a	Motifs PWM Zfp281, Chromatin_Structure DNase-seq Nhdneo, Chromatin_Structure DNase-seq Sknmc, Protein_Binding ChIP-seq SK-N-MC POI_R2A
B103 B104	chr2	21646773	C	T	transition		intergenic	NA	NA	3a	Motifs PWM MYF, Motifs PWM Myf, Motifs PWM NeuroD, Motifs PWM Ascl2, Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Mel2183, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Urotsa, Protein_Binding ChIP-seq HegG2IMAEK

B123	B124	chr8	23211573	T	G	transversion	LOXL2	intronic	NA	NA	3a	Motifs PWM Pou6f1, Motifs PWM Sox15, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure Diffa14d DNase-seq H7es, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Hpaec, Chromatin_Structure DNase-seq Hpdlf, Chromatin_Structure DNase-seq Hmvecdneo, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Hrpe, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq Ag09319, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Prec, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure Est100nm1h DNase-seq Mcf7, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hepe, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure FAIRE Nhbe, Chromatin_Structure DNase-seq M059j, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure DNase-seq Hs5, Chromatin_Structure DNase-seq Hmvecdad, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Hmveclbl, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure ra DNase-seq Sknsh, Chromatin_Structure DNase-seq Hvmf, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq Hrgec, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure Ctfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Hmvecdlyneo, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure Estctrl0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Medullod341, Chromatin_Structure Serumfree DNase-seq Aosmc, Chromatin_Structure DNase-seq Ag04450, Chromatin_Structure DNase-seq Hmvecdlneo, Chromatin_Structure DNase-seq Hct116, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Hsmm, Motifs PWM MAFK, Motifs PWM Mafb, Motifs PWM NRL, Motifs PWM POU6F2, Chromatin_Structure Serumfree DNase-seq Aosmc, Protein_Binding ChIP-seq Hela-S3 EP300
B113	B114	chr11	27959616	C	G	transversion		intergenic	NA	NA	3a	Motifs PWM MAFK, Motifs PWM Mafb, Motifs PWM NRL, Motifs PWM POU6F2, Chromatin_Structure Serumfree DNase-seq Aosmc, Protein_Binding ChIP-seq Hela-S3 EP300
B103	B104	chr20	34319685	T	G	transversion	RBM39	intronic	NA	NA	3a	Motifs PWM FOXC1, Motifs PWM Nkx3-1, Motifs PWM Cdx, Chromatin_Structure Hypoxlac FAIRE Mcf7, Chromatin_Structure DNase-seq Nhek, Protein_Binding ChIP-seq H1-hESC POLR2A, Protein_Binding ChIP-seq HepG2 POI_R2A
LJ LS		chr4	39119053	G	T	transversion	KLHL5	intronic	NA	NA	3a	Motifs PWM Zfp410, Motifs PWM Rxra, Motifs PWM MZF1, Chromatin_Structure DNase-seq T47d, Protein_Binding 02pct ChIP-seq T-47D GATA3, Protein_Binding 02nct ChIP-seq T-47D FOXA1

B53	B54	chr17	41382051	T	C	transition	LINC00854	5upstream	NA	NA	3a	Motifs PWM EBF1, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Hpdlf, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure FAIRE UrotsaUt189, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Cd4naivewb11970640, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure fng4h FAIRE Helas3, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure FAIRE Gm12878, Chromatin_Structure FAIRE Nhbe, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure FAIRE Htr8, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure Ohurea FAIRE K562, Chromatin_Structure DNase-seq Hrgec, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure Ctfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Medullo, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Th2, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Tregwb78495824, Chromatin_Structure DNase-seq Cd4naivewb78495824, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq Saec, Motifs PWM Srf, Motifs PWM FOXP1, Motifs PWM Zfp105, Motifs PWM Mtf1, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase-seq Helas3, Protein_Binding ChIP-seq T2OSISFTDR1, Protein_Binding O2mtl ChIP-seq A549 VV1
B103	B104	chr17	41633492	A	C	transversion		intergenic	NA	NA	3a	

B103	B104	chr10	71169085	T	G	transversion	TACR2	intronic	NA	NA	3a	Motifs PWM MAX, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq K562G1phase, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure DNase-seq Cerebellumoc, Chromatin_Structure DNase-seq Sknsn, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Hs27a, Chromatin_Structure DNase-seq Th1wb54553204, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase-seq Psoasmuscleoc, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Hs5, Chromatin_Structure DNase-seq Urotsa, Protein_Binding ChIP-seq K562 POLL2A, Protein_Binding fng6h ChIP-seq K562 MYC, Protein_Binding fng6h ChIP-seq K562 POLL2A, Protein_Binding fng6h ChIP-seq K562 MYC
B83	B84	chr2	109609615	G	A	transition		intergenic	NA	NA	3a	Motifs PWM Bcl6b, Chromatin_Structure DNase-seq Osteobl, Protein_Binding differential ChIP-seq Caco2 HNF4A, Protein_Binding proliferation ChIP-seq Caco2 HNF4A
B53	B54	chr5	118144221	T	C	transition		intergenic	NA	NA	3a	Motifs PWM FOXM1, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure DNase-seq H7es, Chromatin_Structure Diffa9d DNase-seq H7es. Protein_Binding ChIP-seq HepG2 MAFK
B83	B84	chr4	128589038	T	A	transversion	INTU	intronic	NA	NA	3a	Motifs PWM Foxk1, Motifs PWM Foxj3, Motifs PWM Glis2, Motifs PWM TBP, Motifs PWM Tbp, Motifs PWM FOXC2, Motifs PWM Fox11, Motifs PWM FOXP1, Motifs PWM Foxa2, Chromatin_Structure DNase-seq Sknmc, Protein_Binding ChIP-seq SK-N-MC POLR2A
B62	B63	chr2	183452479	C	T	transition		intergenic	NA	NA	3a	Motifs PWM NF-kappaB, Chromatin_Structure DNase-seq Hah, Chromatin_Structure FAIRE Panislets, Chromatin_Structure DNase-seq Sknmc, Chromatin_Structure FAIRE Gliobla. Protein_Binding ChIP-seq K562 SPI1
B62	B63	chr1	199011594	G	A	transition		intergenic	NA	NA	3a	Motifs PWM Zfp187, Motifs PWM Nkx3-1, Chromatin_Structure DNase-seq Hl60, Protein_Binding ChIP-seq IMR90 CEPB, Protein_Binding ChIP-seq HenG2 CEBPP

B63	chr4	1331141	T	C	transition	MAEA	intronic	NA	NA	4	Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase-seq Hrpe, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Gliobla, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq Hmveclbl, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Phte, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq Myometr, Chromatin_Structure DNase-seq Hmvecdblneo, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Hsmmm, Chromatin_Structure DNase-seq Psoriasismuscleoc, Chromatin_Structure DNase-seq Panislets, Protein_Binding ChIP-seq HUVEC FOS, Protein_Binding ChIP-seq GM12878 BCL3,
B83 B84	chr5	5609554	A	C	transversion	intergenic		NA	NA	4	Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Th1, Protein_Binding ChIP-seq K562 RFX3
B113 B114	chr2	10036253	C	T	transition	TAF1B	intronic	CpG_site	NA	4	Chromatin_Structure FAIRE Panislets, Chromatin_Structure FAIRE Medullo, Chromatin_Structure DNase-seq Mel2183, Chromatin_Structure DNase-seq Colo829, Chromatin_Structure FAIRE Hepg2, Chromatin_Structure FAIRE Nhek, Chromatin_Structure FAIRE Gliobla, Protein_Binding ChIP-seq MCF10Δ_Er_5α DOI_R2Δ, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gcbcell, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Medullo, Protein_Binding ChIP-seq GM12878 QP1
B83	chr8	10745154	T	C	transition	intergenic		NA	NA	4	Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gcbcell, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Medullo, Protein_Binding ChIP-seq MCF10Δ_Er_5α DOI_R2Δ, Chromatin_Structure DNase-seq L02pct ChIP-seq GATA3
B62 B63	chr6	18908561	G	A	transition	intergenic		NA	NA	4	Chromatin_Structure Estctrl0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq T47d, Protein_Binding 02pct ChIP-seq T-47D GATA3

B103	B104	chrX	20489848	T	C	transition	intergenic	NA	NA	4	Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Mel2183, Chromatin_Structure DNase-seq Colo829, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure FAIRE Hepg2, Chromatin_Structure DNase-seq Rpmi7951, Protein_Binding ChIP-seq HepG2 TCF7L2, Protein_Binding ChIP-seq HepG2 MBD4, Protein_Binding ChIP-seq HepG2 FOXA1, Protein_Binding ChIP-seq HepG2 HDAC2, Protein_Binding ChIP-seq HepG2 EP300, Protein_Binding ChIP-seq HepG2 RXRA, Protein_Binding ChIP-seq HepG2 NFIC, Protein_Binding ChIP-seq HepG2 SP1, Protein_Binding ChIP-seq HepG2 TEAD4, Protein_Binding ChIP-seq HepG2 HNF4A, Protein_Binding ChIP-seq VCaP AR, Protein_Binding ChIP-seq HepG2 ARID3A, Protein_Binding ChIP-seq HepG2 CEPB, Protein_Binding ChIP-seq HepG2 HNF4G. Protein_Binding ChIP-seq HepG2 SMC3.	
B114		chr13	21816875	A	G	transition	intergenic	NA	NA	4	Chromatin_Structure DNase-seq T47d, Chromatin_Structure FAIRE Nhbe, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Hmec, Protein_Binding EWS-ERG-fusion ChIP-seq CADO-ES1 ERG, Protein_Binding EWS_ERG_fusion ChIP_seq CADO_ES1 ERG	
B114		chr13	21816893	T	A	transversion	intergenic	NA	NA	4	Chromatin_Structure FAIRE Nhbe, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Hmec, Protein_Binding EWS-ERG-fusion ChIP-seq CADO-ES1 ERG, Protein_Binding EWS_ERG_fusion ChIP_seq CADO_ES1 ERG	
B114		chr13	21816900	G	A	transition	intergenic	CpG_site	NA	4	Chromatin_Structure FAIRE Nhbe, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Hmec, Protein_Binding EWS-ERG-fusion ChIP-seq CADO-ES1 ERG, Protein_Binding EWS_ERG_fusion ChIP_seq CADO_ES1 ERG	
B114		chr13	21816903	A	G	transition	intergenic	NA	NA	4	Chromatin_Structure FAIRE Nhbe, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Hmec, Protein_Binding EWS-ERG-fusion ChIP-seq CADO-ES1 ERG, Protein_Binding EWS_ERG_fusion ChIP_seq CADO_ES1 ERG	
B62	B63	chr20	30972541	G	C	transversion	ASXL1	intronic	NA	NA	4	Chromatin_Structure DNase-seq Hbvp, Chromatin_Structure DNase-seq Th1wb33676984, Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-seq Mel2183, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Colo829, Chromatin_Structure DNase-seq Be2c, Protein_Binding ChIP-seq H2afy2 LINE1, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure Dm002p1h DNase-seq Ecc1, Protein_Binding ChIP-seq HSPC IKZF1
B53	B54	chr13	32042443	G	A	transition	intergenic	NA	NA	4	Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure Dm002p1h DNase-seq Ecc1, Protein_Binding ChIP-seq HSPC IKZF1	
B113	B114	chr20	34539772	A	G	transition	SCAND1	3downstream	NA	NA	4	Chromatin_Structure DNase-seq Tregwb83319432, Protein_Binding ChIP-seq U87 POLR2A, Protein_Binding ChIP-seq HepG2 POLR2A, Protein_Binding ChIP-seq H1-hESC POI.R2A

B123	B124	chr19	35449194	T	C	transition	ZNF792	nonsyn p.N522S	NA	NA	4	Chromatin_Structure Znf4c50c4 DNase-seq K562, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq K562, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure DNase-seq Be2c, Protein_Binding ChIP-seq K562 SETDB1, Protein_Binding ChIP-seq U2OS SETDB1, Protein_Binding ChIP-seq K562 TRIM28, Protein_Binding ChIP-seq K562 RFX3, Protein_Binding ChIP-seq K562 CBX3, Chromatin_Structure DNase-seq HepG2, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Tregwb83319432, Chromatin_Structure Ctcfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifn4h DNase-seq Helas3, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Naivebcell, Chromatin_Structure DNase-seq Cd20ro01778, Protein_Binding ChIP-seq GM12878 ATF2
B93	B94	chr15	39934086	G	A	transition	FSIP1	intronic	NA	NA	4	Chromatin_Structure DNase-seq Hpd6e6e7, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Tregwb83319432, Chromatin_Structure Ctcfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifn4h DNase-seq Helas3, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Naivebcell, Chromatin_Structure DNase-seq Cd20ro01778, Protein_Binding ChIP-seq GM12878 ATF2
B62	B63	chr2	47527796	G	A	transition	AC073283.4	non-coding intronic	NA	NA	4	Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Naivebcell, Chromatin_Structure DNase-seq Cd20ro01778, Protein_Binding ChIP-seq GM12878 ATF2
B93	B94	chr17	49168444	T	C	transition	SPAG9	intronic	NA	NA	4	Chromatin_Structure DNase-seq Hmec, Chromatin_Structure FAIRE K562, Chromatin_Structure FAIRE HepG2, Protein_Binding ChIP-seq A549 POLR2A
B62	B63	chr20	52707477	A	G	transition		intergenic	NA	NA	4	Chromatin_Structure DNase-seq Gm12878, Chromatin_Structure DNase-seq Gm06990, Chromatin_Structure DNase-seq Gm12865, Protein_Binding ChIP-seq GM12878 FBF1
B93	B94	chr19	57037004	C	G	transversion	ZNF471	nonsyn p.A523G	NA	NA	4	Chromatin_Structure DNase-seq Hrgc, Protein_Binding ChIP-seq GM12878 ZNF274, Protein_Binding ChIP-seq U2OS SETDB1, Protein_Binding ChIP-seq NT2-D1 ZNF274, Protein_Binding ChIP-seq HEK293 TRIM28, Protein_Binding O2pct ChIP-seq A549 TCF12, Protein_Binding ChIP-seq K562 ZNF274, Protein_Binding ChIP-seq U2OS TRIM28, Protein_Binding ChIP-seq HepG2 ZNF274, Protein_Binding ChIP-seq GM12878 ZNF274

B123	B124	chr5	64235781	C	A	transversion	CWC27	intronic	NA	NA	4	Motifs Footprinting A549, Chromatin_Structure Diffa14d DNase-seq H7es, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure Znfb34a8 DNase-seq K562, Chromatin_Structure DNase-seq Hpdf, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Hrpe, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure DNase-seq Ag09319, Chromatin_Structure DNase-seq Prec, Chromatin_Structure Est100nm1h DNase-seq Mcf7, Chromatin_Structure Znf4c50c4 DNase-seq K562, Chromatin_Structure DNase-seq Hcpe, Chromatin_Structure DNase-seq Rptec, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Hmveclbl, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure DNase-seq Hvmf, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure Ctcfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Hae, Chromatin_Structure Estctrl0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Ag04450, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure DNase-seq Skmc, Chromatin_Structure Znfa41c6 DNase-seq K562, Chromatin_Structure Znff41b2 DNase-seq K562, Chromatin_Structure DNase-seq Saec, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Hbpv, Chromatin_Structure DNase-seq Nhbera, Chromatin_Structure DNase-seq Hipe, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq K562, Chromatin_Structure Diffa9d DNase-seq H7es, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure DNase-seq Hrce, Chromatin_Structure DNase-seq Hnpce, Chromatin_Structure Diffa2d DNase-seq H7es, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Hgf, Chromatin_Structure DNase-seq Nha, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure DNase-seq Hconf, Chromatin_Structure DNase-seq Hre, Chromatin_Structure DNase-seq Nhdfneo, Chromatin_Structure DNase-seq Hs5, Chromatin_Structure DNase-seq Th1wb33676984, Chromatin_Structure Hypoxlac FAIRE Mcf7, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Msc, Protein_Binding ChIP-seq HEK293 TRIM28
B93	B94	chr5	65445884	C	G	transversion	SREK1	intronic	NA	NA	4	

B83 B84	chr12	66449758	T	C	transition	intergenic	NA	NA	4	Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Gm18507, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure FAIRE Gm12891, Chromatin_Structure DNase-seq Gm18507, Protein_Binding tnfa ChIP-seq GM12878 NFKB1, Protein_Binding ChIP-seq GM12878 ATF2, Protein_Binding ChIP-seq GM12878 TBL1XR1, Protein_Binding tnfa ChIP-seq GM12892 NFKB1, Protein_Binding tnfa ChIP-seq GM12878 NFIC, Protein_Binding ChIP-seq GM12878 EP300, Protein_Binding ChIP-seq GM12878 NFATC1, Protein_Binding ChIP-seq GM12878 POLR2A, Protein_Binding ChIP-seq GM12878 BHLHE40, Protein_Binding ChIP-seq GM12891 POU2F2, Protein_Binding ChIP-seq VCAP AR, Protein_Binding ChIP- Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Hpd6e6e7, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure Estctr0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Medullo341, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Hek293t, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure DNase-seq H9es, Chromatin_Structure DNase-seq Medullo,	
LJ LS	chr16	71264288	C	T	transition	HYDIN	intronic	NA	NA	4	Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Hpd6e6e7, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure Estctr0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Medullo341, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Hek293t, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure DNase-seq H9es, Chromatin_Structure DNase-seq Medullo,
B54	chr12	76731688	G	T	transversion	intergenic	NA	NA	4	Chromatin_Structure DNase-seq Gm04503, Chromatin_Structure DNase-seq Bj, Chromatin_Structure DNase-seq Fibropag20443, Protein_Binding ChIP-seq HUVEC GATA2, Protein_Binding ChIP-seq HeLa-S3 JUN, Protein_Binding ChIP-seq HeLa-S3 EP300, Protein_Binding ChIP-seq IMR90 CEBPB, Protein_Binding ChIP-seq HeLa-S3 JUND, Protein_Binding ChIP-seq HUVEC FOS, Protein_Binding ChIP-	

B113	B114	chr16	86024734	T	C	transition	intergenic	NA	NA	4	Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Gm18507, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Gcbcell, Chromatin_Structure DNase-seq Cll, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Naivebcell, Chromatin_Structure DNase-seq Th1, Chromatin_Structure FAIRE Gm12891, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq Gm18507, Chromatin_Structure DNase-seq Gm12878, Protein_Binding ChIP-seq GM12878 PML, Protein_Binding ChIP-seq GM12878 ATF2, Protein_Binding ChIP-seq GM12878 TAF1, Protein_Binding ChIP-seq GM12878 POT1, Protein_Binding ChIP-seq GM12878 VV1
B123	B124	chr14	104390533	C	T	transition	intergenic	NA	NA	4	Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure Znf4c50c4 DNase-seq K562, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure Znff41b2 DNase-seq K562, Chromatin_Structure Znfa41c6 DNase-seq K562, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq K562, Chromatin_Structure Saha1u72hr DNase-seq K562, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq T47d, Chromatin_Structure Sahactrl DNase-seq K562, Chromatin_Structure Nabut DNase-seq K562, Chromatin_Structure DNase-seq Osteobl, Protein_Binding ChIP-seq SK-N-SH EP300, Protein_Binding ChIP-seq K562 ZBTB7A, Protein_Binding ChIP-seq SH-SY5Y GATA2, Protein_Binding ChIP-seq HUEC GATA2, Protein_Binding shbrg1 ChIP-seq CD36 GATA1, Protein_Binding ChIP-seq K562 TEAD4, Protein_Binding ChIP-seq Jurkat CREBBP, Protein_Binding ChIP-seq SK-N-SH VV1, Protein_Binding ChIP-seq Iuc ChIP-seq CD36 GATA1, Protein_Binding ChIP-seq Iuc ChIP-seq CD36 GATA1

LJ LS	chr3	120315393	G	A	transition	NDUFB4	intronic	NA	CpG_island	4	Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure Lenticon DNase-seq Fibroblgm03348, Chromatin_Structure Ifna4h FAIRE Helas3, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq Ipsnih7, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Hpaec, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq K562, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure DNase-seq Gbccll, Chromatin_Structure DNase-seq Cll, Chromatin_Structure DNase-seq Mel2183, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure Saha1u72hr DNase-seq K562, Chromatin_Structure DNase-seq Ipsnih11, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq Colo829, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Cerebellumoc, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Nhek, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Sknsn, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure DNase-seq Gliobla, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure Sahactrl DNase-seq K562, Chromatin_Structure Nabut DNase-seq K562, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure FAIRE Nhek, Chromatin_Structure DNase-seq Hek293t, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase- Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Gbccl, Chromatin_Structure FAIRE Gm12878, Chromatin_Structure DNase-seq Gm12864, Chromatin_Structure DNase-seq Gm12865, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase-seq Gm12878, Protein_Binding ChIP-seq GM12891 SPII, Protein_Binding ChIP-seq GM12878 RUNX3,
B113	chr4	122576011	T	C	transition		intergenic	NA	NA	4	

B103	B104	chr12	133397352	G	A	transition	GOLGA3	intronic	NA	NA	4	Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Gm18507, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq Gcbcell, Chromatin_Structure DNase-seq Gm12865, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq Gm18507, Chromatin_Structure DNase-seq Gm12870, Chromatin_Structure DNase-seq Gm12870, Chromatin_Structure DNase-seq Gm12870, Motifs Footprinting SkMC, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure Diffa14d DNase-seq H7es, Chromatin_Structure DNase-seq Ipsnih17, Chromatin_Structure DNase-seq Hpdlf, Chromatin_Structure DNase-seq Hmvecdneo, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Hrpe, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Ipsnih11, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure fng4h FAIRE Helas3, Chromatin_Structure DNase-seq Ag09319, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Hcpe, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Rptec, Chromatin_Structure DNase-seq M059j, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure DNase-seq Hcfaa, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Hvmf, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure Serumfree DNase-seq Aosmc, Chromatin_Structure DNase-seq Ag04450, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Hsmmm, Chromatin_Structure DNase-seq Hac, Chromatin_Structure DNase-seq Ag09309, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Skmc, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure Lenticon DNase-seq Fibroblgm03348, Chromatin_Structure fna4h FAIRE Helas3, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq Hipe, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq K562, Chromatin_Structure Diffa9d DNase-seq H7es, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure DNase-seq Hrc, Chromatin_Structure DNase-seq Hnpce, Chromatin_Structure DNase-seq Hl60, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Nhek, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq Hgf, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-
LJ LS		chr9	134281048	C	T	transition		intergenic	CpG_site	NA	4	

B93	B94	chr7	135414085	A	G	transition	SLC13A4	5upstream	NA	NA	4	Chromatin_Structure Znfb34a8 DNase-seq K562, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure Znfe103c6 DNase-seq K562, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Hs5, Chromatin_Structure Ohurea FAIRE K562, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure Znff41b2 DNase-seq K562, Chromatin_Structure Znfa41c6 DNase-seq K562, Chromatin_Structure Lenticon DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Hgf, Chromatin_Structure DNase-seq Hsmtt, Chromatin_Structure FAIRE K562, Chromatin_Structure DNase-seq Hsmmfshd, Chromatin_Structure Znfg54a11 DNase-seq K562, Chromatin_Structure Znfg47d3 DNase-seq K562, Protein_Binding ChIP-Motifs Footprinting HepG2, Chromatin_Structure DNase-seq Rptec, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure FAIRE Gm18507, Chromatin_Structure DNase-seq H7es, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure DNase-seq Hrce, Chromatin_Structure Diffa2d DNase-seq H7es, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Caco2, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Hre, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-seq Gm18507, Chromatin_Structure DNase-seq Gm12878, Chromatin_Structure FAIRE Hepg2, Chromatin_Structure FAIRE A549, Protein_Binding ChIP-seq HepG2 EP300, Protein_Binding ChIP-seq HepG2 RAD21, Protein_Binding ChIP-seq HepG2 NFIC, Protein_Binding ChIP-seq HepG2 SP1, Protein_Binding 02pct ChIP-seq A549 EP300, Protein_Binding differential ChIP-Chromatin_Structure DNase-seq Sae, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Wi38, Chromatin_Structure DNase-seq Hipe, Chromatin_Structure DNase-seq Nhek, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hbmec, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Th17, Chromatin_Structure Ohtam DNase-seq Wi38, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Msc, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Nhdfad, Chromatin_Structure Hypoxlac FAIRE Mcf7, Chromatin_Structure DNase-seq Hnpce, Chromatin_Structure DNase-seq Gm04504, Chromatin_Structure DNase-seq Nha, Chromatin_Structure DNase-seq Nhlf, Protein_Binding 02pct ChIP-seq A549 FOSL2, Protein_Binding ChIP-seq GM19099 POLR2A, Protein_Binding ChIP-seq K562 ARID3A.
B83	B84	chr7	138001305	T	C	transition		intergenic	NA	NA	4	
B83	B84	chr1	150338972	A	G	transition	RPRD2	intronic	NA	NA	4	

B53 B54	chr4	155058899	C	A	transversion	intergenic	NA	NA	4	Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure DNase-seq Hrce, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Ipsi, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Phte, Chromatin_Structure DNase-seq H9es, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Psoasmuscleoc, Chromatin_Structure DNase-
B103 B104	chr5	172158355	G	A	transition	intergenic	CpG_site	NA	4	Chromatin_Structure DNase-seq Hpde6e6e7, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure Hypoxlaccon DNase-seq Mcf7, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure Ctcfshrna DNase-seq Mcf7, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure Ifna4h FAIRE Helas3, Chromatin_Structure DNase-seq K562, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Hu7, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure DNase-seq Hek293t, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Rwpe1, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure DNase-seq Phte, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure Hypoxlac FAIRE Mcf7, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq K562G1phase, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure Dm002p1h DNase-seq Ecc1, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure Randshrna DNase-seq Mcf7, Protein_Binding ChIP-seq HeLa-S3 CHD2, Protein_Binding ChIP-seq HepG2 MAX, Protein_Binding ChIP-seq K562 JUND, Protein_Binding ChIP-seq HepG2 SIN3A, Protein_Binding ChIP-seq HeLa-S3 SMARCC1, Protein_Binding ChIP-seq HeLa-S3 MYC, Protein_Binding differential ChIP-seq Caco2 CDX2, Protein_Binding ChIP-seq HeLa-S3 CEBPB, Protein_Binding ChIP-seq K562 MAX, Protein_Binding ChIP-seq K562 RCP1

B83	B84	chr2	209224680	C	T	transition	PIKFYVE	3downstream	NA	NA	4	Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Olfneurosphere, Chromatin_Structure DNase-seq H9es, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Ipsnih11, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Myometr, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Protein_Binding ChIP-seq GM12878 POLR2A,
B103	B104	chr8	674731	A	G	transition	ERICH1	intronic	NA	NA	5	Chromatin_Structure DNase-seq Ipsnih17, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq H9es, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Medullo341, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure Ifna4h FAIRE Helas3, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Medullo,
B123	B124	chr8	1779112	G	A	transition	ARHGEF10	intronic	CpG_site	NA	5	Chromatin_Structure DNase-seq Hpd1f, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hgf, Chromatin_Structure DNase-seq Nhdfneo, Chromatin_Structure DNase-seq Nhdfad Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Nb4, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq Gm12892,
B62	B63	chr4	1883359	A	G	transition	WHSC1	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hpd1f, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hgf, Chromatin_Structure DNase-seq Nhdfneo, Chromatin_Structure DNase-seq Nhdfad Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Nb4, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq Gm12892,
B123	B124	chr4	2463556	C	T	transition	RP11_503N18.	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hpd1f, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hgf, Chromatin_Structure DNase-seq Nhdfneo, Chromatin_Structure DNase-seq Nhdfad Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Nb4, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq Gm12892,

B93 B94	chr5	3908422	C	T	transition	intergenic	NA	NA	5	Motifs PWM SPIB, Motifs PWM PU.1, Motifs PWM Elf5, Motifs PWM SPIC, Motifs PWM SOX, Motifs PWM Spf1, Motifs PWM PU1, Motifs PWM SII, Motifs PWM Spic, Motifs PWM ELF5, Motifs PWM SPI-B, Chromatin_Structure DNase-seq T47d	
B93 B94 LJ LS	chr2 chr1	6263593 7523207	C G	T A	transition	intergenic CAMTA1	intronic	NA CpG_site	NA	5 5	Chromatin_Structure DNase-seq Werirb1 Motifs PWM Dax1, Motifs PWM RARG, Motifs PWM Zic3, Motifs Footprinting Gm12892 PPARG::RXRA, Motifs PWM PPARG::RXRA, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Werirb1 Motifs Footprinting H1hesc SREBP-1, Motifs PWM VDR, Motifs Footprinting Hsmmt SREBP-1, Motifs Footprinting Gm12892 SREBP, Motifs Footprinting 8988t SREBP, Motifs Footprinting Lncap SREBP-1, Motifs Footprinting Huh7 SREBP-1, Motifs Footprinting Gm12878 SREBP-1, Motifs Footprinting Gm12891 SREBP, Motifs Footprinting Huh7 SREBP, Motifs Footprinting Gm12891 SREBP-1, Motifs Footprinting Gm12878 SREBP, Motifs PWM SREBP, Motifs Footprinting Hpde6e6e7 SREBP, Motifs Footprinting LncapAndro SREBP, Motifs Footprinting Chorion SREBP, Motifs Footprinting LncapAndro SREBP-1, Motifs Footprinting Osteobl SREBP, Motifs Footprinting Gm12892 SREBP-1, Motifs PWM SREBP1, Motifs Footprinting Myometr SREBP, Motifs Footprinting Hpde6e6e7 SREBP-1, Motifs Footprinting Hepatocytes SREBP, Motifs Footprinting H1hesc SREBP, Motifs Footprinting A549 SREBP-1, Motifs Footprinting Hsmmt SREBP, Motifs Footprinting Cll SREBP, Motifs PWM SREBP-1, Motifs Footprinting Osteobl SREBP-1, Motifs Footprinting Lncap SREBP, Motifs Footprinting Chorion SREBP-1, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Pit-1, Motifs Footprinting H1hesc Pit-1, Motifs Footprinting Gm12891 Pit-1, Motifs Footprinting Huh7 Pit-1, Motifs Footprinting LncapAndro Pit-1, Motifs Footprinting Chorion Pit-1, Chromatin_Structure DNase-seq Medullo341, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Medullo,
LJ LS	chr19	8468273	C	T	transition	RAB11B	intronic	CpG_site	NA	5	Motifs Footprinting H1hesc SREBP-1, Motifs PWM VDR, Motifs Footprinting Hsmmt SREBP-1, Motifs Footprinting Gm12892 SREBP, Motifs Footprinting 8988t SREBP, Motifs Footprinting Lncap SREBP-1, Motifs Footprinting Huh7 SREBP-1, Motifs Footprinting Gm12878 SREBP-1, Motifs Footprinting Gm12891 SREBP, Motifs Footprinting Huh7 SREBP, Motifs Footprinting Gm12891 SREBP-1, Motifs Footprinting Gm12878 SREBP, Motifs PWM SREBP, Motifs Footprinting Hpde6e6e7 SREBP, Motifs Footprinting LncapAndro SREBP, Motifs Footprinting Chorion SREBP, Motifs Footprinting LncapAndro SREBP-1, Motifs Footprinting Osteobl SREBP, Motifs Footprinting Gm12892 SREBP-1, Motifs PWM SREBP1, Motifs Footprinting Myometr SREBP, Motifs Footprinting Hpde6e6e7 SREBP-1, Motifs Footprinting Hepatocytes SREBP, Motifs Footprinting H1hesc SREBP, Motifs Footprinting A549 SREBP-1, Motifs Footprinting Hsmmt SREBP, Motifs Footprinting Cll SREBP, Motifs PWM SREBP-1, Motifs Footprinting Osteobl SREBP-1, Motifs Footprinting Lncap SREBP, Motifs Footprinting Chorion SREBP-1, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Pit-1, Motifs Footprinting H1hesc Pit-1, Motifs Footprinting Gm12891 Pit-1, Motifs Footprinting Huh7 Pit-1, Motifs Footprinting LncapAndro Pit-1, Motifs Footprinting Chorion Pit-1, Chromatin_Structure DNase-seq Medullo341, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Medullo,
B93 B94	chr18	12057323	A	C	transversion	intergenic	NA	NA	5	Motifs Footprinting H1hesc Pit-1, Motifs Footprinting Gm12891 Pit-1, Motifs Footprinting LncapAndro Pit-1, Motifs Footprinting Chorion Pit-1, Chromatin_Structure DNase-seq Medullo341, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Medullo,	
B124	chr17	12638660	T	C	transition	MYOCD	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hcf, Chromatin_Structure DNase-seq Osteobl
B124	chr17	12638661	A	G	transition	MYOCD	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hcf, Chromatin_Structure DNase-seq Osteobl

B123	B124	chr3	13085170	C	T	transition	IQSEC1	intronic	CpG_site	NA	5	Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure Nabut DNase-seq K562, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Psoasmuscleoc,
B53	B54	chr9	16205585	C	T	transition	C9orf92	intronic	NA	NA	5	Motifs PWM EWSR1-FLI1, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Hpaf, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq Hcm, Chromatin_Structure DNase-seq Nhdfad, Chromatin_Structure DNase-seq Aacf
B53	B54	chr5	16385105	T	C	transition	RP1_167G20.1	non-coding intronic	NA	NA	5	Chromatin_Structure Znfa41c6 DNase-seq K562, Chromatin_Structure DNase-seq K562
B113	B114	chr11	16923047	T	A	transversion	PLEKHA7	intronic	NA	NA	5	Motifs PWM Nanog, Chromatin_Structure DNase-seq Melano, Chromatin_Structure FAIRE Gm12892
B103	B104	chr9	17578171	A	C	transversion	SH3GL2	5upstream	NA	NA	5	Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase-seq Helas3
B103	B104	chr16	19266444	C	T	transition	SYT17	intronic	NA	NA	5	Chromatin_Structure DNase-seq Ips
B53	B54	chr5	19663794	A	G	transition	CDH18	intronic	NA	NA	5	Motifs PWM MEF-2, Protein_Binding ChIP-seq HSPC SPI1
B103	B104	chr20	19700562	G	A	transition	SLC24A3	intronic	NA	NA	5	Chromatin_Structure DNase-seq Cll
B103	B104	chr17	20145251	C	T	transition	SPECC1	intronic	NA	NA	5	Chromatin_Structure Ctcfshrna DNase-seq Mcf7, Chromatin_Structure DNase-seq Mcf7
B53	B54	chr7	20169410	C	T	transition	AC005062.2	non-coding intronic	CpG_site	NA	5	Chromatin_Structure DNase-seq H7es, Chromatin_Structure Diffa5d DNase-seq H7es
B93	B94	chr12	20304160	A	G	transition	intergenic		NA	NA	5	Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure FAIRE Gm12891, Chromatin_Structure Dm002p1h DNase-seq Ecc1
B103	B104	chr7	21283412	G	A	transition	intergenic		CpG_site	NA	5	Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Th1
B83	B84	chr12	21661853	A	C	transversion	GOLT1B	intronic	NA	NA	5	Chromatin_Structure FAIRE Gm18507, Chromatin_Structure DNase-seq Hl60
B93	B94	chr6	21747135	C	T	transition	CASC15	non-coding intronic	NA	NA	5	Chromatin_Structure DNase-seq H7es

B62 B63	chr22	22003746	T	A	transversion	intergenic	NA	NA	5	Motifs Footprinting Helas3 HNF3beta, Motifs PWM HFH8(FOXF1A), Motifs Footprinting H1hesc FOXD3, Motifs Footprinting Gm12892 HNF3beta, Motifs Footprinting Gm12892 HFH4(FOXJ1), Motifs Footprinting Gm12892 HFH3(FOXII), Motifs PWM HNF3beta, Motifs PWM HNF3, Motifs PWM HFH4(FOXJ1), Motifs Footprinting Gm12878 HNF3beta, Motifs PWM Foxj3, Motifs Footprinting Helas3 HFH8(FOXF1A), Motifs Footprinting Chorion HNF3beta, Motifs Footprinting Chorion Foxd3, Motifs PWM FOXC1, Motifs Footprinting Helas3 FOXfactors, Motifs Footprinting Helas3 FOXO3A, Motifs Footprinting Chorion Freac-7, Motifs Footprinting Gm12878 FOXII1, Motifs Footprinting K562 FOXO3A, Motifs Footprinting Gm12892 Freac-7, Motifs Footprinting Gm12878 HFH4(FOXJ1), Motifs Footprinting Gm12891 Cdx, Motifs Footprinting Gm12892 FOXO3A, Motifs Footprinting Helas3 FOXII1, Motifs PWM Cdx, Motifs Footprinting Gm12878 FOXO3A, Motifs Footprinting K562 HFH4(FOXJ1), Motifs PWM ONECUT3, Motifs PWM Tlx2, Motifs Footprinting K562 HNF3, Motifs Footprinting Gm12892 HFH8(FOXF1A), Motifs Footprinting Gm12878 FOXfactors, Motifs Footprinting Gm12878 Foxd3, Motifs Footprinting Huh7 HFH3(FOXII), Motifs Footprinting Gm12892 Cdx, Motifs Footprinting H1hesc HNF3, Motifs Footprinting H1hesc HFH3(FOXII), Motifs Footprinting Gm12878 HFH4(FOXJ1), Motifs Footprinting Gm12891 Cdx, Motifs Footprinting Gm12892 FOXO3A, Motifs Footprinting Helas3 FOXII1, Motifs Footprinting Helas3 HFH4(FOXJ1), Motifs Footprinting Gm12878 FOXfactors, Motifs Footprinting Foxl1, Motifs Footprinting K562 Cdx, Motifs Footprinting Chorion HFH4(FOXJ1), Motifs Footprinting H1hesc Freac-7, Motifs Footprinting Helas3 Cdx, Motifs Footprinting FOXP1, Motifs Footprinting Chorion HFH3(FOXII), Motifs Footprinting H1hesc Foxd3, Motifs Footprinting Helas3 HFH3(FOXII), Motifs Footprinting Gm12878 Freac-7, Motifs Footprinting Gm12878 Cdx, Motifs Footprinting FOXJ2, Motifs Footprinting Gm12878 HFH8(FOXF1A), Motifs Footprinting Chorion Cdx, Motifs Footprinting K562 HFH3(FOXII), Motifs Footprinting Gm12892 FOXII1, Motifs Footprinting Hoxd8, Motifs Footprinting Chorion HFH8(FOXF1A), Motifs Footprinting Chorion FOXO3A, Motifs Footprinting Gm12878 HNF3, Motifs Footprinting Chorion FOXfactors, Motifs Footprinting Helas3 Freac-7, Motifs Footprinting Chorion HNF3, Motifs Footprinting Gm12892 FOXfactors, Motifs Footprinting H1hesc HFH4(FOXJ1), Motifs Footprinting Gm12892 FOXL1, Motifs Footprinting Gm12878 FOXO3A, Motifs Footprinting Gm12878 FOXII, Motifs Footprinting Gm12878 Foxd3,
---------	-------	----------	---	---	--------------	------------	----	----	---	--

B62 B63	chr1	23120852	C	T	transition	EPHB2	intronic	CpG_site	NA	5	Motifs Footprinting Gm12892 HSF, Motifs Footprinting Gm12892 HSF1, Motifs PWM HSF1, Motifs Footprinting Gm12891 HSF1, Motifs Footprinting Hsmmm HSF1, Motifs Footprinting Hsmmt HSF1, Motifs PWM HSF, Motifs Footprinting Osteobl HSF1, Motifs Footprinting Hsmmt HSF, Motifs Footprinting Gm12891 HSF, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Hsmmm, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure Nabut DNase-seq K562, Chromatin_Structure DNase-seq K562G2mphase, Chromatin_Structure DNase-
LJ LS B103 B104	chr10 chr8	24822488 25459305	T G	C T	transition transversion	KIAA1217 intergenic	intronic	NA NA	NA NA	5 5	Motifs PWM Pitx1, Chromatin_Structure DNase-seq Hbvp Motifs PWM FOXG1, Chromatin_Structure DNase-seq Hmveclbl, Chromatin_Structure DNase-seq Hmvecdblad, Chromatin_Structure DNase-seq Me12183, Chromatin_Structure DNase-seq Colo829
B113 B114 LJ LS	chr17 chr4	26144055 27144101	A G	G A	transition transition	intergenic intergenic		NA CpG_site	NA NA	5 5	Chromatin_Structure DNase-seq Werirb1 Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq H7es, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure DNase-seq H9es
B113 B114	chr13	28557037	C	T	transition	URAD	intronic	NA	NA	5	Motifs Footprinting Hepatocytes STAT5A(homotetramer), Motifs PWM STAT5A(homotetramer), Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Mycmet
B83 B84	chr4	29361176	T	A	transversion	intergenic		NA	NA	5	Motifs PWM Cdx, Motifs PWM Sox4, Motifs PWM FOXP1, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Monocd14
B93 B94 B62 B63 B83 B84 LJ LS B93 B94 B93 B94 LJ LS LJ LS	chr3 chr11 chr14 chr3 chr11 chr11 chr7 chr6	30351562 30391709 35915225 36418949 36428450 36428499 36863766 37921491	T A C C G C C G	C C T T C A T A	transition transversion transition transition transversion transversion transition transition	intergenic intergenic intergenic intergenic PRR5L PRR5L intergenic ZFAND3	intronic intronic intronic intronic intronic intronic intronic intronic	NA NA NA NA NA NA NA NA	NA NA NA NA NA NA NA NA	5 5 5 5 5 5 5 5	Chromatin_Structure DNase-seq H7es Motifs PWM NR2E1, Motifs PWM FOXC2, Protein_Binding ChIP-seq VCaP AR Chromatin_Structure DNase-seq Jurkat Chromatin_Structure DNase-seq Rptec Chromatin_Structure DNase-seq Hgf Chromatin_Structure DNase-seq Hgf Chromatin_Structure DNase-seq Werirb1 Chromatin_Structure DNase-seq M059j, Chromatin_Structure DNase-seq Hae, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Hipe, Chromatin_Structure DNase-seq Werirb1, Chromatin_Structure DNase-seq Nhdfad, Chromatin_Structure DNase-seq Rc?c Motifs PWM HNF4A, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure DNase-seq Medulloblastoma341, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure Hypoxlac DNase-seq Mcf7, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Mcf7, Chromatin_Structure DNase-seq 12002
B53 B54	chr17	39595049	G	T	transversion	KRT38	nonsyn p.P265H	NA	NA	5	Chromatin_Structure DNase-seq Nhek Motifs PWM Srf, Protein_Binding ChIP-seq K562 EP300, Protein_Binding ChIP-seq K562 TAL1 Motifs PWM NF-Y, Motifs PWM alpha-CP1, Chromatin_Structure DNase-seq M059j, Chromatin_Structure DNase-seq H7es, Chromatin_Structure Diffa5d DNase-seq H7es
B83 B84 B123 B124	chr6 chr21	40413089 40959364	G T	A C	transition transition	LRFN2 intergenic	intronic	CpG_site NA	NA NA	5 5	
B93 B94	chr8	41046737	A	T	transversion	intergenic		NA	NA	5	

B53 B54	chr13	42917019	C	G	transversion	intergenic	NA	NA	5	Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Monocd14	
B53 B54	chr22	43077904	T	C	transition	intergenic	NA	NA	5	Protein_Binding proliferation ChIP-seq Caco2 HNF4A	
B83 B84	chr14	43204114	A	G	transition	intergenic	NA	NA	5	Motifs PWM Freac-7, Motifs PWM FOXJ3, Motifs PWM TEF, Motifs PWM FOXJ2, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seal Hff	
B93 B94	chr1	43646521	C	T	transition	WDR65	intronic	NA	NA	5	Motifs PWM VDR, Chromatin_Structure DNase-seq Th1, Chromatin_Structure DNase-seq Th17
B62 B63	chr22	45313501	A	G	transition	PHF21B	intronic	NA	NA	5	Motifs PWM GATA-6, Motifs Footprinting Cl GATA-6, Motifs Footprinting H1hesc GATA-6, Motifs PWM Gata1, Motifs Footprinting Fibrop GATA-6, Motifs Footprinting Hsmmm GATA-6, Motifs Footprinting Hsmmm Gata1, Motifs Footprinting Gm12892 Gata1, Motifs Footprinting Chorion Gata1, Motifs Footprinting GM12892 Nrf-2, Motifs Footprinting Hsmmt GATA-6, Motifs Footprinting 8988t GATA-6, Motifs Footprinting Lymphoblastoid Nrf-2, Motifs Footprinting LncapAndro Gata1, Motifs Footprinting Lncap GATA-6, Motifs Footprinting Lymphoblastoid Eip74EF, Motifs Footprinting GM12892 Eip74EF, Motifs Footprinting Gm12892 GATA-6, Motifs Footprinting Lymphoblastoid dl, Motifs Footprinting Lymphoblastoid GABPA, Motifs Footprinting Lymphoblastoid ELK4, Motifs Footprinting Chorion GATA-6, Motifs Footprinting Lymphoblastoid dl_1, Motifs Footprinting Hepatocytes GATA-6, Motifs Footprinting LncapAndro GATA-6, Motifs Footprinting H1hesc Gata1, Motifs Footprinting GM12892 ELK4, Motifs Footprinting GM12892 GABPA, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm12892, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Hsmmm, Chromatin_Structure DNase-seq Psoasmucleoc, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase- seq Frontal_cortex, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Cerebellumoc
B123 B124	chr11	47691714	A	G	transition	AGBL2	intronic	NA	NA	5	Motifs PWM AP-2gamma, Motifs PWM TCFAP2A, Motifs PWM AP-2alpha, Motifs PWM HIC1, Motifs PWM Zic3, Motifs PWM TFAP2C, Motifs PWM TFAP2A, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Phte, Chromatin_Structure DNase-seq Frontal_cortex, Chromatin_Structure DNase-seq Cerebrumfrontaloc
B113 B114	chr22	49740309	G	C	transversion	intergenic	CpG_site	NA	5	Motifs PWM AP-2gamma, Motifs PWM TCFAP2A, Motifs PWM AP-2alpha, Motifs PWM HIC1, Motifs PWM Zic3, Motifs PWM TFAP2C, Motifs PWM TFAP2A, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Phte, Chromatin_Structure DNase-seq Frontal_cortex, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Cerebellumoc	
LJ LS B93 B94	chr7 chr3	50434049 50574244	G T	A C	transition transition	IKZF1 intergenic	intronic	NA NA	NA NA	5 5	Chromatin_Structure DNase-seq Cd4naivewb11970640, Chromatin_Structure DNase-seq Hbvp, Chromatin_Structure FAIRE UrotsaUt189, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq 8988t
LJ LS B123 B124	chr20 chr3	50688780 51651550	G C	T	transversion transition	intergenic RAD54L2	intronic	NA NA	NA NA	5 5	Protein_Binding ChIP-seq HepG2 MAFK, Motifs PWM Hoxb13, Motifs PWM Hoxd10, Motifs PWM Hoxd13, Chromatin_Structure DNase-seq Monocd14ro1746, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Monocd14
B103 B104	chr4	53162451	C	T	transition	intergenic		NA	NA	5	Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure Est10nm30m DNase-seq Ishikawa

B53	B54	chr15	77314816	G	A	transition	PSTPIP1	intronic	NA	NA	5	Chromatin_Structure DNase-seq Progfib, Chromatin_Structure DNase-seq Htr8, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Huvec, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure DNase-seq UrotsaUt189, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Gm19240, Chromatin_Structure DNase-seq Melano, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure DNase-seq Gm10248, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Fibropag08396, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure Lenticon DNase-seq Fibroblgm03348, Chromatin_Structure Ifna4h FAIRE Helas3, Chromatin_Structure DNase-seq Adultcd4th0, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure DNase-seq Werirb1, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq A549, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure DNase-seq Urotsa, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Phte, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure Lentimyod DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Hsmmfshd, Chromatin_Structure DNase-seq Hsmmemb, Chromatin_Structure DNase-seq Fibropag20443, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq Chorion, Chromatin_Structure DNase-
B123	B124	chr9	77915529	G	A	transition		intergenic	CpG_site	NA	5	Motifs PWM FAC1, Motifs PWM CLOCK, Chromatin_Structure DNase-seq M059j, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Th2, Chromatin_Structure DNase-seq Th2wh54553?04, Chromatin_Structure DNase-seq Th1
B123	B124	chr5	79459636	T	C	transition	SERINC5	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hnpce, Chromatin_Structure DNase-seq Lhcnm2

B62 B63	chr16	81489957	A	C	transversion	CMIP	intronic	NA	NA	5	Motifs Footprinting Gm12891 ARP-1(COUP-TF2), Motifs Footprinting Hsmm ARP-1(COUP-TF2), Motifs Footprinting Hmec ARP-1(COUP-TF2), Motifs Footprinting Gm12892 ARP-1(COUP-TF2), Motifs PWM RAR _A , Motifs Footprinting Medullo ARP-1(COUP-TF2), Motifs Footprinting Gm19238 ARP-1(COUP-TF2), Motifs PWM THRA, Motifs Footprinting 8988t ARP-1(COUP-TF2), Motifs Footprinting Gm12892 GCNF, Motifs Footprinting Lncap ARP-1(COUP-TF2), Motifs PWM ARP-1(COUP-TF2), Motifs Footprinting Chorion ARP-1(COUP-TF2), Motifs Footprinting Gm19239 ARP-1(COUP-TF2), Motifs Footprinting Osteobl ARP-1(COUP-TF2), Motifs Footprinting A549 GCNF, Motifs Footprinting H1hesc ARP-1(COUP-TF2), Motifs Footprinting Huh75 ARP-1(COUP-TF2), Motifs PWM TTF-1(Nkx2-1), Motifs Footprinting 8988t GCNF, Motifs Footprinting 8988t TTF-1(Nkx2-1), Motifs Footprinting Gm12892 TTF-1(Nkx2-1), Motifs PWM GCNF, Motifs Footprinting Hepg2 ARP-1(COUP-TF2), Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq H7es, Chromatin_Structure Znfa41c6 DNase-seq K562, Chromatin_Structure Znff41b2 DNase-seq K562, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq K562, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-
B83 B84	chr9	82567879	T	C	transition		intergenic	NA	NA	5	Chromatin_Structure DNase-seq Panisd, Chromatin_Structure FAIRE Gliobla, Chromatin_Structure DNase-seq Colo829
B93 B94	chr11	83191270	C	A	transversion	DLG2	intronic	NA	NA	5	Motifs PWM Gf11, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure DNase-seq Hah
LJ LS	chr11	84078617	G	T	transversion	DLG2	intronic	NA	NA	5	Motifs PWM ATF4, Motifs PWM Duxl, Motifs PWM Cphx, Motifs PWM DUXA, Protein_Binding ChIP-seq VCaP ERG
B103 B104	chr14	85591083	T	C	transition		intergenic	NA	NA	5	Motifs PWM LHX8, Motifs PWM LHX6, Chromatin_Structure DNase-seq H7es, Chromatin_Structure Diffa5d DNase-seq H7es
B62 B63	chr15	90929885	T	C	transition	IQGAP1	5upstream	NA	NA	5	Motifs PWM Nkx3-1, Motifs Footprinting Hepatocytes Nkx2-9, Motifs Footprinting A549 Nkx2-9, Motifs PWM Nkx2-9, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Osteobl
B93 B94	chr10	94297199	C	A	transversion	IDE	syn	NA	NA	5	Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Hae, Chromatin_Structure Hypoxlac FAIRE Mcf7

B93	B94	chr7	97860074	G	A	transition	TECPR1	intronic	NA	NA	5	Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Hsmm, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure DNase-seq Panislets, Chromatin_Structure DNase-seq Gm13976, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Adultcd4th1, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure Nabut DNase-seq K562, Chromatin_Structure DNase-seq Hsmmt, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure DNase-seq Osteobl, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-seq Hsmmfshd, Chromatin_Structure DNase-seq Medullo,
B113	B114	chr3	98242563	T	C	transition	CLDND1	Supstream	NA	NA	5	Chromatin_Structure DNase-seq Th1wb33676984, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm12865, Chromatin_Structure Znfp5 DNase-seq K562, Chromatin_Structure FAIRE Gm12892, Chromatin_Structure DNase-seq Th1
LJ	LS	chr11	100687399	G	A	transition	ARHGAP42	intronic	NA	NA	5	Motifs PWM Nkx6-1, Motifs PWM GSX2, Motifs PWM RAX, Motifs PWM HMX2, Motifs PWM HMX1, Motifs PWM HMX3, Motifs PWM RFX. Chromatin_Structure DNase-seq Hmec
B103	B104	chr13	101552351	C	A	transversion	NALCN_AS1	non-coding intronic	NA	NA	5	Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Th2, Chromatin_Structure DNase-seq Th2wb54553204, Chromatin_Structure DNase-seq Gm12865 Chromatin_Structure DNase-seq Th1
B103	B104	chr14	104320775	G	A	transition	LINC00637	non-coding intronic	CpG_site	NA	5	Chromatin_Structure DNase-seq Chorion
B103	B104	chr12	104900233	A	G	transition	CHST11	intronic	NA	NA	5	Protein_Binding ChIP-seq Jurkat CREBBP
LJ	LS	chr12	105193487	A	G	transition		intergenic			5	Chromatin_Structure Ohureal FAIRE K562, Chromatin_Structure DNase-seq H9es
B93	B94	chr3	113657145	G	A	transition	GRAMD1C	intronic	NA	NA	5	Protein_Binding ChIP-seq HepG2 MAFK
B83	B84	chr5	116550935	G	A	transition		intergenic	CpG_site	NA	5	Chromatin_Structure DNase-seq Th17, Chromatin_Structure DNase-seq Th1
B113	B114	chr10	117458502	T	A	transversion	ATRNL1	intronic	NA	NA	5	Motifs PWM Srf, Protein_Binding ChIP-seq HepG2 MAFK
B103	B104	chr3	118923152	T	C	transition	UPK1B	3utr	NA	NA	5	Chromatin_Structure DNase-seq K562
B103	B104	chr8	121565361	A	G	transition	SNTB1	intronic	NA	NA	5	Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Hcpe, Chromatin_Structure FAIRE Htr8, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Hbvp, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure FAIRE Urotsa, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Fibrop
B113	B114	chr10	123074966	T	C	transition		intergenic	NA	NA	5	

B113	B114	chr9	124373251	T	C	transition	DAB2IP	intronic	NA	NA	5	Motifs PWM HFF8(FOXF1A), Motifs PWM HNF3, Motifs PWM HFF4(FOXJ1), Motifs Footprinting Hmec FOXJ2, Motifs PWM FOXC1, Motifs PWM FOXP1, Motifs PWM FOXL1, Motifs PWM FOXD3, Motifs PWM FOXI1, Motifs PWM Foxd3, Motifs Footprinting LncapAndro FOXJ2, Motifs Footprinting LncapAndro HNF3, Motifs Footprinting H9es FOXJ2, Motifs Footprinting Medullo HFF4(FOXJ1), Motifs PWM FOXC2, Motifs Footprinting LncapAndro HFF8(FOXF1A), Motifs PWM FOXJ2, Motifs Footprinting Medullo FOXJ2, Motifs Footprinting LncapAndro HFF4(FOXJ1),
LJ LS	chr6	124510527	C	T	transition	NKAIN2	intronic	NA	NA	5	Chromatin_Structure DNase-seq Lhcnm2	
LJ LS	chr6	124515065	A	T	transversion	NKAIN2	intronic	NA	NA	5	Motifs PWM SREBP, Motifs PWM Os2, Motifs PWM SREBP1, Chromatin_Structure DNase-seq H7es	
B103	B104	chr11	126855973	C	A	transversion	KIRREL3	intronic	NA	NA	5	Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq T47d
B113	B114	chr5	127441266	A	G	transition	SLC12A2	intronic	NA	NA	5	Chromatin_Structure FAIRE Gm19239, Chromatin_Structure DNase-seq Nhlf, Chromatin_Structure DNase-seq Aoaf
B62	B63	chr7	127503274	G	A	transition	SND1	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hcpe, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure DNase-seq Hbvp, Chromatin_Structure DNase-seq Hbvsme
B123	B124	chr4	128289126	C	T	transition	intergenic		CpG_site	NA	5	Motifs PWM HOXA10, Motifs PWM Hoxb9, Motifs PWM HOXC11, Motifs PWM Hoxa9, Motifs PWM Hoxa10, Motifs PWM hoxa9, Motifs PWM HOXA11, Chromatin_Structure DNase-seq Osteoblast
B123	B124	chr6	138054222	T	C	transition	intergenic		NA	NA	5	Motifs PWM Sox4, Protein_Binding ChIP-seq HepG2 POLR2A
B93	B94	chr6	139472949	A	G	transition	HECA	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Hasp, Chromatin_Structure FAIRE Gm19239
B83	B84	chr9	139573616	C	T	transition	AGPAT2	intronic	NA	NA	5	Motifs Footprinting Chorion TFII-I, Motifs Footprinting H1hesc TFII-I, Motifs Footprinting LncapAndro TFII-I, Motifs Footprinting Lncap TFII-I, Motifs Footprinting Gm12892 TFII-I, Motifs PWM TFII-I, Chromatin_Structure DNase-seq Cd20ro01794, Chromatin_Structure DNase-seq Heartoc, Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq Sknsh, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Cerebrumfrontaloc, Chromatin_Structure DNase-seq Gm12891, Chromatin_Structure DNase-seq 8988t, Chromatin_Structure DNase-seq Gm20000, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Monocd14, Chromatin_Structure Ifna4h FAIRE Helas3, Chromatin_Structure Andro DNase-seq Lncap, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Hepatocytes, Chromatin_Structure DNase-seq Gm10266, Chromatin_Structure DNase-seq Frontalcortexoc, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq Huh7, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure DNase-seq Gm19238, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure DNase-seq Gm19239, Chromatin_Structure DNase-seq Huh75, Chromatin_Structure DNase-seq Hmec, Chromatin_Structure DNase-seq Osteoblast, Chromatin_Structure DNase-seq Gm13977, Chromatin_Structure DNase-

B123	B124	chr9	139787912	G	T	transversion	TRAF2	intronic	NA	NA	5	Chromatin_Structure Ifng4h FAIRE Helas3, Chromatin_Structure DNase-seq Lncap, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure DNase-seq Th1
B123	B124	chr6	150229677	G	T	transversion	RAET1E_AS1	non-coding intronic	NA	NA	5	Protein_Binding ChIP-seq K562 SETDB1, Protein_Binding ChIP-seq K562 TRIM28, Protein_Binding ChIP-seq HEK293 TRIM28, Protein_Binding ChIP-seq K562 CBX3
B93	B94	chr1	151829054	G	C	transversion		intergenic	NA	NA	5	Chromatin_Structure Est10nm30m DNase-seq T47d, Chromatin_Structure DNase-seq Medulloid341, Chromatin_Structure DNase-seq Medullo, Chromatin_Structure DNase-seq T47d, Chromatin_Structure DNase-seq Werirh1
B114		chr6	154215825	T	C	transition		intergenic	NA	NA	5	Motifs Footprinting CII MEF2A, Motifs Footprinting CII MEF-2, Motifs PWM MEF2A, Motifs PWM MEF-2, Motifs Footprinting Helas3 Ifna4h MEF-2, Motifs Footprinting Helas3 Ifna4h MEF2A, Chromatin_Structure FAIRE Panislets, Chromatin_Structure Znf4c50c4 DNase-seq K562, Chromatin_Structure FAIRE Gm12878, Chromatin_Structure Nabut FAIRE K562, Chromatin_Structure FAIRE H1hesc, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq Th1wb33676984, Chromatin_Structure DNase-seq K562, Chromatin_Structure DNase-seq Helas3, Chromatin_Structure Ifna4h DNase-seq Helas3, Chromatin_Structure FAIRE K562, Chromatin_Structure Hypoxlac FAIRE Mcf7,
LJ	LS	chr2	167170722	C	T	transition	SCN9A	intronic	CpG_site	NA	5	Motifs PWM MAFK, Motifs PWM Pax-4, Chromatin_Structure DNase-seq Gm04503, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Nhdfneo, Chromatin_Structure DNase-seq Hipe, Chromatin_Structure DNase-seq Hvmf, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq Hffmyc, Chromatin_Structure DNase-seq Ag04449, Chromatin_Structure FAIRE Urotsa, Chromatin_Structure FAIRE UrotsaUt189, Chromatin_Structure Serumfree DNase-seq Aosmc, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Ag09309,
B113	B114	chr3	171864901	G	C	transversion	FNDC3B	intronic	NA	NA	5	Chromatin_Structure DNase-seq K562, Chromatin_Structure Zfp5 DNase-seq K562
B123	B124	chr2	172411647	G	C	transversion	CYBRD1	3utr	NA	NA	5	Motifs PWM Prx1, Chromatin_Structure DNase-seq Hcm, Chromatin_Structure DNase-seq Hmveccly, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq Hmf, Chromatin_Structure DNase-seq Nhdfad, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Hpf, Chromatin_Structure DNase-seq Mcf7
B53	B54	chr5	177563147	C	T	transition	RMND5B	intronic	CpG_site	NA	5	Motifs Footprinting Mcf7 Dax1, Motifs PWM COUPTF, Motifs PWM Dax1, Chromatin_Structure Est100nm1h DNase-seq Mcf7, Chromatin_Structure Estctrl0h DNase-seq Mcf7, Chromatin_Structure DNase-seq Mcf7
B123	B124	chr3	181670965	C	T	transition	LINC01206	non-coding intronic	NA	NA	5	Motifs PWM Zfp740, Chromatin_Structure DNase-seq Nt2d1, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq H7es
LJ	LS	chr2	184856690	T	C	transition		intergenic	NA	NA	5	Motifs PWM MyoD, Motifs Footprinting A549 MyoD, Motifs PWM RARG, Protein_Binding ChIP-seq HeLa-S3 GTF2B, Protein_Binding ChIP-seq HeLa-S3 POI.R3A
B62	B63	chr2	188360514	A	G	transition	TFPI	intronic	NA	NA	5	Motifs PWM Gata5, Chromatin_Structure DNase-seq Hasp

LJ LS	chr2	218660590	C	G	transversion	intergenic	NA	NA	5	Motifs PWM Sp1, Chromatin_Structure Znf41c6 DNase-seq K562, Chromatin_Structure DNase-seq H1hesc, Chromatin_Structure DNase-seq Progfib, Chromatin_Structure Diffa14d DNase-seq H7es, Chromatin_Structure DNase-seq Hah, Chromatin_Structure DNase-seq Fibrobl, Chromatin_Structure DNase-seq Ipsihi7, Chromatin_Structure DNase-seq Hipe, Chromatin_Structure Est10nm30m DNase-seq Ecc1, Chromatin_Structure Diffa5d DNase-seq H7es, Chromatin_Structure DNase-seq Hff, Chromatin_Structure DNase-seq Hcm, Chromatin_Structure DNase-seq K562, Chromatin_Structure Diffa9d DNase-seq H7es, Chromatin_Structure Tam10030 DNase-seq Ishikawa, Chromatin_Structure DNase-seq Fibropag08395, Chromatin_Structure DNase-seq Stellate, Chromatin_Structure Diff4d DNase-seq Lhcnm2, Chromatin_Structure DNase-seq Imr90, Chromatin_Structure Lentimyod DNase-seq Fibroblgm03348, Chromatin_Structure DNase-seq Hepg2, Chromatin_Structure DNase-seq Ipscwr1, Chromatin_Structure DNase-seq Lhcnm2, Chromatin_Structure DNase-seq Fibropag20443, Chromatin_Structure DNase-seq Hnpce, Chromatin_Structure DNase-seq Hsmmemb, Chromatin_Structure DNase-seq Nhlf, Chromatin_Structure Est10nm30m DNase-seq Ishikawa, Chromatin_Structure DNase-seq Ipsihi11, Chromatin_Structure DNase-seq Ips, Chromatin_Structure DNase-seq Hcf, Chromatin_Structure DNase-seq Fibrop, Chromatin_Structure DNase-seq Panisd, Chromatin_Structure DNase-seq Cmk, Chromatin_Structure DNase-seq Fibroblgm03348, Chromatin_Structure Dm002p1h DNase-seq Ecc1, Chromatin_Structure DNase-seq Ag10803, Chromatin_Structure DNase-seq H7es, Chromatin_Structure DNase-seq Fibronag08396. Chromatin_Structure DNase- Chromatin_Structure DNase-seq Hvmf	
B53 B54	chr2	230096243	G	A	transition	PID1	intronic	NA	NA	5	Motifs PWM TCF11 MafG, Protein_Binding ChIP-seq PBDE GATA1
B103 B104	chr2	231640380	T	G	transversion	CAB39	intronic	NA	NA	5	Chromatin_Structure DNase-seq Hgf
B53 B54	chr2	241555733	C	A	transversion	GPR35	intronic	CpG_site	NA	5	Motifs PWM IRF
B93 B94	chr18	508117	A	C	transversion	intergenic		NA	NA	6	Motifs PWM HFH8(FOXF1A), Motifs Footprinting A549 HFH8(FOXF1A), Motifs PWM HNF3beta, Motifs PWM HNF3, Motifs PWM HFH4(FOXJ1), Motifs Footprinting LncapAndro Freac-7, Motifs PWM FOXC1, Motifs Footprinting A549 HFH3(FOXI1), Motifs Footprinting LncapAndro FOXO3A, Motifs PWM Dbx1, Motifs PWM Freac-7, Motifs PWM Cdx, Motifs Footprinting Gm12878 FOXO3A, Motifs Footprinting A549 Cdx, Motifs Footprinting A549 Foxd3, Motifs PWM Foxl1, Motifs Footprinting Helas3Ifna4h Cdx, Motifs PWM FOXP1, Motifs Footprinting Cll Freac-7, Motifs Footprinting A549 Freac-7, Motifs Footprinting A549 HFH4(FOXJ1), Motifs PWM FOXL1, Motifs PWM FOXD3, Motifs PWM Foxd3, Motifs PWM POU3F1, Motifs Footprinting A549 HNF3, Motifs PWM Foxj3, Motifs PWM ONECUT3, Motifs PWM Tlx2, Motifs PWM FOXC2, Motifs PWM FOXO3A, Motifs Footprinting A549 FOXO3A, Motifs Footprinting LncapAndro Cdx, Motifs Footprinting Cll Cdx, Motifs Footprinting Gm12878 Cdx, Motifs PWM FOXJ2, Motifs Footprinting Cll FOXO3A, Motifs Footprinting A549 HNF3beta, Motifs PWM Foxo3
B53 B54	chr12	634274	T	G	transversion	B4GALNT3	intronic	NA	NA	6	

B53 B54	chr7	2384889	C	G	transversion	intergenic		NA	NA	6	Motifs PWM AR, Motifs PWM Ar, Motifs PWM GR
B83 B84	chr19	4421191	C	T	transition	CHAF1A	intronic	NA	NA	6	Motifs PWM LUN-1, Chromatin_Structure FAIRE Helas3
B113 B114	chr18	6348505	T	C	transition	L3MBTL4	intronic	NA	NA	6	Motifs PWM ESRRA, Motifs PWM Hbp1
B93 B94	chr18	10325164	C	T	transition	intergenic		NA	NA	6	Motifs PWM Oct-1
B103 B104	chr9	12145048	G	A	transition	intergenic		NA	NA	6	Motifs PWM Six6, Motifs PWM Sox7, Motifs PWM DMRT3, Motifs PWM Six3
B62 B63	chr20	13918102	A	G	transition	SEL1L2	intronic	NA	NA	6	Motifs PWM FOXG1
LJ LS	chr9	15398550	G	C	transversion	intergenic		NA	NA	6	Motifs PWM Pitx2, Motifs Footprinting CII Pitx2
B83 B84	chr21	16181595	A	G	transition	intergenic		NA	NA	6	Motifs PWM Gfi1b
B83	chr8	16483051	T	C	transition	intergenic		NA	NA	6	Motifs PWM NR2E1
B93 B94	chr2	16519005	G	C	transversion	intergenic		NA	NA	6	Motifs PWM Cart-1
B62 B63	chr3	16887859	C	T	transition	intergenic		NA	NA	6	Motifs PWM Hand1:E47, Motifs PWM SMAD4
B93 B94	chr7	19081637	C	A	transversion	intergenic		NA	NA	6	Motifs PWM HeliosA
B62 B63	chr19	20630678	T	C	transition	intergenic		NA	NA	6	Motifs PWM CIZ
B62 B63	chr21	21885587	G	A	transition	intergenic		NA	NA	6	Motifs PWM ATF4
B62 B63	chr10	24095479	T	C	transition	KIAA1217	intronic	NA	NA	6	Motifs PWM POU4F3, Motifs PWM POU4F2, Motifs PWM POU4F1
B53 B54	chr11	30052740	G	C	transversion	intergenic		NA	NA	6	Motifs PWM Meis1, Motifs PWM MAFB, Motifs PWM TGIF
B53 B54	chr11	31215844	C	G	transversion	intergenic		NA	NA	6	Motifs PWM PTF1-beta, Motifs PWM RARA, Motifs PWM RARB
B123 B124	chr19	31507732	T	G	transversion	intergenic		NA	NA	6	Motifs PWM HoxB5, Motifs PWM Hoxc8, Motifs PWM Pax-4, Motifs PWM HOXC-8, Motifs PWM Dlx5, Motifs PWM Dlx3, Motifs PWM Hoxa6, Motifs PWM Dlx2, Motifs PWM Dlx4, Motifs PWM Hoxb5, Motifs PWM H1vho
B113 B114	chr5	31616208	A	G	transition	intergenic		NA	NA	6	Motifs PWM Tcf3
B62 B63	chr10	31767758	G	A	transition	ZEB1	intronic	NA	NA	6	Motifs PWM TBX15, Motifs PWM FOXB1
B93 B94	chr2	32706159	A	G	transition	BIRC6	intronic	NA	NA	6	Motifs PWM POU3F3, Motifs PWM Mafb, Motifs PWM POU3F2, Motifs PWM POU3F1
B123 B124	chr5	34827544	A	C	transversion	RAI14	intronic	NA	NA	6	Motifs PWM HIC1, Motifs PWM RFX4
B62 B63	chr11	34949538	A	G	transition	PDHX	intronic	NA	NA	6	Motifs PWM POU3F3, Motifs PWM Oct-1, Motifs PWM POU3F1, Motifs PWM POU2F1, Motifs PWM POU2F2
B62	chr11	35864072	A	T	transversion	intergenic		NA	NA	6	Motifs PWM SPIB, Motifs PWM Srf, Motifs PWM SPIC, Motifs PWM SPI1, Motifs PWM SPI-B
B53 B54	chr14	37701455	T	C	transition	MIPOL1	intronic	NA	NA	6	Motifs PWM HNF1A
LJ LS	chr14	39073589	G	C	transversion	intergenic		NA	NA	6	Motifs PWM Mtf1, Motifs PWM Oct-4(POU5F1), Motifs PWM DMRT5
B53 B54	chr1	40982533	A	G	transition	EXO5	3downstream	NA	NA	6	Motifs PWM AIRE
B123 B124	chr2	41332857	C	T	transition	intergenic		NA	NA	6	Motifs PWM COMP1
B62 B63	chr12	41970530	A	G	transition	intergenic		NA	NA	6	Motifs PWM IRX5, Motifs PWM IRX2, Motifs PWM IRX3
B103 B104	chr12	42352274	A	C	transversion	intergenic		NA	NA	6	Motifs PWM Gfi-1
B123	chr14	43121265	G	A	transition	intergenic		CpG_site	NA	6	Motifs PWM FOXC1, Motifs PWM Freac-3, Motifs PWM Sox7
B53 B54	chr1	43588564	C	T	transition	intergenic		NA	NA	6	Motifs PWM FOXP1, Motifs PWM Six6
B84	chr13	45830708	C	T	transition	GTF2F2	intronic	NA	NA	6	Motifs PWM Oct-1
B93 B94	chr3	46151596	A	G	transition	intergenic		NA	NA	6	Motifs PWM Pax-4, Chromatin_Structure FAIRE Gm19239
B83	chr8	50040053	G	A	transition	intergenic		NA	NA	6	Motifs PWM HeliosA
B113	chr20	52177664	A	G	transition	RP4_724E16.2 non-coding	intronic	NA	NA	6	Motifs Footprinting A549 EWSR1-FLII, Motifs PWM EWSR1-FLII
B83 B84	chr3	53325297	C	T	transition	DCP1A	intronic	NA	NA	6	Motifs PWM POU3F3
B83 B84	chr5	53365411	A	C	transversion	ARL15	intronic	NA	NA	6	Motifs PWM Srf, Motifs PWM FOXP1, Motifs PWM Zfp105, Motifs PWM Mtf1, Motifs PWM AIRE
B113 B114	chr19	53997821	C	T	transition	ZNF813	3downstream	CpG_site	NA	6	Motifs PWM ZNF524
LJ LS	chr7	54008241	C	T	transition	intergenic		CpG_site	NA	6	Motifs PWM Obox1
B62 B63	chr14	54397605	T	C	transition	intergenic		NA	NA	6	Motifs PWM Dbx1, Motifs PWM C/EBPgammad, Motifs PWM SOX5
B93 B94	chr17	54433363	A	G	transition	ANKFN1	intronic	NA	NA	6	Motifs PWM Oct-1
B93 B94	chr17	54433372	T	C	transition	ANKFN1	intronic	NA	NA	6	Motifs PWM Oct-1

B123 B124	chr4	55065295	T	A	transversion	intergenic		NA	NA	6	Motifs PWM Barhl2, Motifs PWM Nkx5-2
B93 B94	chrX	55365659	C	A	transversion	intergenic		NA	NA	6	Motifs PWM HNF3alpha, Chromatin_Structure FAIRE Astrocy
B93 B94	chrX	55595693	C	A	transversion	intergenic		NA	NA	6	Motifs PWM MEF-2, Chromatin_Structure FAIRE A549
B103 B104	chr15	56398367	T	C	transition	RFX7	intronic	NA	NA	6	Motifs PWM Bcl6b, Chromatin_Structure FAIRE A549
B123 B124	chr6	56978734	T	G	transversion	ZNF451	intronic	NA	NA	6	Motifs PWM Tcf7, Motifs PWM Tbp
B53 B54	chr3	57476826	A	C	transversion	DNAH12	intronic	NA	NA	6	Motifs PWM FAC1, Motifs PWM Foxk1, Motifs PWM FOXC1, Motifs PWM Mtf1, Motifs PWM FOXO3, Motifs PWM Srf, Motifs PWM BCL6, Motifs PWM FOXO1, Motifs PWM Zfn105, Motifs PWM FOXO1
B53 B54	chr8	57645596	T	A	transversion	intergenic		NA	NA	6	Motifs PWM MEF-2
B93 B94	chr17	58787185	G	A	transition	BCAS3	intronic	NA	NA	6	Motifs Footprinting Mcf7 SRF, Motifs Footprinting Mcf7Hypoxlac SRF, Motifs PWM SRF
B123 B124	chr17	60527119	A	G	transition	METTL2A	3utr	NA	NA	6	Motifs Footprinting GM12892 , Motifs Footprinting HelaS3
B103 B104	chr16	63843879	C	T	transition	intergenic		NA	NA	6	Motifs PWM NR3C1
B93 B94	chr14	64047343	A	G	transition	intergenic		NA	NA	6	Motifs PWM Dbx2, Motifs PWM Vsx1, Motifs PWM Rhox6, Motifs PWM Pou4f3, Motifs PWM Hoxd8, Motifs PWM Sox1, Motifs PWM Isl2, Motifs PWM Pon3f1, Chromatin_Structure FAIRE Htr8
B113 B114	chr13	65852160	C	A	transversion	intergenic		NA	NA	6	Motifs PWM HOXA10, Motifs PWM HOXC11, Motifs PWM Hoxa9, Motifs PWM Cdx2, Motifs PWM Hoxd10, Motifs PWM Cdx1, Motifs PWM Cdx- 2, Motifs PWM hoxa9, Motifs PWM HOXA11, Motifs PWM Hoxb9, Motifs PWM Sox7, Motifs PWM HOXD11
B103 B104	chr8	66834673	A	G	transition	intergenic		NA	NA	6	Motifs PWM AREB6, Motifs PWM Pitx2, Motifs PWM TCF4, Motifs PWM MEIS2
B114	chr9	66861389	T	C	transition	intergenic		NA	NA	6	Motifs Footprinting Helas3Ifna4h HSF2, Motifs Footprinting Gm12878 HSF2, Motifs Footprinting Hpd6e6e7 HSF2, Motifs PWM HSF1, Motifs PWM HSF2
B114	chr12	67837996	G	A	transition	intergenic				6	Motifs PWM RAR
B53 B54	chr2	68543768	A	T	transversion	CNRIP1	intronic	CpG_site	NA	6	Motifs PWM Six6, Motifs PWM Tbp, Motifs PWM Dobox4
B93 B94	chr3	70268930	A	C	transversion	intergenic		NA	NA	6	Motifs PWM NR1H2::RXRA
B93 B94	chr15	70747621	C	G	transversion	intergenic		NA	NA	6	Motifs PWM Pitx2, Motifs Footprinting Mcf7 Pitx2
B93 B94	chr18	73917867	G	A	transition	intergenic		NA	NA	6	Motifs PWM CREB3L1, Motifs PWM CREB3L2, Chromatin_Structure FAIRE Medullo
B83 B84	chr15	75719027	T	C	transition	SIN3A		intronic	NA	NA	6
B62 B63	chr3	76252246	C	T	transition	intergenic		CpG_site	NA	6	Motifs PWM TCFAP2A, Motifs PWM TFAP2B, Motifs PWM Tcfap2e, Motifs PWM TFAP2C, Motifs PWM TFAP2A, Motifs PWM Tcfap2b
LJ LS	chr16	76707882	C	T	transition	intergenic		NA	NA	6	Motifs PWM Lmx1a, Motifs PWM Lhx1, Motifs PWM HMGIY, Motifs PWM Arid3a, Motifs PWM Lmx1b, Motifs PWM Lhx5, Motifs PWM Hoxd13
B53 B54	chr3	76877788	A	G	transition	intergenic	intronic	NA	NA	6	Motifs PWM Tlx2
B103 B104	chr15	78321335	T	G	transversion	TBC1D2B		NA	NA	6	Motifs PWM Sox18, Motifs PWM Sry, Motifs PWM EMX2, Motifs PWM Cdx, Motifs PWM Nkx2-5, Motifs PWM Hmx2, Motifs PWM Sox5, Motifs PWM EN1, Motifs PWM Sox21, Motifs PWM Hmx1, Motifs PWM Nkx5- 2, Motifs PWM Sox15, Motifs PWM HMX1, Motifs PWM MSX1, Motifs PWM Hmx3, Motifs PWM Sox12, Motifs PWM EMX1 Motifs PWM AP-2alphaA
B103 B104	chr12	79083015	G	A	transition	intergenic		NA	NA	6	Motifs PWM AP-2alphaA

B93 B94	chr2	79194279	T	A	transversion	intergenic		NA	NA	6	Motifs PWM NF-AT, Chromatin_Structure FAIRE Gm12891
LJ LS	chr1	79265388	C	T	transition	intergenic		NA	NA	6	Motifs PWM FOXJ3, Motifs PWM Zfp187, Motifs PWM FOXJ2
B93 B94	chr13	79350867	A	G	transition	intergenic		NA	NA	6	Motifs PWM VDR
B113 B114	chr12	80731159	G	A	transition	OTOG	intronic	CpG_site	NA	6	Motifs PWM MECP2
B62 B63	chr1	81565925	A	G	transition	intergenic		NA	NA	6	Motifs PWM BCL6
B53 B54	chr4	81742124	C	T	transition	C4orf22	intronic	NA	NA	6	Motifs PWM SPDEF
B93 B94	chr7	85163682	G	A	transition	intergenic		NA	NA	6	Motifs PWM HNF3, Motifs PWM MEF2A, Motifs PWM MEF-2, Motifs PWM FOXD3, Motifs PWM RSRFC4, Motifs PWM Freac-4, Motifs PWM Foxi3, Motifs PWM Hoxd8, Motifs PWM Foxa2
B62 B63	chr13	85569294	C	T	transition	intergenic		NA	NA	6	Motifs PWM AIRE, Chromatin_Structure FAIRE Panislets, Chromatin_Structure FAIRE Gliobla
B62	chr13	87882896	A	C	transversion	intergenic		NA	NA	6	Motifs PWM Dbx1, Motifs PWM FOXJ3, Motifs PWM FOXP1, Motifs PWM MEF-2, Motifs PWM Tlx2, Motifs PWM FOXJ2, Motifs PWM Sox8
B103 B104	chr3	89209126	T	G	transversion	EPHA3	intronic	NA	NA	6	Motifs PWM Srf, Motifs PWM FOXP1
B113 B114	chr4	89304876	G	A	transition	HERC6	intronic	NA	NA	6	Motifs PWM Zbtb3
B83	chr8	94260080	G	T	transversion	intergenic		NA	NA	6	Motifs PWM FOXJ3, Motifs PWM FOXJ2
B84	chr9	94497890	T	C	transition	ROR2	intronic	NA	NA	6	Motifs PWM Srf, Motifs PWM FOXP1, Motifs PWM Zfp105, Motifs PWM Mtf1
B62 B63	chr15	95452112	T	A	transversion	intergenic		NA	NA	6	Motifs PWM Tcf3, Motifs PWM Srf, Motifs PWM Tcfap2e, Motifs PWM Tcf7, Motifs PWM Zfp105
B103 B104	chr3	95967254	T	G	transversion	intergenic		NA	NA	6	Motifs PWM AR, Motifs PWM Gfi1b
B113	chr15	96155017	C	A	transversion	intergenic		NA	NA	6	Motifs PWM Barhl-1, Motifs PWM Nkx5-2
B62 B63	chrX	96306108	C	T	transition	DIAPH2	intronic	NA	NA	6	Motifs PWM ZFP652, Motifs PWM Gfi-1
B123 B124	chr5	97928714	T	G	transversion	intergenic		NA	NA	6	Motifs PWM Oct-1, Motifs PWM HMGIY, Motifs PWM HSF1, Motifs PWM Sox7, Motifs PWM NFAT5, Motifs PWM Nanog, Motifs PWM HSF2
B113 B114	chr1	101012039	G	T	transversion	intergenic		CpG_site	NA	6	Motifs PWM RBP-Jkappa
B53 B54	chr11	101016266	T	C	transition	LOC10105452 ^c	non-coding	NA	NA	6	Motifs PWM Hoxa5, Motifs PWM Lhx4, Motifs PWM Evx1, Motifs PWM Meox1, Motifs PWM Hoxa6, Motifs PWM Hoxa2, Motifs PWM Hoxd3, Motifs PWM Dlx2, Motifs PWM Hoxb3, Motifs PWM Gch2, Motifs PWM Infl Motifs PWM HOXD3
B62 B63	chr4	102822941	A	T	transversion	BANK1	intronic	NA	NA	6	Motifs PWM DMRT5
B113 B114	chr7	108858722	C	A	transversion	intergenic		NA	NA	6	Motifs PWM FOXC1, Motifs PWM SPIC
B53 B54	chr7	113264399	A	G	transition	intergenic		NA	NA	6	Motifs PWM Srf, Motifs PWM FOXP1, Motifs PWM Zfp105, Motifs PWM Mtf1
LJ LS	chr4	115909880	T	A	transversion	NDST4	intronic	NA	NA	6	Motifs PWM NF-E2, Motifs PWM AP-1
LJ LS	chr4	115909881	C	A	transversion	NDST4	intronic	NA	NA	6	Motifs PWM NF-E2, Motifs PWM AP-1
B103 B104	chr8	118537055	G	A	transition	MED30	intronic	CpG_site	NA	6	Motifs PWM Tcf3, Motifs PWM IRF8, Motifs PWM IRF2, Motifs PWM HMGA2, Motifs PWM IRF3, Motifs PWM ISGF-3, Motifs PWM IRF7, Motifs PWM DMRT7, Motifs PWM STAT1
B62 B63	chr2	122634222	A	G	transition	intergenic		NA	NA	6	Motifs PWM Brachyury
LS	chr12	123034737	A	T	transversion	KNTC1	intronic	NA	NA	6	Motifs PWM Pou2f3, Motifs PWM POU5F1P1, Motifs PWM Pou2f2, Motifs PWM POU3F3, Motifs PWM Oct-1, Motifs PWM Octamer, Motifs PWM POU2F3, Motifs PWM POU3F1, Motifs PWM OCT-x, Motifs PWM POU3F4, Motifs PWM POU2F1, Motifs PWM POU1F1, Motifs PWM POU1F2 Motifs PWM POU1F2
B62 B63	chrX	123121415	C	T	transition	STAG2	intronic	NA	NA	6	Motifs PWM Elf3, Motifs PWM Gata6
B62 B63	chr10	127420577	G	A	transition	EDRF1	intronic	CpG_site	NA	6	Motifs PWM Mybl1, Motifs PWM Myb
B113	chr9	127918364	G	A	transition	PPP6C	intronic	NA	NA	6	Motifs PWM Oct-1, Motifs PWM C/EBP, Motifs PWM Mafb, Motifs PWM Oct-4(POU5F1)
LJ LS	chr4	129175380	T	G	transversion	intergenic		NA	NA	6	Motifs PWM STAT1:STAT1
B93 B94	chr11	131293441	T	C	transition	NTM	intronic	NA	NA	6	Motifs PWM FOXP3, Chromatin_Structure FAIRE Nhbe

B123 B124	chr3	139887457	G	T	transversion	CLSTN2	intronic	NA	NA	6	Motifs PWM BARX1
B113 B114	chr6	145886359	G	C	transversion	intergenic		NA	NA	6	Motifs PWM FXR/RXR-alpha
LJ LS	chr2	146609689	G	A	transition	intergenic		NA	NA	6	Motifs PWM HNF4A
B93 B94	chr4	154787026	A	G	transition	intergenic		NA	NA	6	Motifs PWM HIC2
LJ LS	chr3	155766601	G	A	transition	intergenic		NA	NA	6	Motifs PWM MTF1, Motifs PWM C/EBP
B93 B94	chr6	163212623	C	A	transversion	PACRG	intronic	CpG_site	NA	6	Motifs PWM AR
B83 B84	chr3	164228118	A	C	transversion	intergenic		NA	NA	6	Motifs PWM RUSH-1alpha, Motifs PWM Hltf
B93 B94	chr6	165600454	T	C	transition	intergenic		NA	NA	6	Motifs PWM TP53, Motifs PWM p53
B62 B63	chr5	166938981	A	G	transition	TENM2	intronic	NA	NA	6	Motifs PWM GR
B123 B124	chr3	168345596	T	G	transversion	EGFEM1P	non-coding	intronic	NA	NA	Motifs PWM E2F2, Motifs PWM Foxa2
LS	chr2	174321128	C	G	transversion	intergenic		NA	NA	6	Motifs PWM EWSR1-FLI1
B113 B114	chr3	174392522	T	C	transition	intergenic		NA	NA	6	Motifs PWM PU.1
B53 B54	chr1	182887203	C	A	transversion	SHCBP1L	intronic	NA	NA	6	Motifs PWM Tbp
B93 B94	chr2	182975989	A	T	transversion	PPP1R1C	intronic	NA	NA	6	Motifs PWM HFH8(FOXF1A), Motifs PWM Freac-7, Motifs PWM FOXO6, Motifs PWM FOXO3, Motifs PWM Zscan4, Motifs PWM FOXO4, Motifs PWM FOXO1
B103 B104	chr2	185753314	C	A	transversion	ZNF804A	intronic	NA	NA	6	Motifs PWM Zbtb3
LJ LS	chr2	186167711	T	G	transversion	intergenic		NA	NA	6	Motifs PWM HNF3alpha, Motifs PWM FOXJ2, Motifs PWM HFH3(FOXI1)
LJ LS	chr2	186167712	A	T	transversion	intergenic		NA	NA	6	Motifs PWM HNF3alpha, Motifs PWM FOXJ2, Motifs PWM HFH3(FOXI1)
B93 B94	chr1	186835728	A	G	transition	PLA2G4A	intronic	NA	NA	6	Motifs PWM FOXD3, Motifs PWM Irx2, Motifs PWM Irx6, Motifs PWM Irx-3, Motifs PWM BARHL2, Motifs PWM Tlx2, Motifs PWM Irx5, Motifs PWM Irx3, Motifs PWM RSRFC4, Motifs PWM Irx4, Motifs PWM AFP1
B83 B84	chr3	189327482	C	T	transition	intergenic		NA	NA	6	Motifs PWM Oct-1, Chromatin_Structure FAIRE Gliobla
B83 B84	chr1	218352394	A	G	transition	intergenic		NA	NA	6	Motifs PWM ISGF-3
B83 B84	chr2	230145171	G	A	transition	intergenic		NA	NA	6	Motifs PWM TFE, Motifs PWM Sox17, Motifs PWM Sox8
B53 B54	chr2	230274367	C	T	transition	DNER	intronic	CpG_site	NA	6	Motifs PWM SP1, Motifs PWM Sp1, Motifs PWM Ascl2
LJ LS	chr2	238513142	C	T	transition	intergenic		NA	NA	6	Motifs Footprinting Fibrop E2A, Motifs Footprinting Hpde6e6e7 E2A, Motifs Footprinting Cll E2A, Motifs Footprinting Medullo E2A, Motifs Footprinting Hepg2 E2A, Motifs Footprinting Myometr E2A, Motifs Footprinting Gliobla E2A, Motifs Footprinting Gm12891 E2A, Motifs Footprinting Huh75 E2A, Motifs Footprinting LnCapAndro E2A, Motifs Footprinting Gm12878 E2A, Motifs Footprinting Gm12892 E2A, Motifs Footprinting Gm19238 E2A, Motifs PWM E2A, Motifs Footprinting Htr8 E2A, Motifs Footprinting 8988t E2A,
B62 B63	chr2	242283282	A	G	transition	SEPT2	nonsyn p.N271S	NA	NA	6	Motifs PWM ESR2
B103 B104	chr17	2421858	C	T	transition	intergenic		CpG_site	NA	7	No data
LJ	chr19	3354318	T	C	transition	intergenic		NA	NA	7	No data
B123 B124	chr3	3361010	G	C	transversion	intergenic		NA	NA	7	No data
LJ LS	chr8	3381974	C	A	transversion	CSMD1	intronic	NA	NA	7	No data
B103 B104	chr8	5214640	G	A	transition	intergenic		NA	NA	7	No data
B93 B94	chr4	5962283	A	G	transition	intergenic		NA	NA	7	No data
B123 B124	chr3	7391648	G	A	transition	GRM7	intronic	CpG_site	NA	7	No data
B84	chr7	8681356	T	C	transition	NXPH1	intronic	NA	NA	7	No data
B123 B124	chr9	11027822	C	T	transition	intergenic		NA	NA	7	No data
B83 B84	chr5	11250985	G	A	transition	CTNND2	intronic	CpG_site	NA	7	No data
B103 B104	chr9	11417777	A	T	transversion	intergenic		NA	NA	7	No data
B103 B104	chr12	11507171	G	A	transition	PRB1	intronic	NA	NA	7	No data
B62 B63	chr17	12179178	A	G	transition	intergenic		NA	NA	7	No data
B53 B54	chr4	12439530	C	A	transversion	intergenic		NA	NA	7	No data

LJ LS	chr17	13733612	C	T	transition	intergenic		NA	NA	7	No data
B83 B84	chr17	13734863	G	A	transition	intergenic		NA	NA	7	No data
B53 B54	chr1	14607191	C	T	transition	intergenic		NA	NA	7	No data
B123 B124	chr9	15437524	C	T	transition	SNAPC3	intronic	NA	NA	7	No data
B123 B124	chr10	15589618	C	T	transition	ITGA8	intronic	NA	NA	7	No data
B123 B124	chr4	15798530	G	T	transversion	CD38	intronic	NA	NA	7	No data
B83 B84	chr7	16224899	A	G	transition	ISPD	intronic	NA	NA	7	No data
B62 B63	chr2	17073960	C	T	transition	intergenic		NA	NA	7	No data
B62 B63	chr12	17246962	A	G	transition	intergenic		NA	NA	7	No data
B53 B54	chr6	17265790	C	T	transition	intergenic		NA	NA	7	No data
LJ LS	chrY	17459241	A	G	transition	intergenic		NA	NA	7	No data
B83 B84	chr21	17868492	A	G	transition	LINC00478	non-coding intronic	NA	NA	7	No data
B93 B94	chr7	17914832	C	A	transversion	SNX13	intronic	NA	NA	7	No data
B123 B124	chr12	19654480	A	T	transversion	AEBP2	intronic	NA	NA	7	No data
B123 B124	chr10	20845041	C	T	transition	intergenic		CpG_site	NA	7	No data
B62 B63	chr13	20923983	G	A	transition	intergenic		CpG_site	NA	7	No data
B93 B94	chr19	21062165	C	T	transition	intergenic		NA	NA	7	No data
B123 B124	chr3	21088283	C	T	transition	intergenic		NA	NA	7	No data
B83 B84	chr10	21149815	G	C	transversion	NEBL	intronic	NA	NA	7	No data
B93 B94	chr10	21979949	C	G	transversion	MLLT10	intronic	NA	NA	7	No data
B93 B94	chr8	22898794	T	C	transition	TNFRSF10B	intronic	NA	NA	7	No data
B113 B114	chr16	24377252	A	G	transition	intergenic		NA	NA	7	No data
B53 B54	chr7	24594488	T	C	transition	intergenic		NA	NA	7	No data
B123 B124	chr9	24903909	G	A	transition	intergenic		NA	NA	7	No data
B62 B63	chr3	24933640	C	T	transition	intergenic		NA	NA	7	No data
B93 B94	chr9	25091094	G	C	transversion	intergenic		CpG_site	NA	7	No data
B83 B84	chr21	25258273	G	C	transversion	intergenic		CpG_site	NA	7	No data
B62 B63	chr9	25628943	T	C	transition	intergenic		NA	NA	7	No data
B113	chr12	26742696	A	G	transition	ITPR2	intronic	NA	NA	7	No data
B62 B63	chr12	28411429	G	T	transversion	CCDC91	intronic	NA	NA	7	No data
B83 B84	chr14	30014118	A	G	transition	MIR548AI	non-coding intronic	NA	NA	7	No data
B83 B84	chr3	31020364	A	G	transition	intergenic		NA	NA	7	No data
B83 B84	chr17	31223286	C	T	transition	intergenic		NA	NA	7	No data
B113 B114	chr18	32683856	G	C	transversion	MAPRE2	intronic	NA	NA	7	No data
B123 B124	chr4	33398722	C	T	transition	intergenic		CpG_site	NA	7	No data
B113 B114	chr6	34401949	G	A	transition	intergenic		CpG_site	NA	7	No data
B53 B54	chr9	34617204	C	T	transition	DCTN3	intronic	NA	NA	7	No data
B103 B104	chr20	34752676	C	T	transition	EPB41L1	intronic	CpG_site	NA	7	No data
B123 B124	chr16	35013471	T	A	transversion	intergenic		NA	NA	7	No data
B53 B54	chr13	36215362	A	C	transversion	NBEA	intronic	NA	NA	7	No data
B84	chr4	36920104	A	C	transversion	intergenic		NA	NA	7	No data
B103 B104	chr20	37200837	G	T	transversion	RALGAPB	intronic	NA	NA	7	No data
LJ LS	chr18	38732423	A	G	transition	intergenic		NA	NA	7	No data
B93 B94	chr15	39986654	C	G	transversion	FSIP1	intronic	NA	NA	7	No data
B123 B124	chr21	42082429	G	A	transition	DSCAM	intronic	CpG_site	NA	7	No data
B103 B104	chrX	42471130	T	G	transversion	intergenic		NA	NA	7	No data
B103 B104	chr13	43170170	G	A	transition	TNFSF11	intronic	CpG_site	NA	7	No data
B53 B54	chr8	43297461	T	G	transversion	intergenic		NA	NA	7	No data
B62 B63	chr2	43609005	G	C	transversion	THADA	intronic	NA	NA	7	No data
B62 B63	chr4	46434769	G	A	transition	intergenic		NA	NA	7	No data

B123	B124	chr14	46455725	T	G	transversion	intergenic		NA	NA	7	No data
B103	B104	chr8	47649158	A	G	transition	intergenic		NA	NA	7	No data
LJ LS	chr16	47797029	T	C	transition	intergenic		NA	NA	7	No data	
B83	B84	chr15	48506290	T	C	transition	SLC12A1	intronic	NA	NA	7	No data
LJ LS	chr10	53069364	T	C	transition	PRKG1	intronic	NA	NA	7	No data	
B62	B63	chr18	53310663	A	G	transition	intergenic		NA	NA	7	No data
B123	B124	chr20	55421177	T	G	transversion	intergenic		NA	NA	7	No data
B53	B54	chr8	56129326	G	A	transition	XKR4	intronic	NA	NA	7	No data
B123	B124	chr6	56419279	C	T	transition	DST	intronic	NA	NA	7	No data
B53	B54	chr11	57175782	C	T	transition	SLC43A3	intronic	NA	NA	7	No data
B62	B63	chr2	57552402	T	A	transversion	intergenic		NA	NA	7	No data
B62	B63	chrX	58380768	A	C	transversion	intergenic		NA	NA	7	No data
B103	B104	chr12	58842837	C	G	transversion	intergenic		NA	NA	7	No data
B123	B124	chr3	60760326	C	G	transversion	FHIT	intronic	NA	NA	7	No data
B62	B63	chr8	62392971	G	A	transition	CLVS1	intronic	CpG_site	NA	7	No data
B103	B104	chr5	63649074	T	A	transversion	RNF180	intronic	NA	NA	7	No data
B123	chr1	63702337	A	C	transversion	LINC00466	non-coding	intronic	NA	NA	7	No data
B93	B94	chr1	64191122	G	A	transition	intergenic		NA	NA	7	No data
B103	B104	chr18	64205104	G	A	transition	CDH19	intronic	NA	NA	7	No data
B62	B63	chr7	64318208	C	T	transition	intergenic		NA	NA	7	No data
B62	B63	chr14	64473123	C	T	transition	SYNE2	intronic	CpG_site	NA	7	No data
B83	B84	chr13	64665172	G	T	transversion	intergenic		NA	NA	7	No data
B62	B63	chr6	64923693	C	T	transition	EYS	intronic	CpG_site	NA	7	No data
LJ LS	chr10	64995156	A	G	transition	JMJD1C	intronic	NA	NA	7	No data	
B93	B94	chr4	66240782	A	G	transition	EPHA5	intronic	NA	NA	7	No data
B103	B104	chr17	66580685	C	T	transition	FAM20A	intronic	CpG_site	NA	7	No data
B114	chr9	66861395	T	C	transition	intergenic		NA	NA	7	No data	
B62	B63	chr2	68293158	A	G	transition	intergenic		NA	NA	7	No data
LJ LS	chr4	70243771	G	C	transversion	intergenic		NA	NA	7	No data	
B93	B94	chr18	72619462	C	T	transition	ZNF407	intronic	NA	NA	7	No data
B103	B104	chr13	72813354	G	T	transversion	intergenic		NA	NA	7	No data
B93	B94	chr10	72842217	A	G	transition	intergenic		NA	NA	7	No data
LJ LS	chr10	73143664	G	A	transition	intergenic		NA	NA	7	No data	
LJ LS	chr13	73798380	G	A	transition	intergenic		CpG_site	NA	7	No data	
B103	B104	chr13	74635919	C	T	transition	KLF12	intronic	CpG_site	NA	7	No data
B83	B84	chr10	76753997	G	A	transition	KAT6B	intronic	NA	NA	7	No data
B123	B124	chr13	76971899	G	C	transversion	intergenic		NA	NA	7	No data
B93	B94	chr9	77343121	T	A	transversion	TRPM6	intronic	NA	NA	7	No data
B103	B104	chr14	78571385	T	C	transition	intergenic		NA	NA	7	No data
B123	B124	chr5	78999514	T	C	transition	CMYA5	intronic	NA	NA	7	No data
B93	B94	chr16	81098864	G	A	transition	C16orf46	intronic	NA	NA	7	No data
B103	B104	chrX	81626563	A	G	transition	intergenic		NA	NA	7	No data
B103	B104	chr2	82569412	T	C	transition	intergenic		NA	NA	7	No data
B123	B124	chr9	82729515	A	G	transition	intergenic		NA	NA	7	No data
B83	B84	chr11	87164141	C	T	transition	intergenic		CpG_site	NA	7	No data
B123	B124	chr10	87386677	A	C	transversion	GRID1	intronic	NA	NA	7	No data
B93	B94	chr3	88598299	A	G	transition	intergenic		NA	NA	7	No data
B103	B104	chr2	88639215	C	T	transition	intergenic		CpG_site	NA	7	No data
B93	B94	chr9	89456759	C	T	transition	intergenic		CpG_site	NA	7	No data
B113	B114	chr16	89961140	C	A	transversion	TCF25	intronic	CpG_site	NA	7	No data

B93 B94	chr11	90849241	A	G	transition	intergenic		NA	NA	7	No data
B62 B63	chr5	92461001	A	C	transversion	intergenic		NA	NA	7	No data
B103 B104	chr11	94241750	C	A	transversion	intergenic		NA	NA	7	No data
B62 B63	chr10	95127010	C	T	transition	MYOF	intronic	CpG_site	NA	7	No data
LJ LS	chr11	97265884	G	C	transversion	intergenic		NA	NA	7	No data
B123 B124	chr9	99136093	G	C	transversion	SLC35D2	intronic	CpG_site	NA	7	No data
B62 B63	chr3	99546146	G	A	transition	CMSS1	intronic	CpG_site	NA	7	No data
B123 B124	chr8	100386318	A	G	transition	VPS13B	intronic	NA	NA	7	No data
B93 B94	chr4	103002276	C	T	transition	intergenic		CpG_site	NA	7	No data
B113 B114	chr12	103506236	C	T	transition	intergenic		NA	NA	7	No data
B93 B94	chr7	103763484	C	T	transition	intergenic		NA	NA	7	No data
B62 B63	chr11	104908704	C	T	transition	intergenic		NA	NA	7	No data
B83 B84	chr1	105309724	G	A	transition	intergenic		CpG_site	NA	7	No data
LJ LS	chr9	105381829	C	T	transition	LINC00587	non-coding intronic	CpG_site	NA	7	No data
B103 B104	chr8	106280832	T	G	transversion	intergenic		NA	NA	7	No data
B62 B63	chr4	106901329	G	A	transition	intergenic		CpG_site	NA	7	No data
B62 B63	chrX	108826763	C	T	transition	intergenic		CpG_site	NA	7	No data
B113 B114	chr7	108858732	G	T	transversion	intergenic		NA	NA	7	No data
B123 B124	chr3	111154558	G	T	transversion	intergenic		NA	NA	7	No data
B62 B63	chrX	111417043	G	A	transition	ZCCHC16	intronic	NA	NA	7	No data
B53 B54	chr8	111879636	T	A	transversion	intergenic		NA	NA	7	No data
B62 B63	chr5	113042944	C	T	transition	intergenic		CpG_site	NA	7	No data
B53 B54	chr4	114760417	T	A	transversion	intergenic		NA	NA	7	No data
B93 B94	chr11	116421503	A	T	transversion	intergenic		NA	NA	7	No data
LJ LS	chr10	118484331	C	T	transition	HSPA12A	intronic	NA	NA	7	No data
LJ LS	chr1	118539438	T	C	transition	SPAG17	intronic	NA	NA	7	No data
B113 B114	chr5	118782822	G	A	transition	intergenic		CpG_site	NA	7	No data
B83 B84	chr10	118876496	A	G	transition	KIAA1598	intronic	NA	NA	7	No data
B83	chr4	119231662	C	T	transition	PRSS12	intronic	CpG_site	NA	7	No data
B53 B54	chr12	120569035	C	G	transversion	GCN1L1	nonsyn p.S2506T	NA	NA	7	No data
B53 B54	chr6	123917045	A	G	transition	TRDN	intronic	NA	NA	7	No data
B53 B54	chr2	124712603	A	C	transversion	intergenic		NA	NA	7	No data
B123 B124	chr7	125495096	C	T	transition	intergenic		NA	NA	7	No data
B103 B104	chr10	128776253	G	T	transversion	DOCK1	intronic (splice_site)	NA	NA	7	No data
B83 B84	chr4	132378202	C	T	transition	intergenic		NA	NA	7	No data
B93 B94	chr7	132653469	G	A	transition	CHCHD3	intronic	CpG_site	NA	7	No data
B53 B54	chr6	133263812	G	C	transversion	intergenic		NA	NA	7	No data
B83 B84	chr2	137786478	C	T	transition	THSD7B	intronic	CpG_site	NA	7	No data
B93 B94	chr2	138486744	A	G	transition	intergenic		NA	NA	7	No data
B83 B84	chr8	138597674	G	C	transversion	intergenic		NA	NA	7	No data
B53 B54	chr6	140126330	A	G	transition	LOC100132735	non-coding	NA	NA	7	No data
B103 B104	chr3	140390293	C	T	transition	intronic		NA	NA	7	No data
B113 B114	chr6	142436831	C	G	transversion	intergenic		NA	NA	7	No data
B123 B124	chrX	144020346	G	A	transition	intergenic		NA	NA	7	No data
B93 B94	chr6	146456876	G	A	transition	GRM1	intronic	NA	NA	7	No data
B62 B63	chr2	149183277	G	A	transition	MBD5	intronic	CpG_site	NA	7	No data
B84	chr6	149959476	T	A	transversion	KATNA1	intronic	NA	NA	7	No data
B62 B63	chrX	150711814	G	T	transversion	intergenic		NA	NA	7	No data
B103 B104	chr7	152835331	A	G	transition	intergenic		NA	NA	7	No data
B104	chr2	153664053	G	A	transition	intergenic		CpG_site	NA	7	No data

B93 B94	chr2	156799239	A	G	transition	intergenic		NA	NA	7	No data
B103 B104	chr4	162128420	C	T	transition	intergenic		NA	NA	7	No data
B62 B63	chr6	163579077	G	A	transition	PACRG	intronic	NA	NA	7	No data
B93 B94	chr2	164338918	G	A	transition	intergenic		NA	NA	7	No data
B84	chr2	167878685	A	G	transition	XIRP2	intronic	NA	NA	7	No data
B83 B84	chr2	172987977	C	T	transition	intergenic		NA	NA	7	No data
B103 B104	chr3	178375930	C	T	transition	KCNMB2	intronic	NA	NA	7	No data
B53 B54	chr2	179436794	C	T	transition	TTN	nonsyn p.V23048I	NA	NA	7	No data
B53 B54	chr1	191364591	T	G	transversion	intergenic		NA	NA	7	No data
B113 B114	chr2	207501019	T	C	transition	intergenic		NA	NA	7	No data
B93 B94	chr2	212168526	A	G	transition	intergenic		NA	NA	7	No data
B113 B114	chr2	220870844	T	C	transition	intergenic		NA	NA	7	No data
B93 B94	chr1	224422130	T	C	transition	NVL	intronic	NA	NA	7	No data
B62 B63	chr1	231116417	A	G	transition	TTC13	5upstream	NA	NA	7	No data
B93 B94	chr2	241957475	G	A	transition	SNED1	intronic	CpG site	NA	7	No data

* Notes:

1a -- eQTL + TF binding + matched TF motif + matched DNase Footprint + DNase peak

1b -- eQTL + TF binding + any motif + DNase Footprint + DNase peak

1c -- eQTL + TF binding + matched TF motif + DNase peak

1d -- eQTL + TF binding + any motif + DNase peak

1e -- eQTL + TF binding + matched TF motif

1f -- eQTL + TF binding / DNase peak

2a -- TF binding + matched TF motif + matched DNase Footprint + DNase peak

2b -- TF binding + any motif + DNase Footprint + DNase peak

2c -- TF binding + matched TF motif + DNase peak

3a -- TF binding + any motif + DNase peak

3b -- TF binding + matched TF motif

4 -- TF binding + DNase peak

5 -- TF binding or DNase peak

6 -- other